

New Paradigm for Charge Variant Characterization Using cIEF-MS and 'Discovery' Assisted Fractionation

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CASSS Mass Spec, 2025

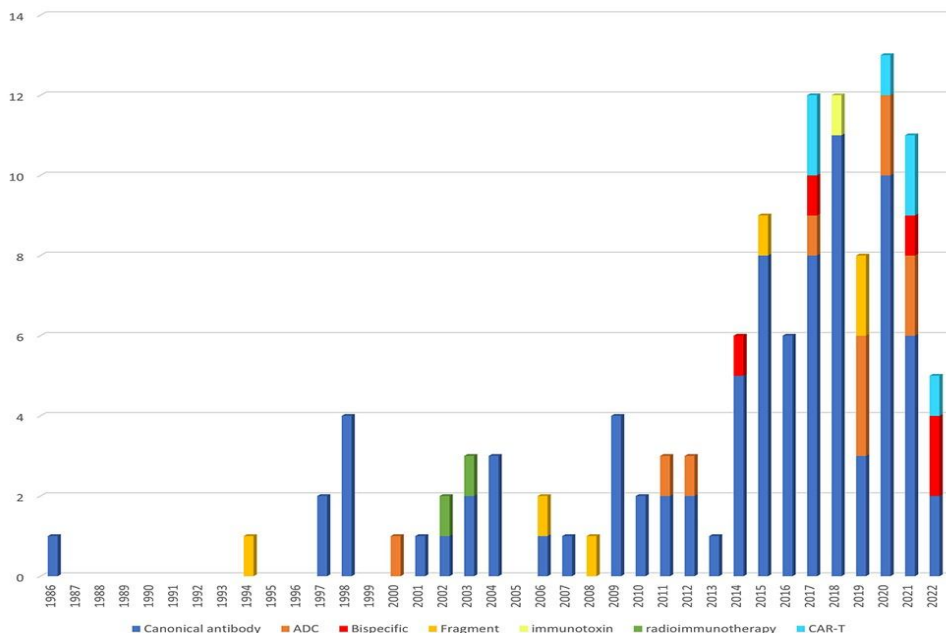
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01

Introduction of Charge Variant Characterization

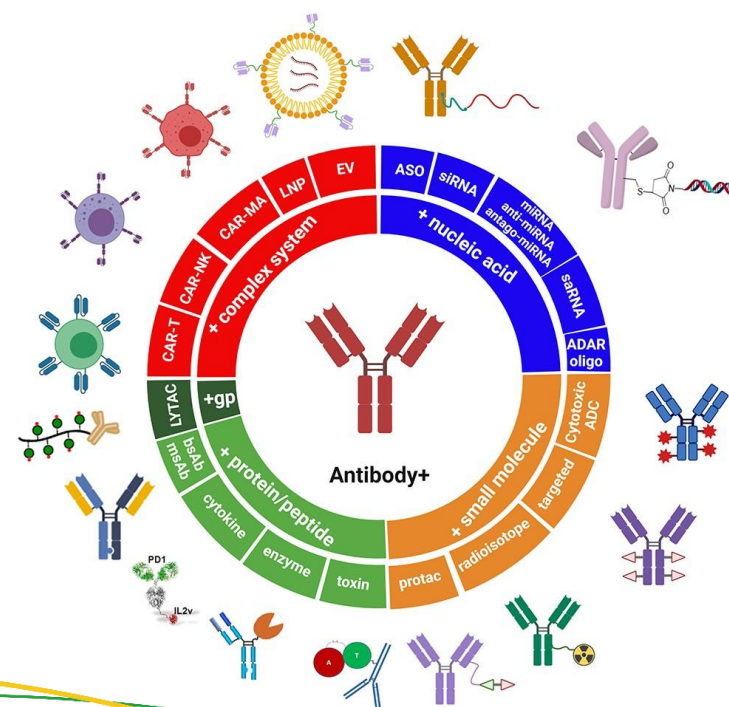
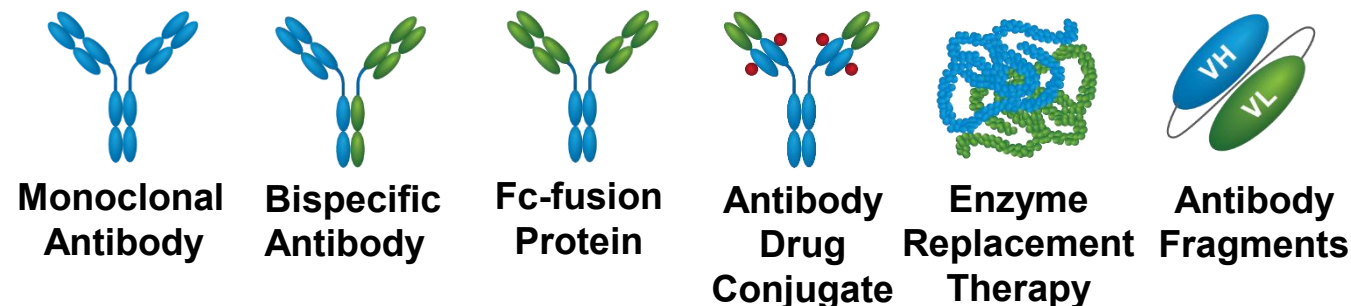


The drug market of protein therapeutics has shown a remarkable expansion in last two decades.



Antibody or antibody-containing therapeutics approved by the FDA as of 31 March 2022.

Antibody Ther, Volume 5, Issue 4, October 2022, Pages 280–287

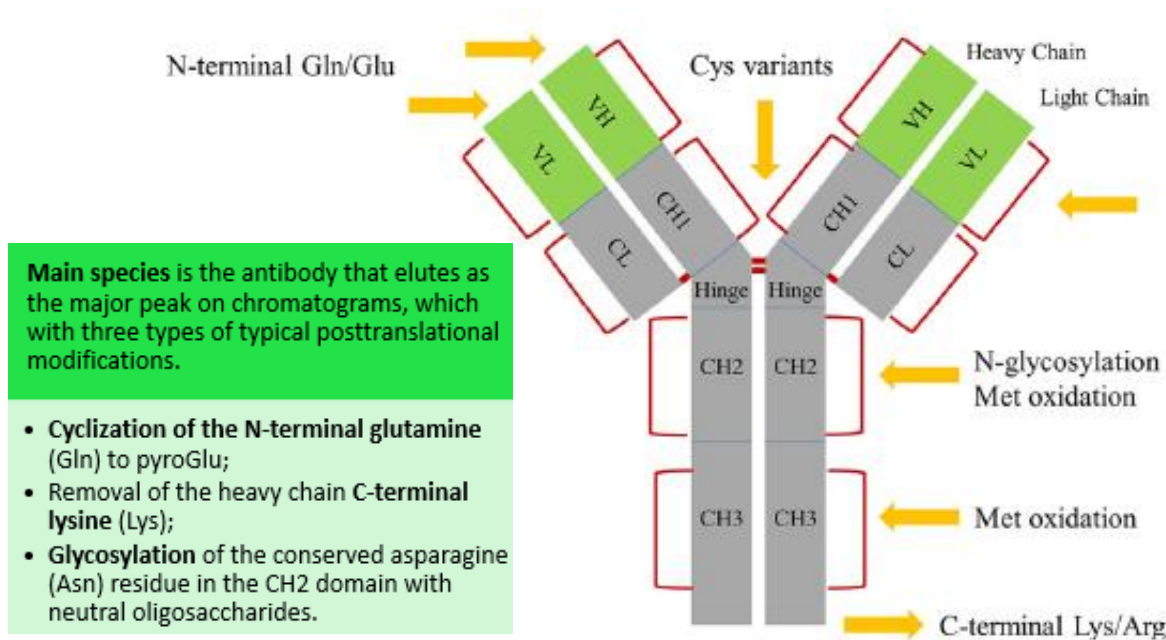




Charge Variant Characterization: Acidic & Basic Species

- Charge variant is one of the most common heterogeneity types for biotherapeutics due to structural modifications and degradation during or after the production process.
- Charge variants including acidic and basic species are common CQAs for protein therapeutics.
- A variety of structural modifications are known to cause the formation of acidic or basic variants.

Acidic species
Sialic acid
Deamidation
Non-classical disulfide linkage
Trisulfide bonds
High mannose
Thiosulfide modification
Glycation
Modification by maleuric acid
Cysteinylation
Reduced disulfide bonds
Non-reduced species
Fragments



Basic species
C-terminal Lys
N-terminal Glu
Isomerization of Asp
Succinimide
Met oxidation
Amidation
Incomplete disulfide bonds
Incomplete removal of leader sequence
Mutation from Ser to Arg
Aglycosylation
Fragments
Aggregates

The background features a light blue gradient with several protein ribbon structures in various colors (teal, light blue, pink, and magenta) and curved lines in yellow, green, and blue. The text is centered in a bold blue font.

Charge Variant Characterization by Online cIEF-MS

02

Characterization of Charge Variants at Different Drug Discovery and CMC Development Stages



Understand the structure of each charge variant; limited functional testing

- 1 Identification of variants significantly changed during developability and stability, force degradation study
- 2 Troubleshooting / investigation during process and formulation development
- 3 Preliminary variants identification to define target range for CQA before major process change or process development for biosimilar product

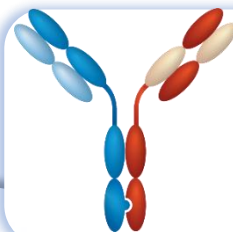


Method Optimization:

- ☐ **Sample preparation:**
- ☐ **cIEF Separation:**
 - Capillary: Neutral-hydrophilic coatings and lengths.
 - Injection amount:
 - Focus voltage:
- ☐ **Interface:**
 - Tip position:
 - Sheath solution:

cIEF-MS to Support Early Stability and Developability Assessment

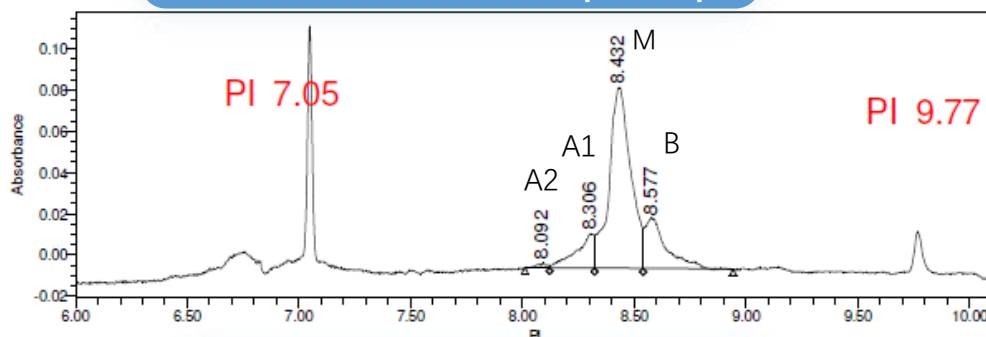
- Charge profile is the most susceptible attribute under thermo stress (40°C) condition.
- icIEF and cIEF-MS show consistent charge profile for unstressed and stressed samples



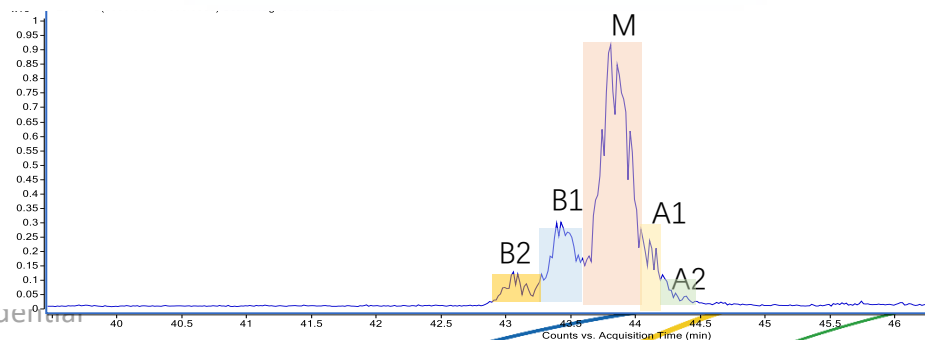
Molecular Information

- Bispecific antibody
- 2 N-glycosylation sites

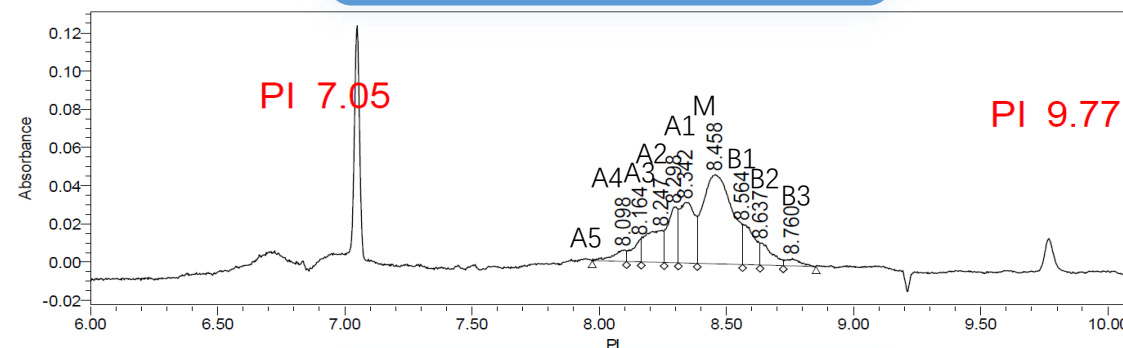
icIEF result of unstressed sample sample



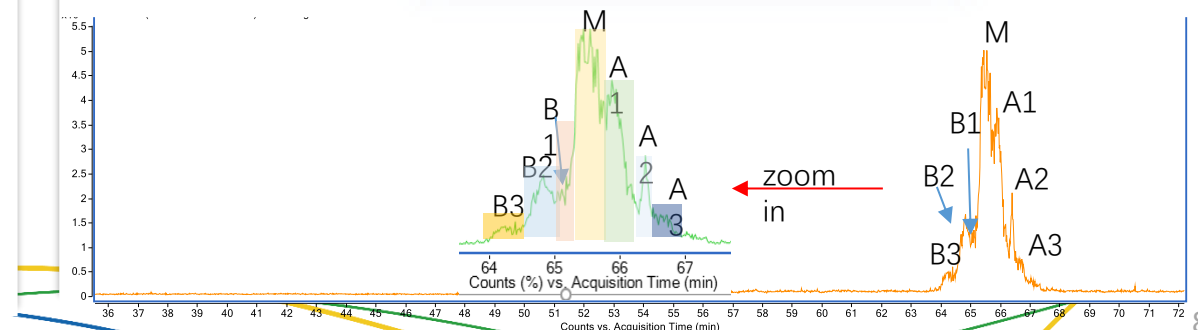
cIEF-Mass result of unstressed sample



icIEF result of stressed sample sample

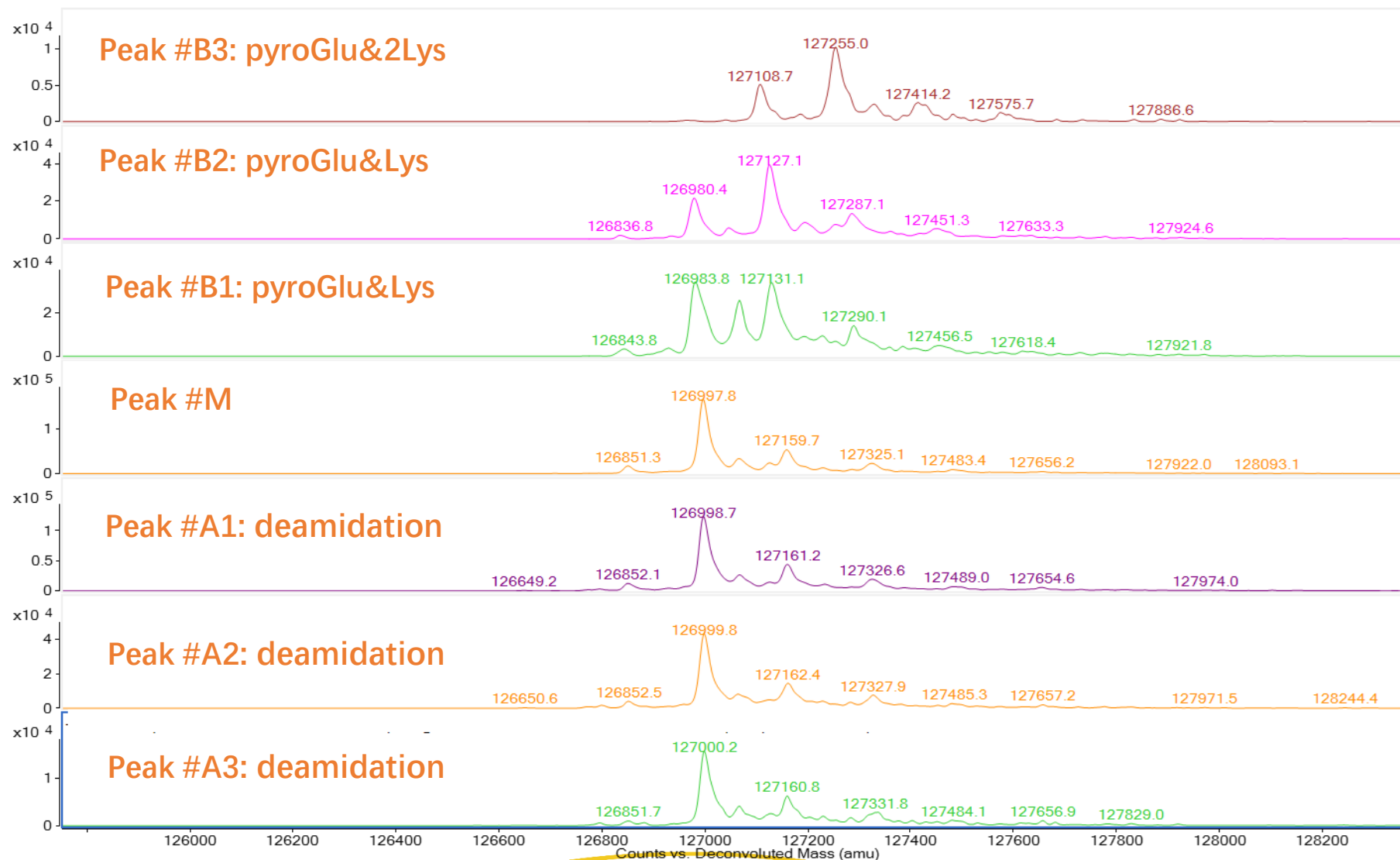


cIEF-Mass result of stressed sample

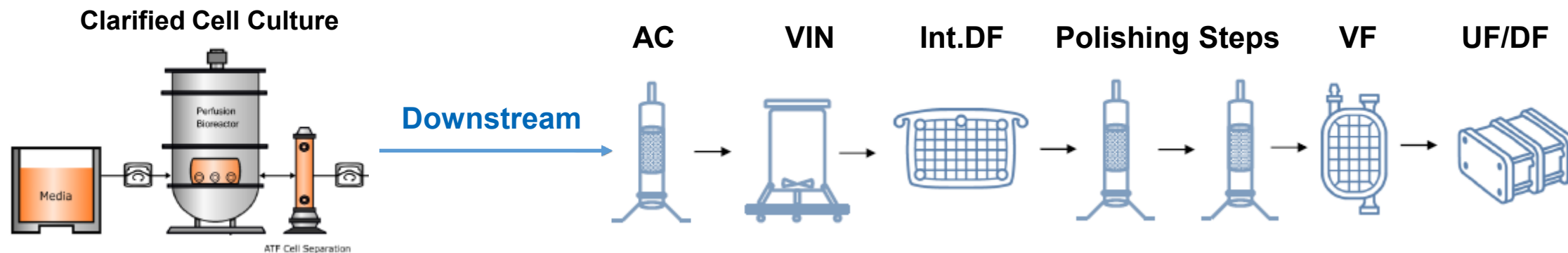




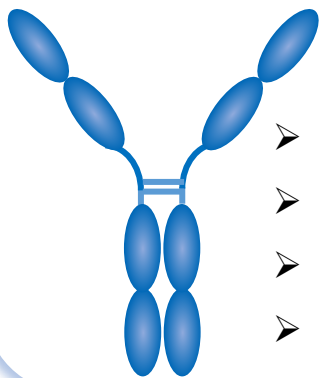
icIEF VS cIEF-MS: Unstressed & Stressed Sample



cIEF-MS to Support Process Development and Manufacturing Investigation/Troubleshooting



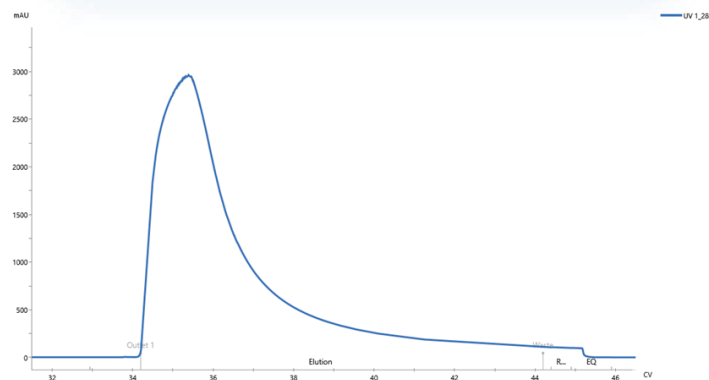
Molecular Information



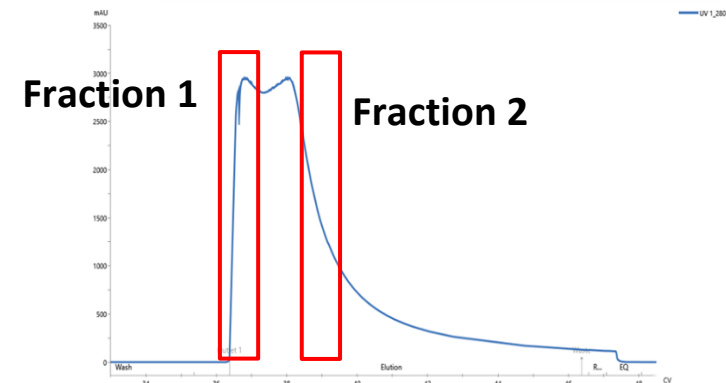
- Fc fusion Protein
- 2 N-glycosylation sites
- 4 O-glycosylation sites
- 20 disulfide bonds

Doublet Peak Issue during Downstream Process

Low Loading with single peak



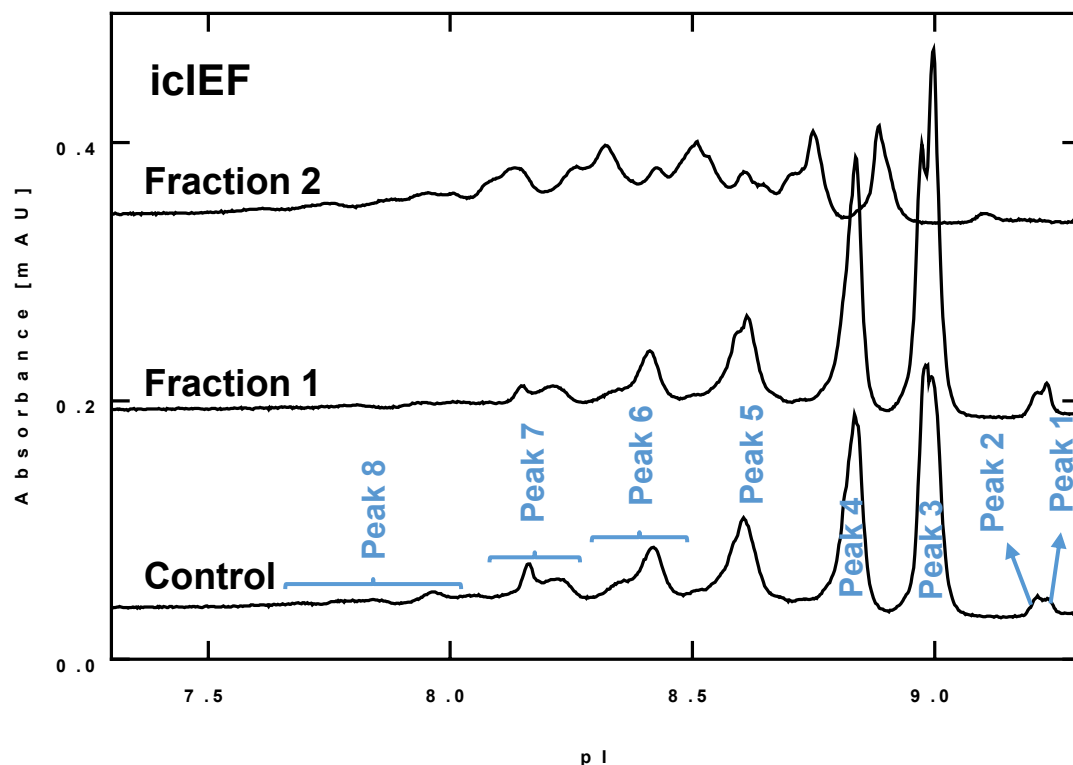
High Loading with doublet peak



Process Investigation: cIEF-MS to Decipher the ID of Doublet Peaks

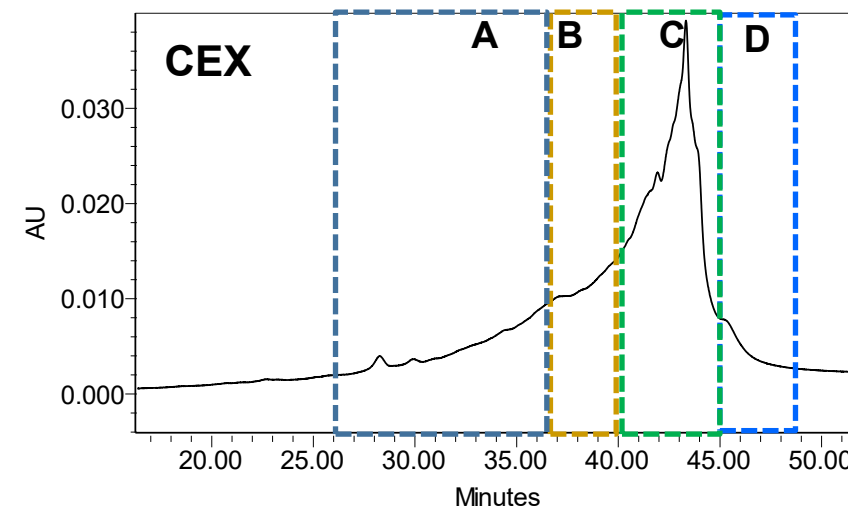
Investigation:

- ✓ Fraction 1 shows higher peak 3-4
- ✓ Fraction 2 shows higher peak 5-8



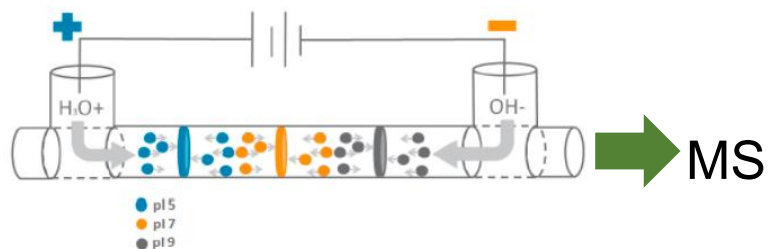
Challenge

- CEX fractionation profile is not aligned with icIEF profile



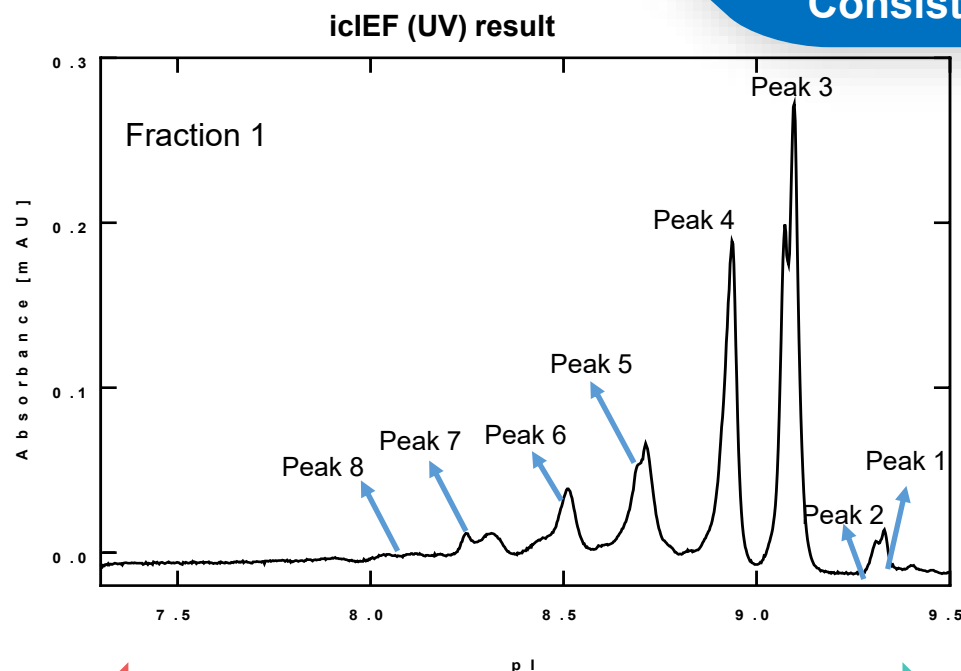
Orthogonal cIEF-MS method become the only choice to identify each charge variants in icIEF profile

Process Investigation: cIEF-MS to Decipher the ID of Doublet Peaks



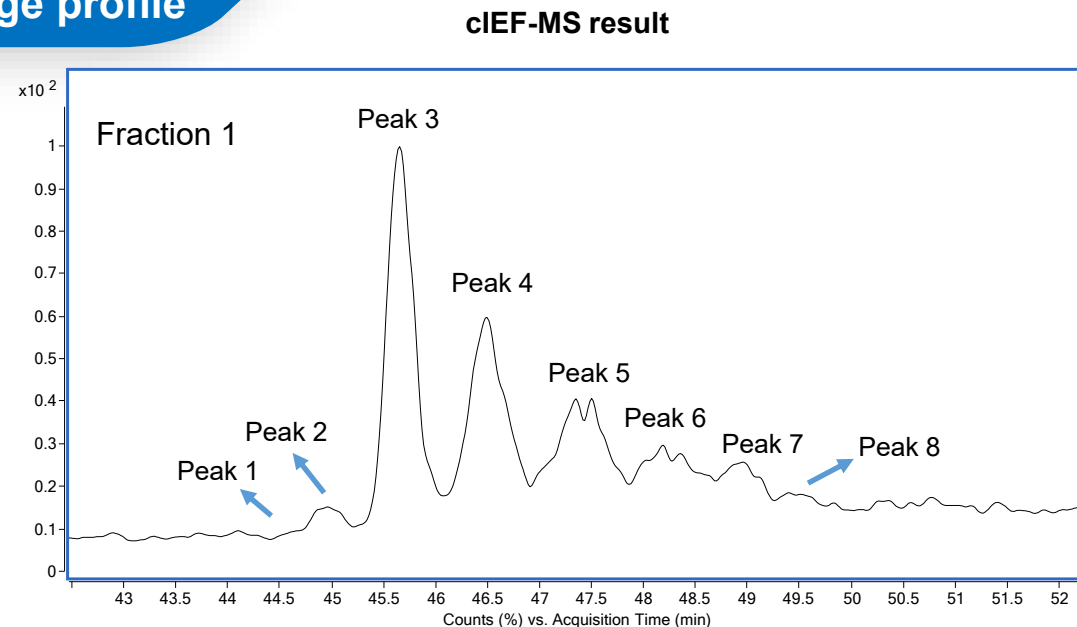
- icIEF and cIEF-MS show very consistent charge profile

Consistent charge profile



Acidic peak

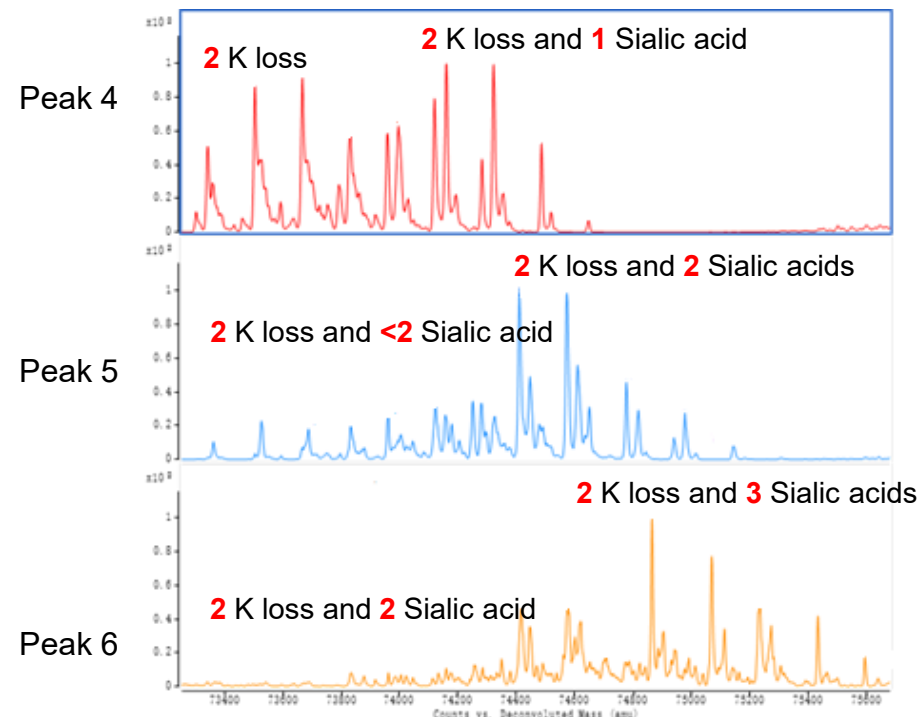
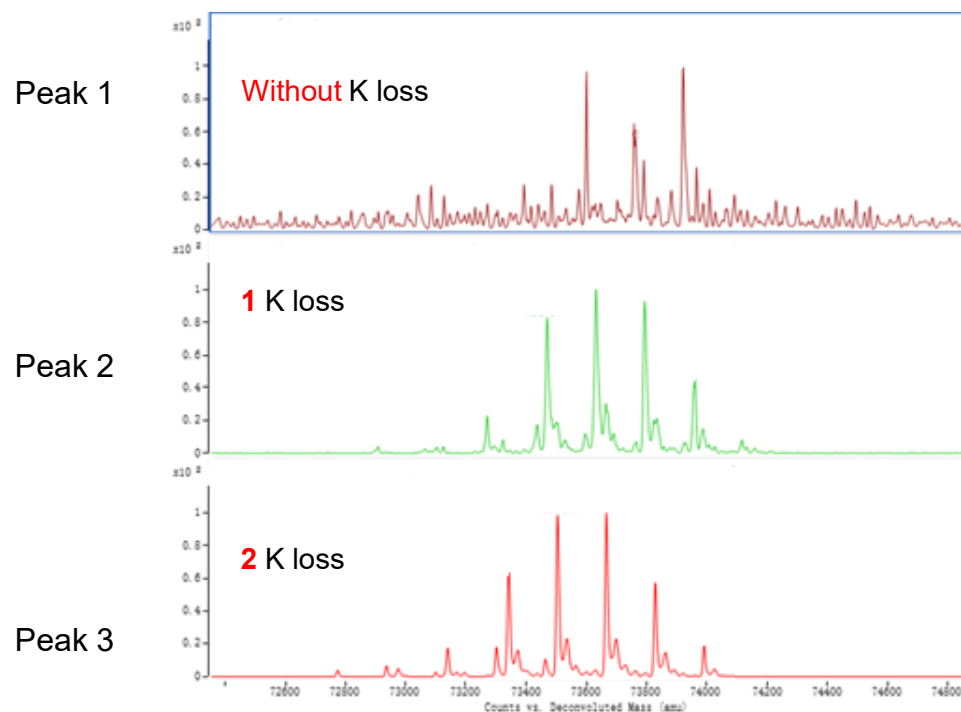
Basic peak



Basic peak

Acidic peak

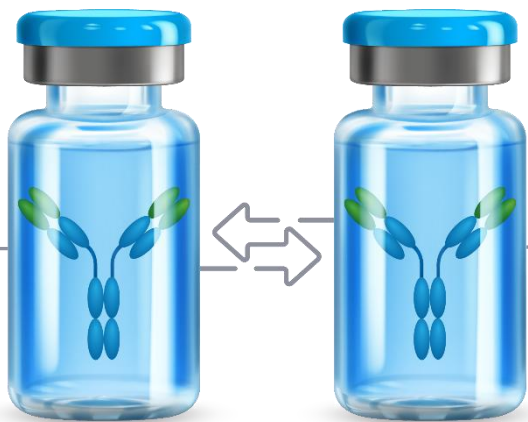
Process Investigation: cIEF-MS to Decipher the ID of Doublet Peaks



Peak # in cIEF	Modifications
1	Neutral glycan +C-terminal K
2	Neutral glycan +C-terminal K Loss
3	Neutral glycan +2XC-terminal K Loss
4	Acidic glycan (0-1 SA)+2XC-terminal K Loss
5	Acidic glycan (1-3 SA)+2XC-terminal K Loss
6-8	Acidic glycan (>3 SA)+2XC-terminal K Loss

- The doublet peak is mainly due to different sialylation contents
- Both peaks represented product-related variants and can be combined in one pool

Charge Variant Identification for Biosimilar Program & Post-approval changes for 2nd Generation Product

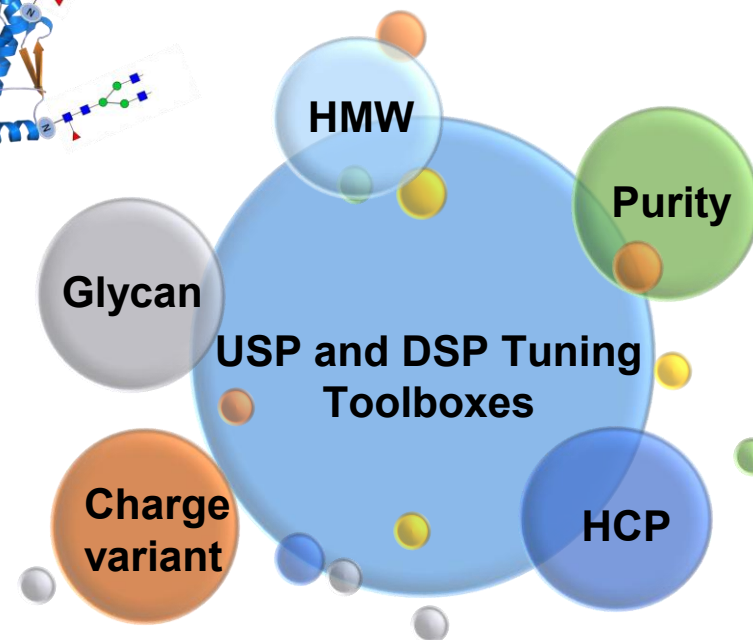
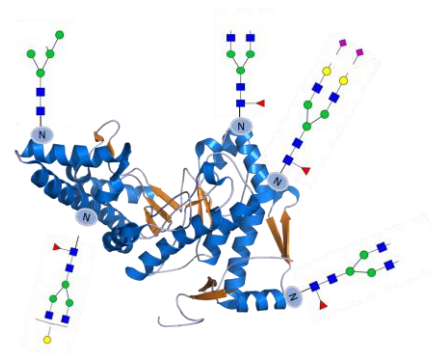


Reference Product

Biosimilar

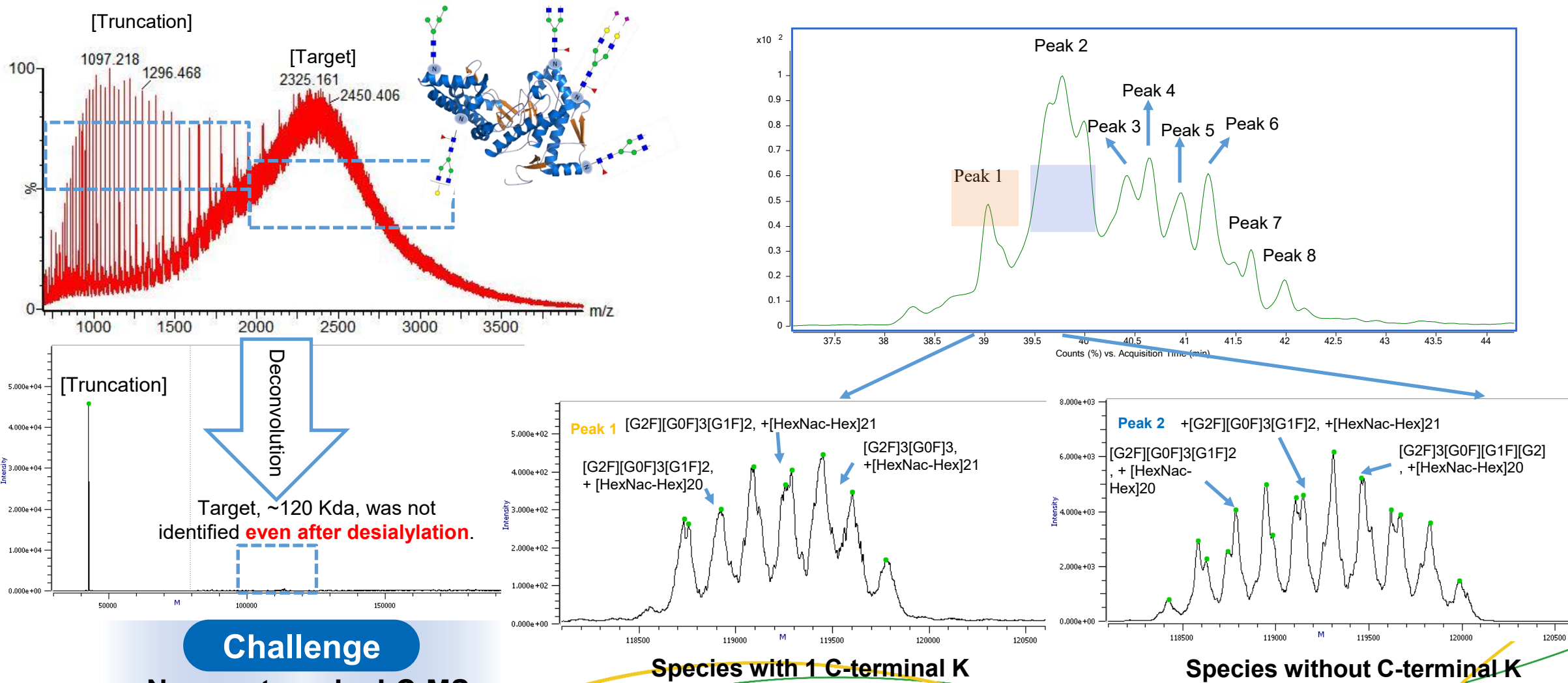
Biosimilar Program & Post-approval Changes for 2nd Generation Commercial Product

- Meet commercial supply demands with the lowest possible production costs (CoGs)
- Additional optimization and fine-tuning of CQA, especially **charge variants**, are essential during process development to meet comparability/similarity acceptance criteria



Defining target ranges through preliminary charge variant characterization is essential prior to initiating process development

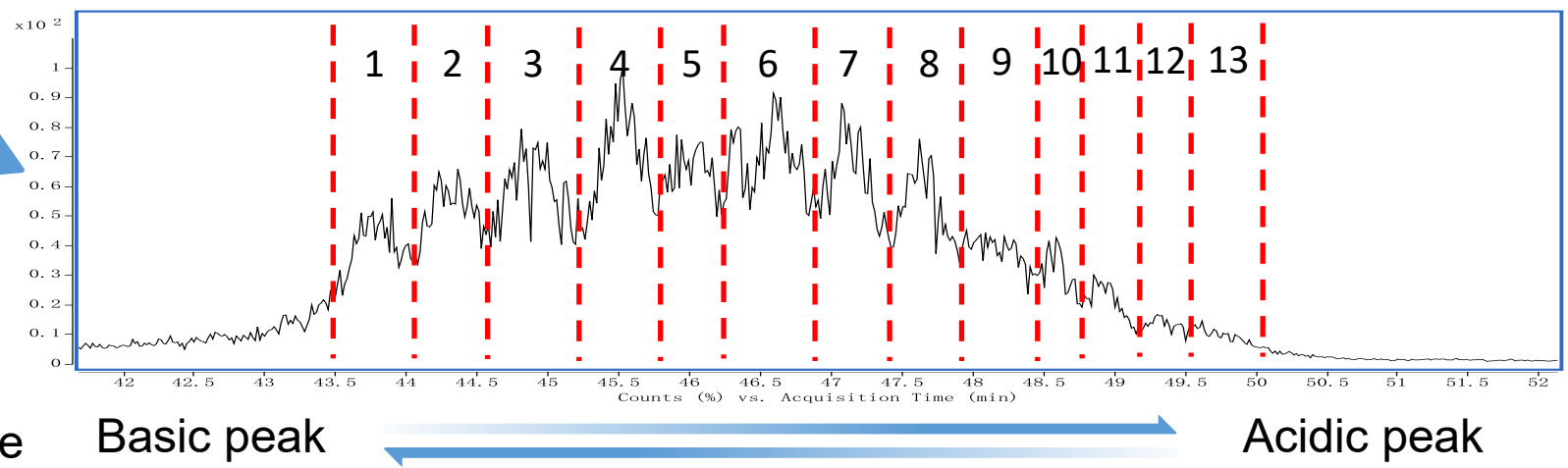
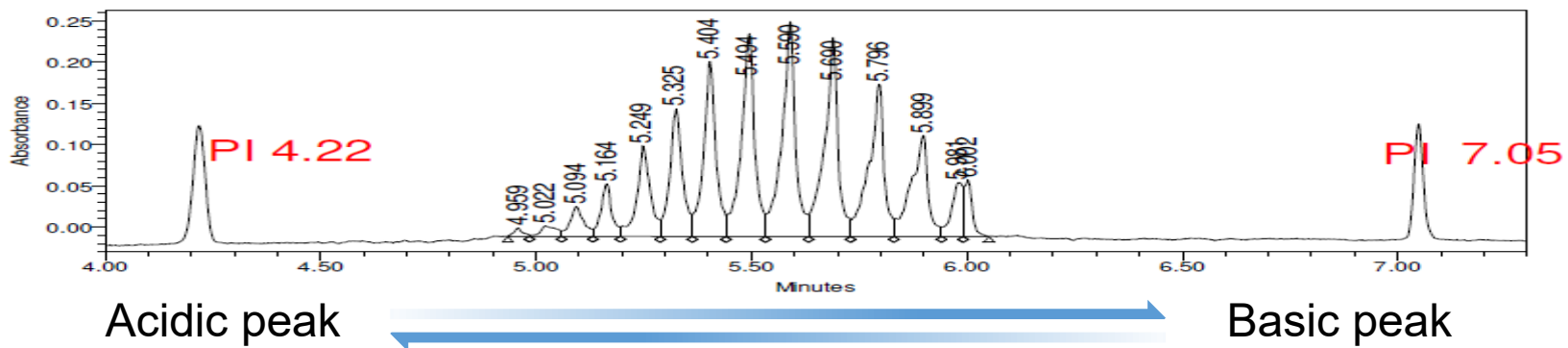
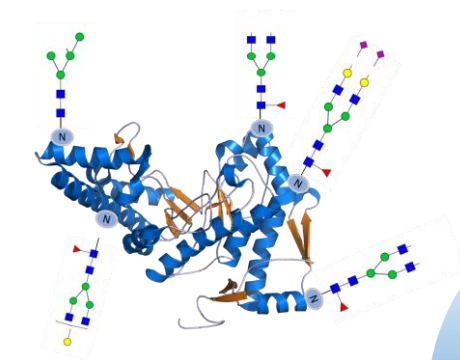
Complex biotherapeutics – Highly O-glycosylated variants identified from cIEF-MS





cIEF-MS Based Characterization of Highly Sialylated Protein

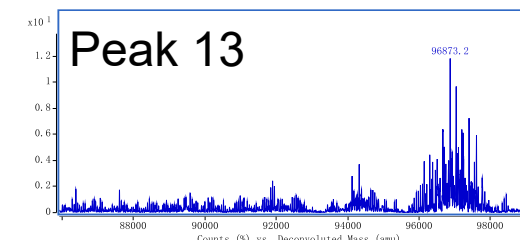
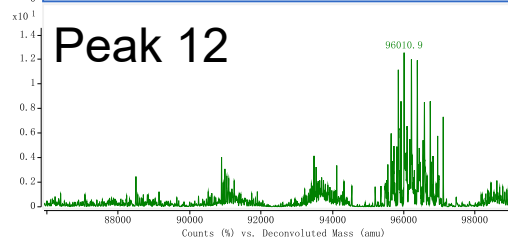
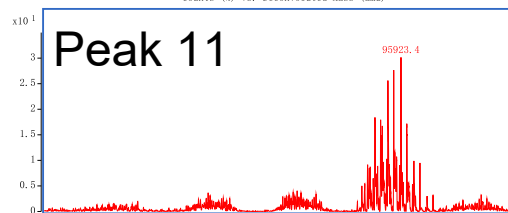
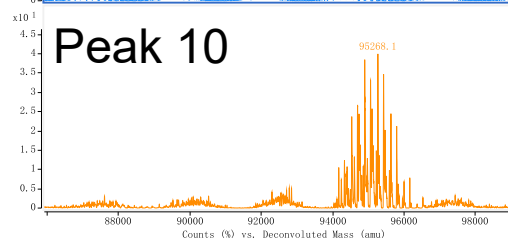
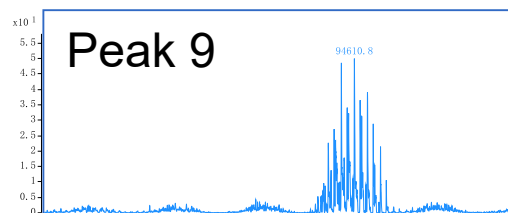
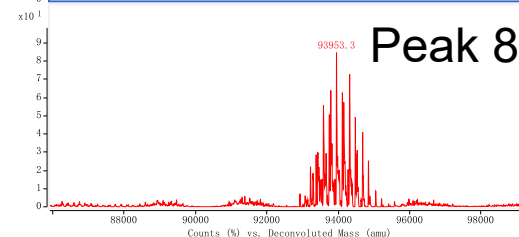
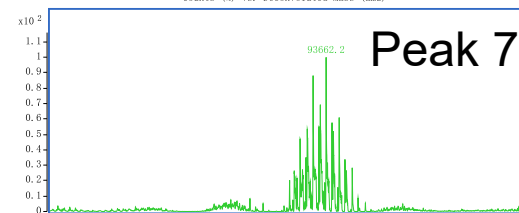
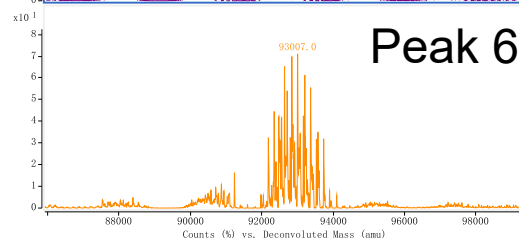
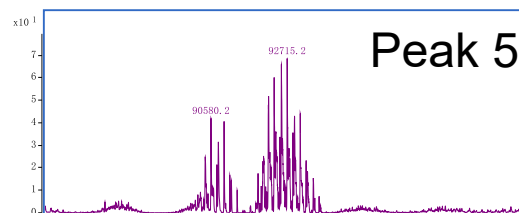
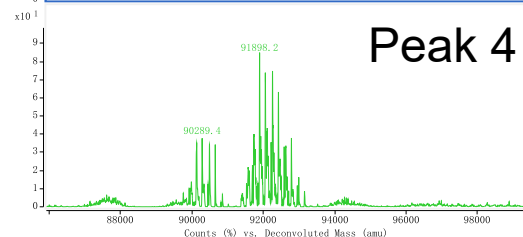
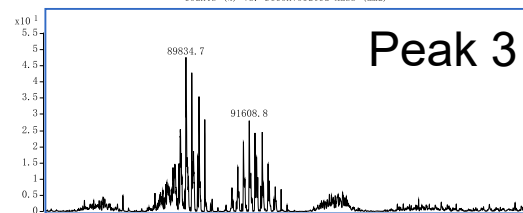
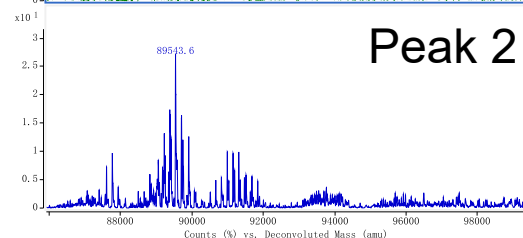
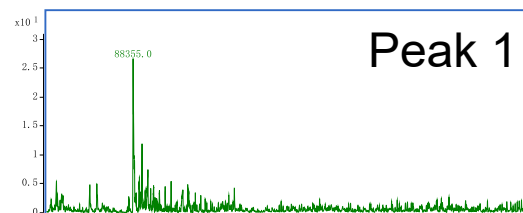
Complex biotherapeutics – Highly glycosylate/sialylated variants identified by cIEF-MS





cIEF-MS Based Characterization of Highly Sialylated Protein

Complex biotherapeutics – Highly glycosylate/sialylated variants identified by cIEF-MS

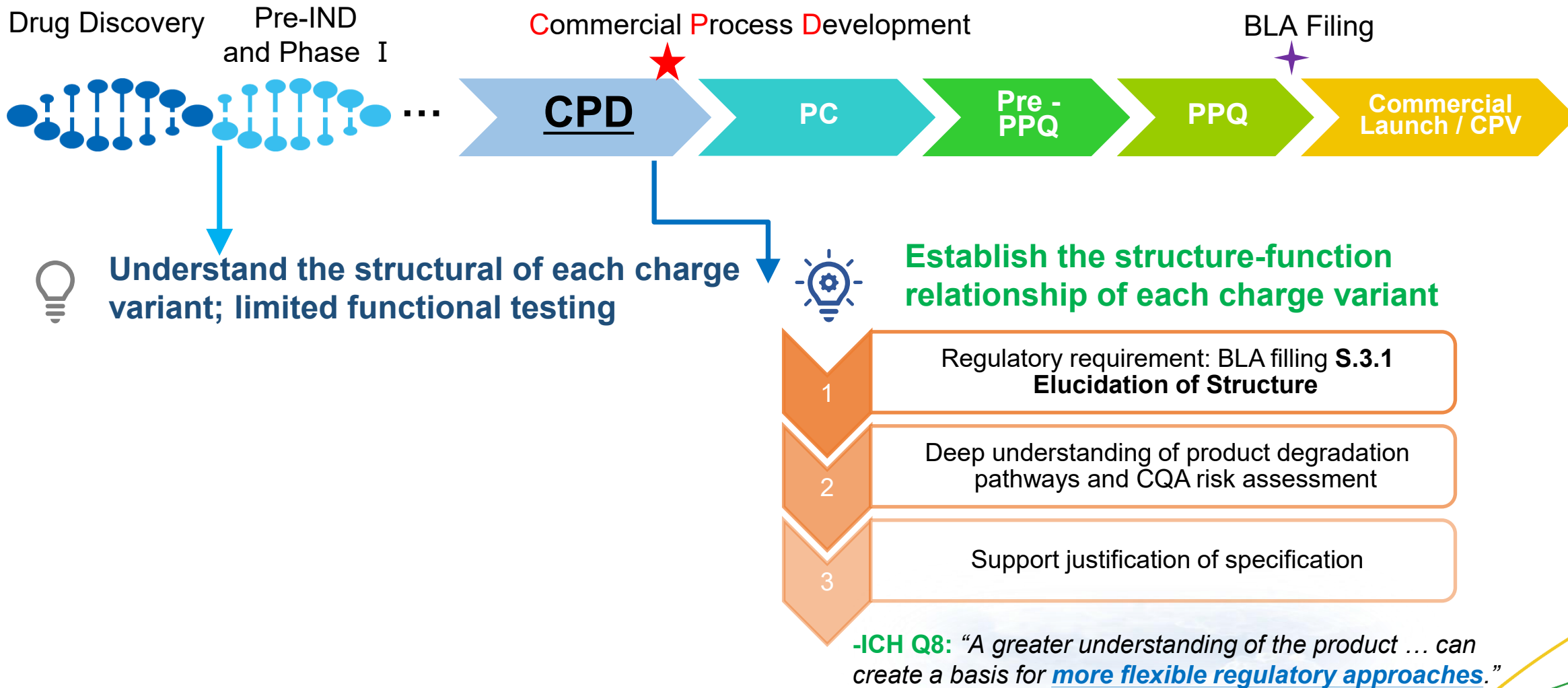


Peak #	Number of Neu5Ac
1	0
2	1
3	2
4	3
5	4
6	5
7	6
8	7
9	8
10	9
11	10
12	11
13	12

03 Charge Variant Characterization by Offline 'Discovery' Assisted Fractionation

The background features a light blue gradient with a large, semi-transparent water droplet on the right side. Several thin, curved lines in blue, green, and yellow sweep across the lower half of the image, creating a dynamic, scientific feel.

Characterization of Charge Variants at Different Drug Discovery and CMC Development Stages



Offline Charge Variant Characterization by Fractionation

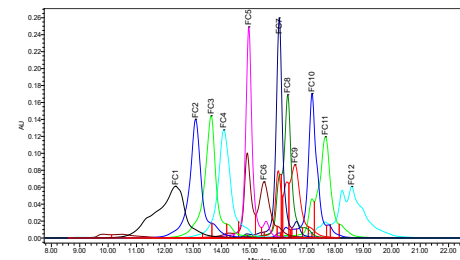
-Traditional Strategy



Scale up to semi-preparative column
and check performance



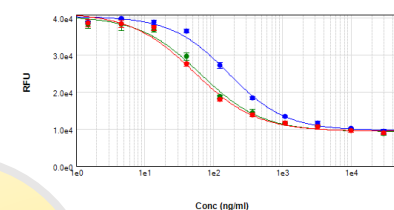
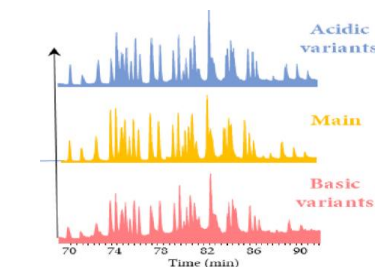
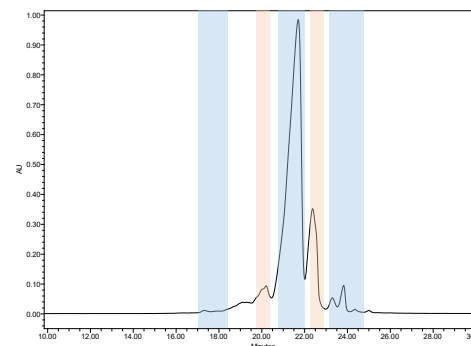
iCIEF and intact MS
analysis to determine
fraction pooling



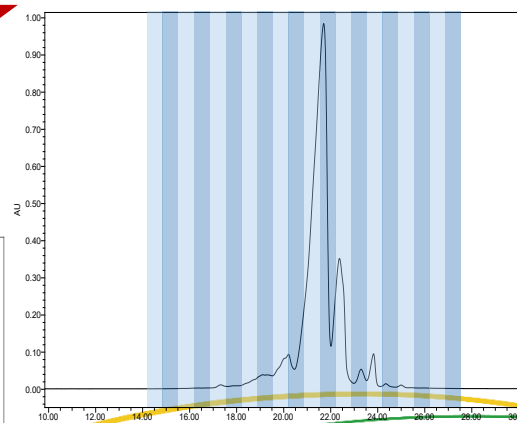
Purity Confirmation

Structure/function
characterization

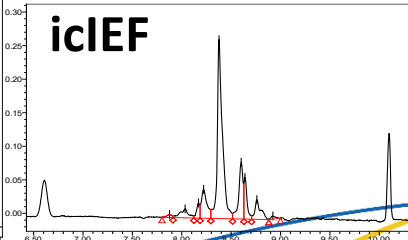
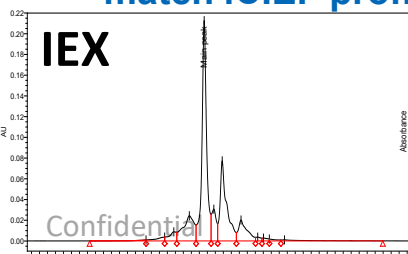
Formal fractionation collection



Pre-study before fractionation



IEX method development on
analytical column to
match iCIEF profile



Control
Strategy
Define CQA/non-
CQA





Challenge 1: IEX Method Development and Scale up

✓ IEX Development Process:

Method development on analytical column

Scale up of analytical method

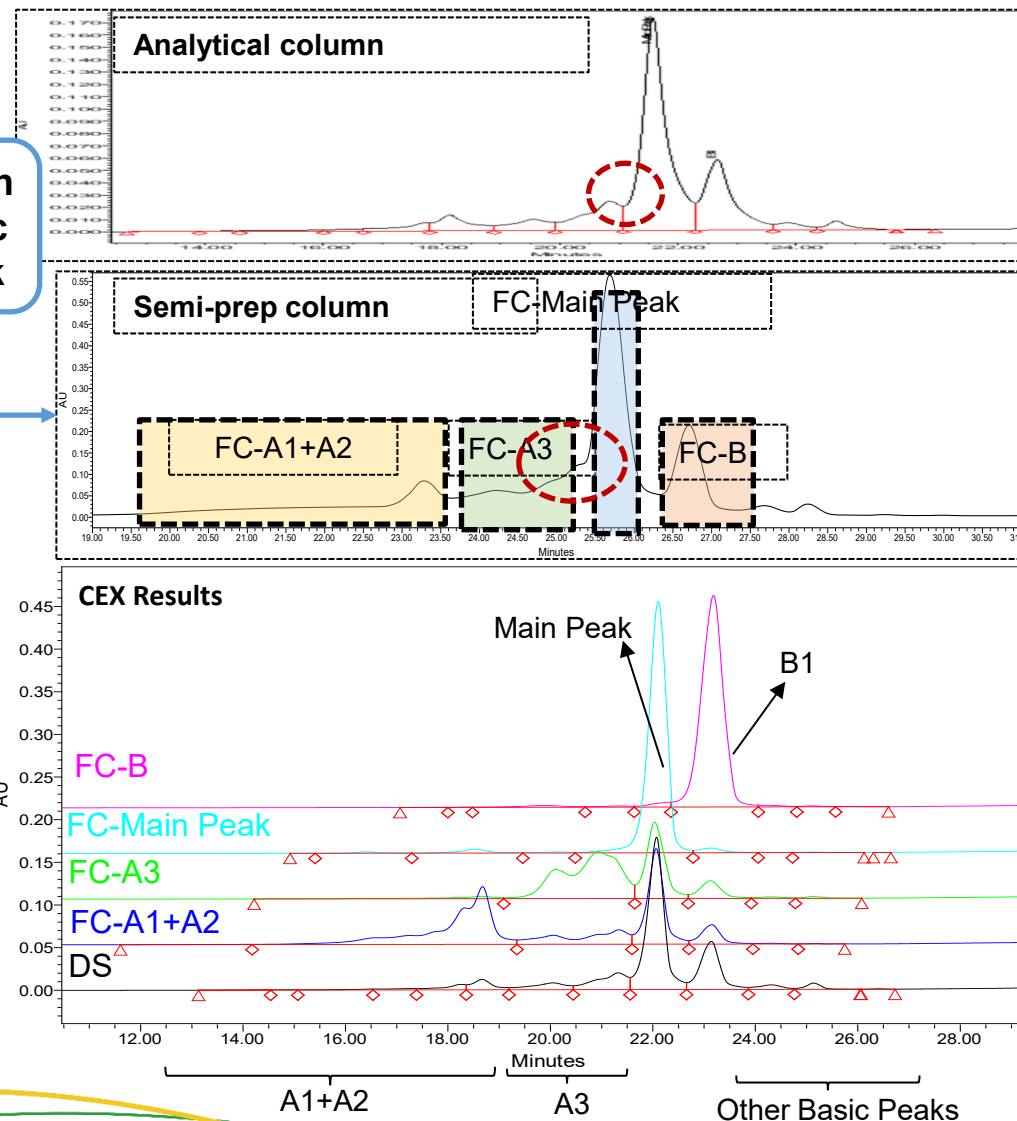
Optimization of Flow rate and loading amount

CEX Purity Analysis for Fractions

$$F2 = F1 \times \frac{d2^2}{d1^2}$$
$$M2 = M1 \times \frac{L2}{L1} \times \frac{d2^2}{d1^2}$$

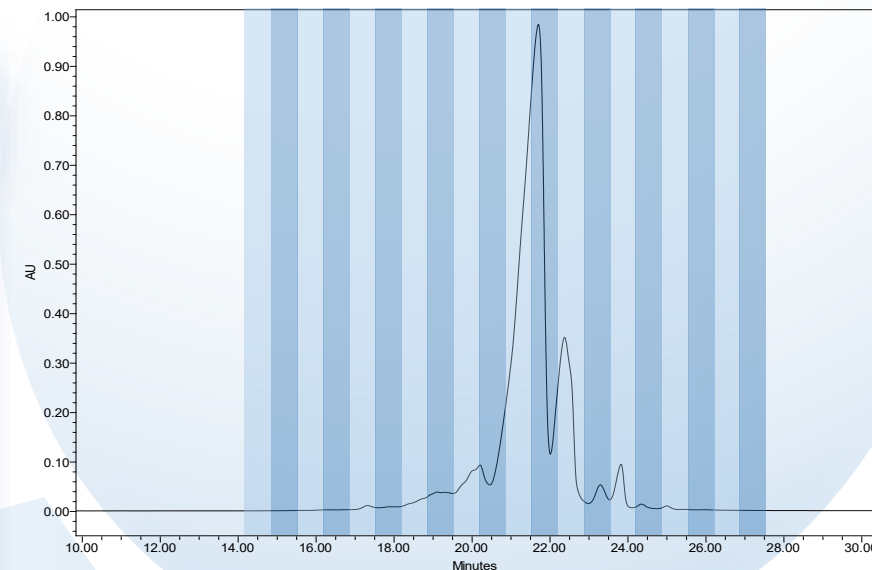
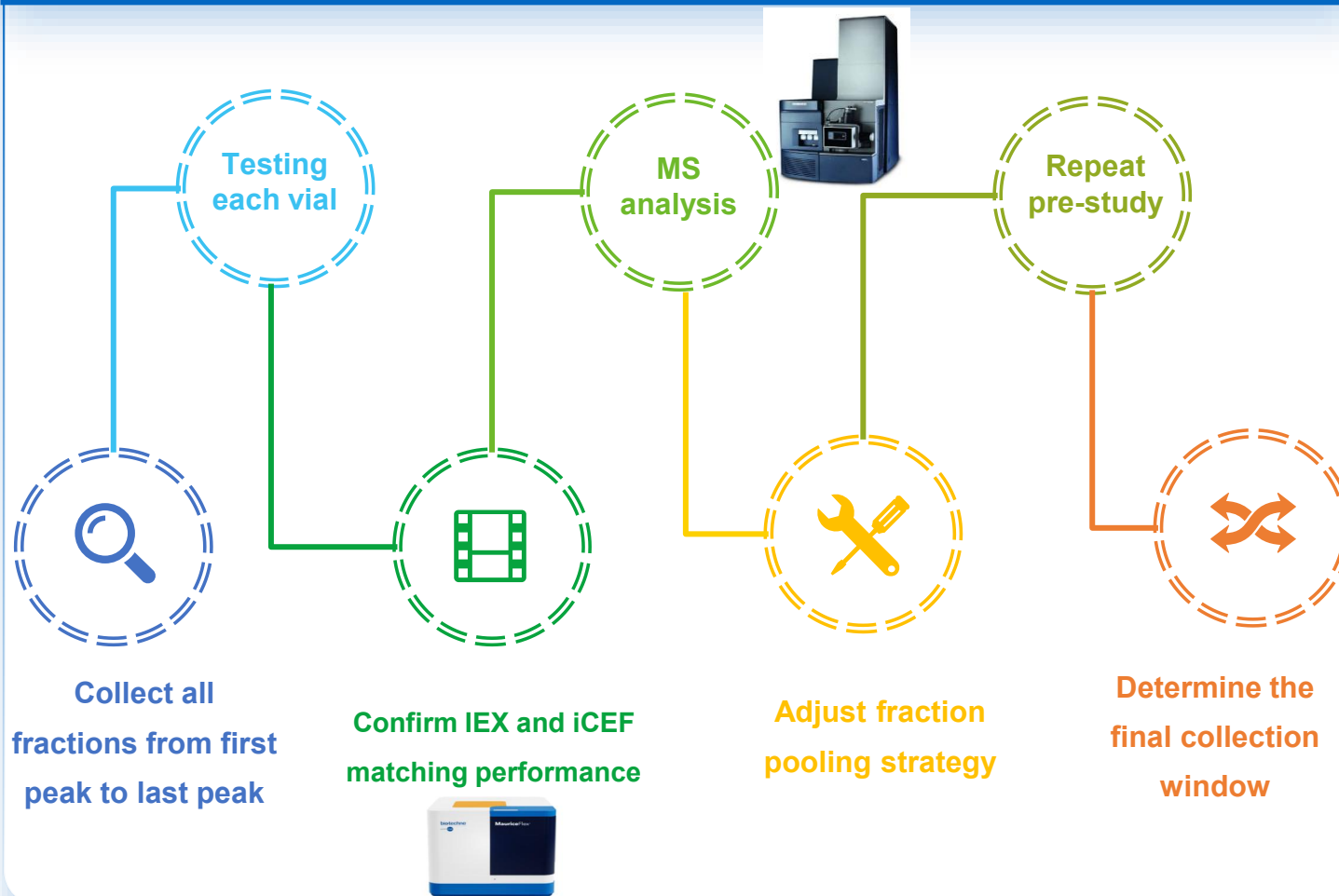
Poor resolution between acidic and main peak

Low CEX purity of acidic fractions

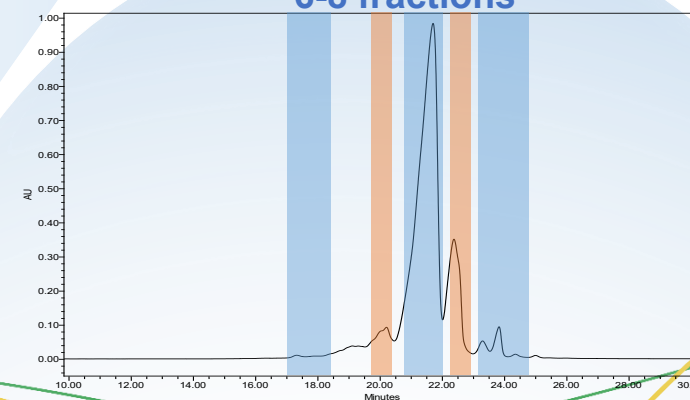


Challenge2: Pre-study to Determine Peak Collection Window and Fraction Pooling Strategy

Fractionation Windows Determine Workflow



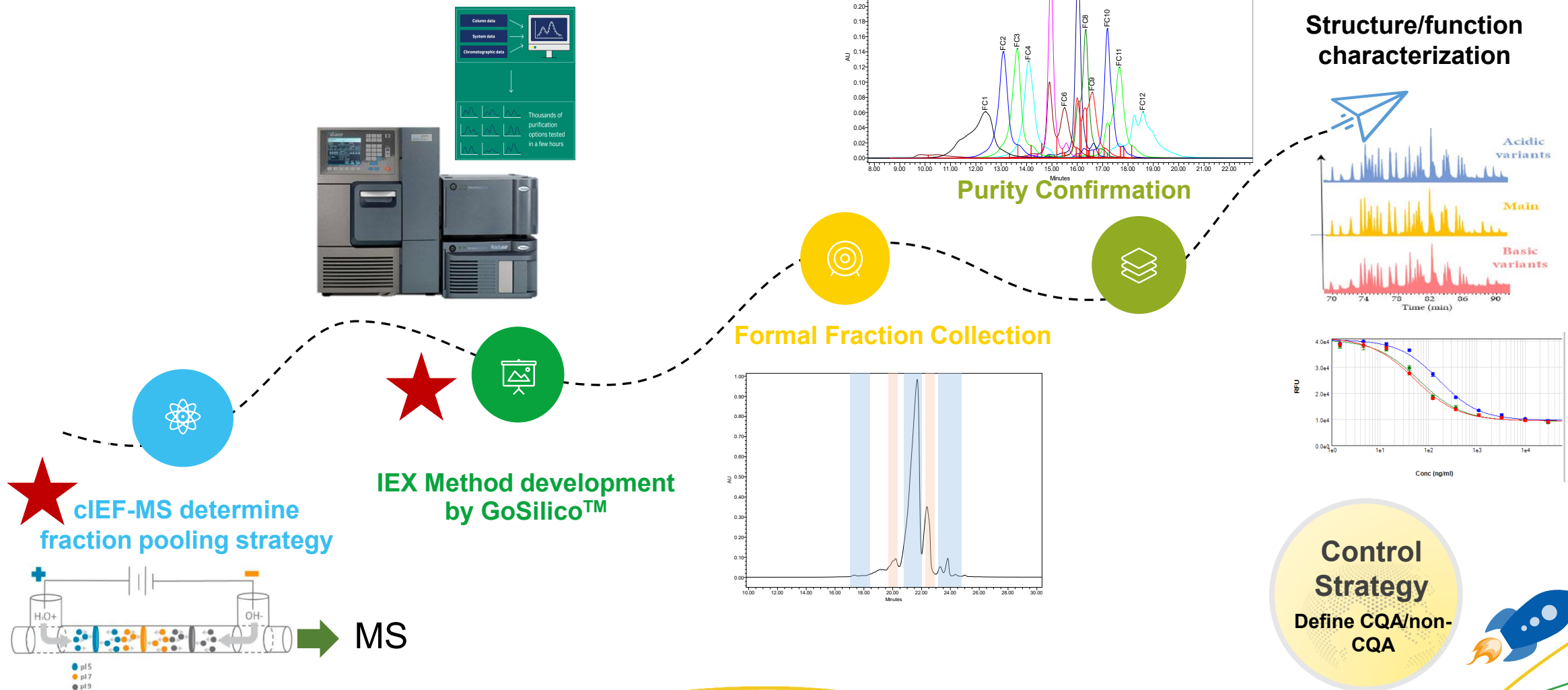
**Before pooling
20-30 fractions**



**After pooling
6-8 fractions**

Offline Charge Variant Characterization by Fractionation

-‘Discovery’ Assisted Strategy





IEX Method Development –Gosilico™ Modelling

Significantly reduced rounds of experiments through application of mechanistic models using computer simulations (GoSilico™ Chromatography Modeling Software).

Column data

System data

Chromatographic data

A variety of models available and keep updating

$K_{L,i} \frac{\partial q_i}{\partial t} = k_{eq,i} \cdot q_{max,i} \cdot \left(1 - \sum_{j=1}^N \frac{q_j}{q_{max,j}}\right) \cdot c_{p,i} - q_i$	Langmuir ("Langmuir"), cf. Schmidt-Traub et al., 2012, [ISBN:978-3-527-64930-3]
$K_{L,i} \frac{\partial q_i}{\partial t} = k_{eq,i} \cdot q_{max,i} \cdot \left(1 - \sum_{j=1}^N \frac{q_j}{q_{max,j}}\right) \cdot c_{p,i} - q_i$	Langmuir with Modifier ("Langmuir with Modifier")
$K_{SMA,i} \frac{\partial q_i}{\partial t} = k_{eq,i} \cdot q_{max,i} \cdot c_{p,i} - q_i \cdot c_{p,i}^{n_{p,i}}$ with $q_{max,i} = A_{SMA,i} \cdot \sum_{j=1}^N \frac{v_j + d_j}{f_j} \cdot q_j$	Steric mass action ("SMA"), cf. Schmidt-Traub et al., 2012, [ISBN:978-3-527-64930-3]
$K_{GIE,i} \frac{\partial q_i}{\partial t} = k_{eq,i} \cdot q_{max,i} \exp(K_{GIE,i} \cdot c_{p,i} + K_{proton,i} \cdot c_{p,i}) \cdot c_{p,i} - q_i \cdot c_{p,i}^{n_{p,i}}$	Generalized Ion Exchange isotherm ("IEC 2008") by Møllerup et al., 2008, [DOI:10.1002/ceat.200800082]
$K_{pH,i} \frac{\partial q_i}{\partial t} = k_{eq,i} \exp(K_{pH,i} \cdot pH + K_{GIE,i} \cdot pH^2) \cdot q_{max,i} \cdot c_{p,i} - q_i \cdot c_{p,i}^{n_{p,i}}$ with $v(\text{pH}) = v_1 + v_2 \cdot \text{pH}$	pH-extended SMA ("SMA With pH 2017") adapted from Hunt et al., 2017, [DOI:10.1002/9781119031116.ch13]
$K_{pH,i} \frac{\partial q_i}{\partial t} = \exp(-G_{pH,i} + v_2 G_{pH,i} \cdot pH) \cdot q_{max,i} \cdot c_{p,i} - q_i \cdot c_{p,i}^{n_{p,i}}$ with $v_1 = \frac{-N_{GIE,i}}{1 + 10^{pH - pK_{a,i}}} + \frac{-N_{pH,i}}{1 + 10^{pH - pK_{a,i}}} + \frac{-N_{GIE,i}}{1 + 10^{pH - pK_{a,i}}} + \frac{-N_{pH,i}}{1 + 10^{pH - pK_{a,i}}}$ + $\frac{N_{GIE,i}}{1 + 10^{pH - pK_{a,i}}}$	pH-dependent Ion Exchange isotherm ("IEC 2014") by Schmidt et al., 2014, [DOI:10.1002/jssc.201301007]

Thousands of purification options tested in a few hours

1

2

3

4

5

6

System characterization

Column characterization

Protein chromatography experiments

Model calibration (with software)

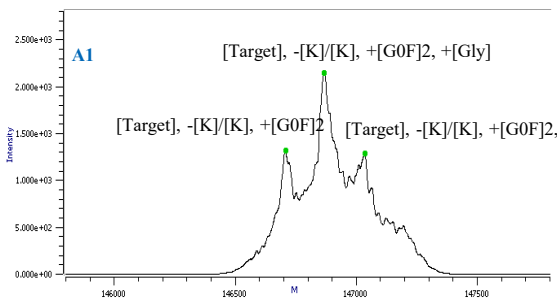
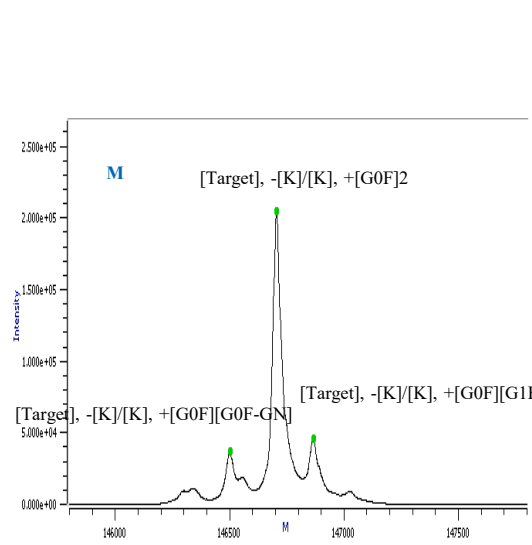
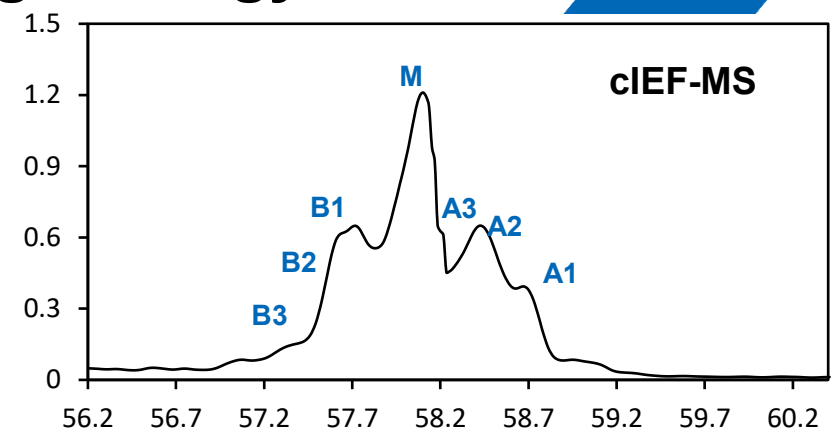
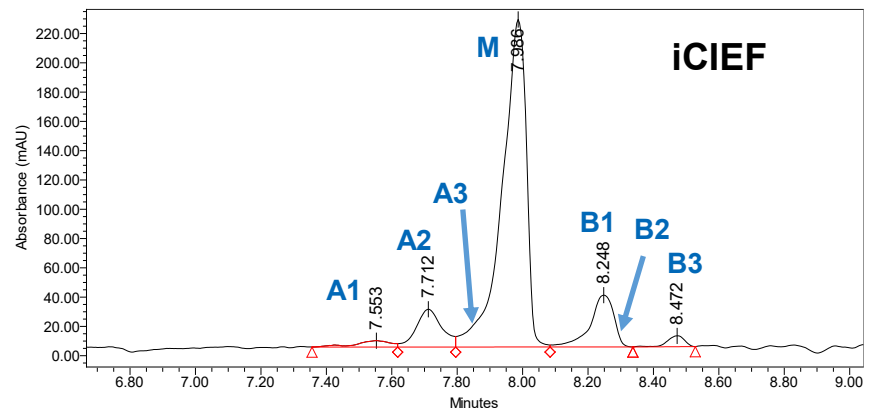
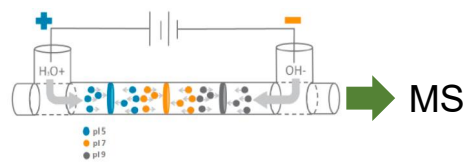
Model utilization (simulated experiments)

Model validation

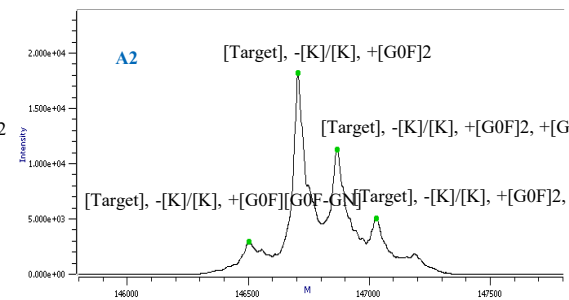
Molecule Type	Timeline-1	Timeline-2	Timeline-3	Timeline-4	Total Timeline
mAb	2-3 days	2 days	1 day	1 day	< 1 weeks with significantly reduced Timeline
Complex Molecule		2 days	2 days	1 day	



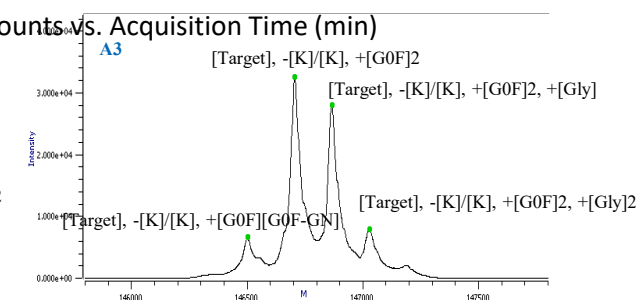
cIEF-MS Determine Peak Boundary and Pooling Strategy



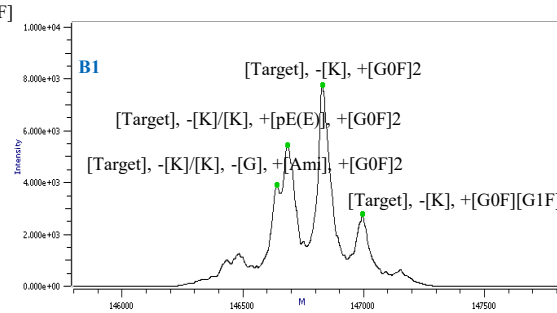
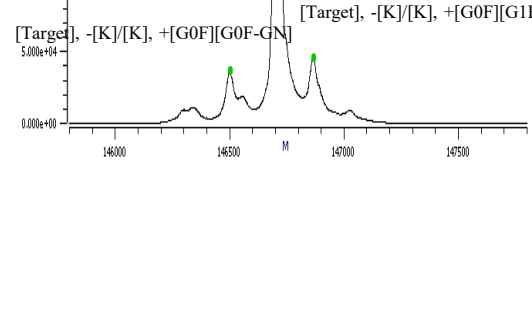
Peak #A1: Glycation ?



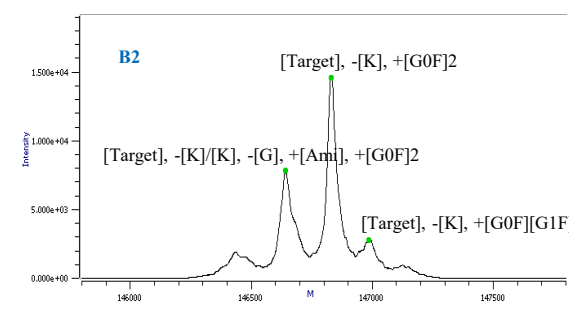
Peak #A2: deamidation/glycation



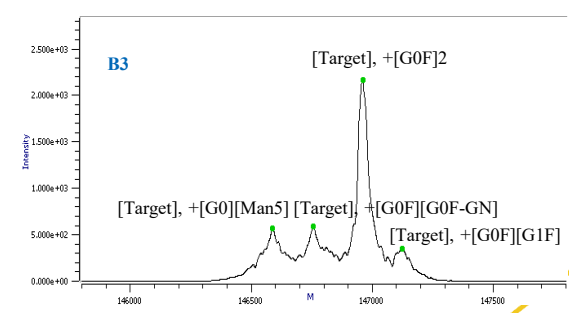
Peak #A3: Glycation



Peak #B1: pyroGlu(E)&Lys



Peak #B2: Lys



Peak #B3: 2Lys

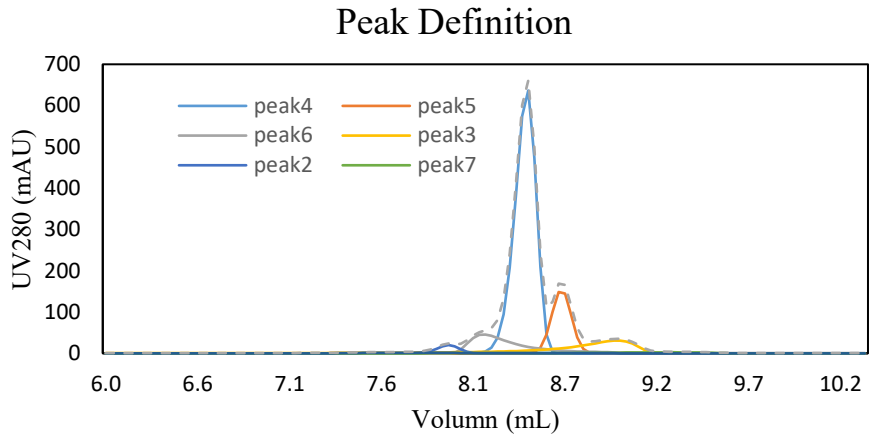


IEX (HPLC) Method Development –Gosilico™ Modelling

Calibration run

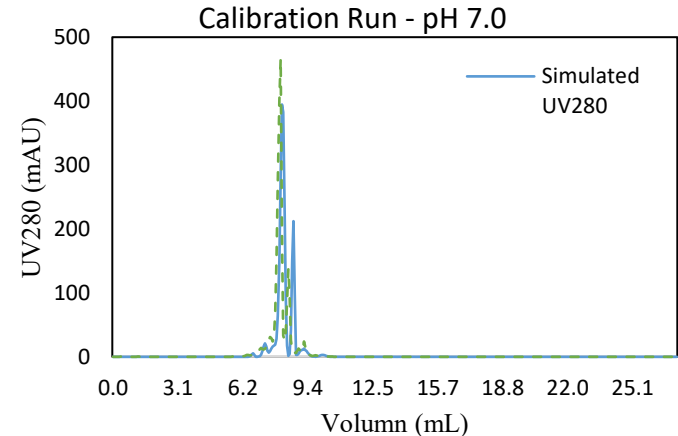
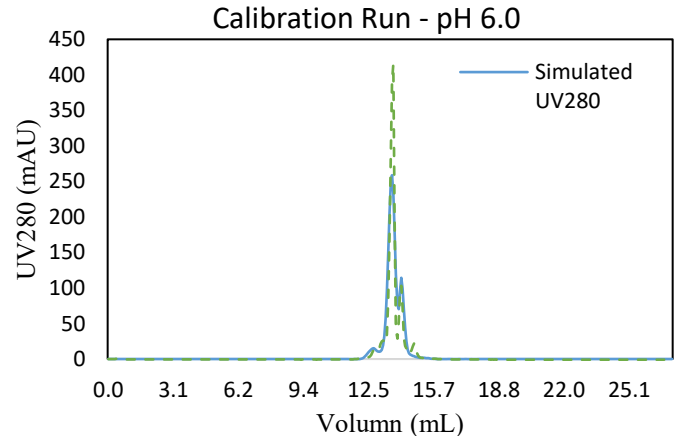
Experiment run	Load density (g/L resin)	Elution length	pH
Gradient 1	1	0~100% 10 CV	6.0
Gradient 2	1	0~100% 20 CV	6.0
Gradient 3	1	0~100% 30 CV	6.0
Gradient 4	1	0~100% 20 CV	5.5
Gradient 5	1	0~100% 20 CV	7.0

Define Peak Component: 7 peaks based on cIEF-MS identification



Model Selection and Calibration in GoSilico

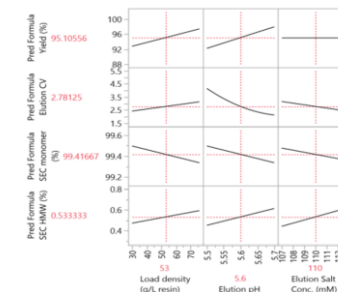
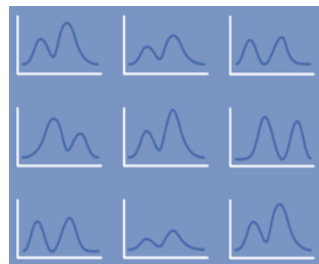
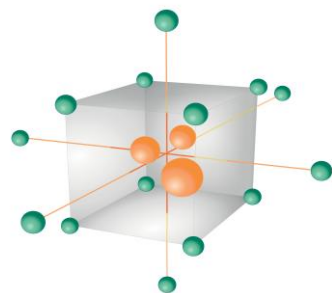
Column Model	Equilibrium Dispersive
Pore Model	No Pore Model
Adsorption Model	SMA with pH 2017





IEX (HPLC) Method Development –Gosilico Simulation

Model Utilization and Simulated Chromatography



- Good resolution
- Appropriate RT
- Good peak response

Model Recommended Process Parameters and Simulated Chromatography

Process parameters	Parameters range
--------------------	------------------

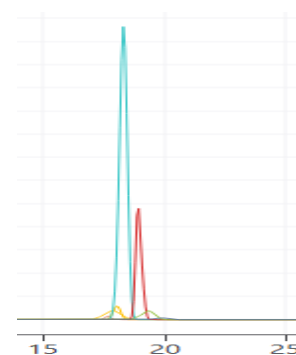
Buffer A pH	6.0-7.0
-------------	---------

Buffer B pH	6.0-7.0
-------------	---------

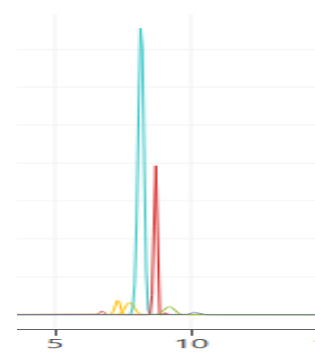
Buffer B NaCl conc. (mM)	300-600
--------------------------	---------

Stage 1 (0-20 min) A% gradient	85-95
--------------------------------	-------

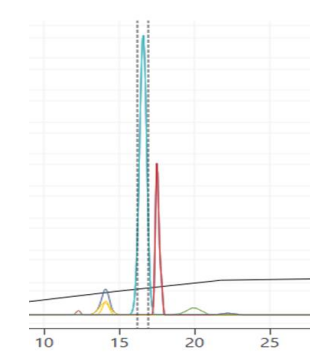
Stage 2 (20-52 min) A% gradient	50-85
---------------------------------	-------



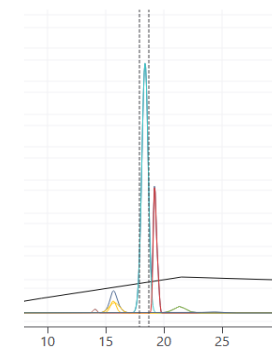
pH=5.5; 500mm NaCl
Condition1



pH=7.0; 500mm NaCl
Condition2



pH=7.0; 500mm NaCl
Condition3



pH=6.8; 300mm NaCl
Condition4

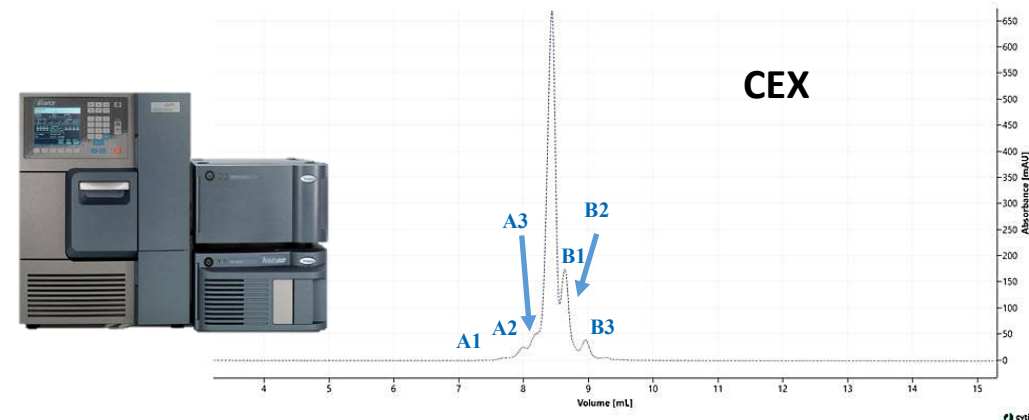


IEX (HPLC) Method Development –Validation

Model Calibration Input and Original Chromatography

Process parameters Parameters range

Buffer A pH	6.0-7.0
Buffer B pH	6.0-7.0
Buffer B NaCl conc. (mM)	300-600
Stage 1 (0-20 min) A% gradient	85-95
Stage 2 (20-52 min) A% gradient	50-85



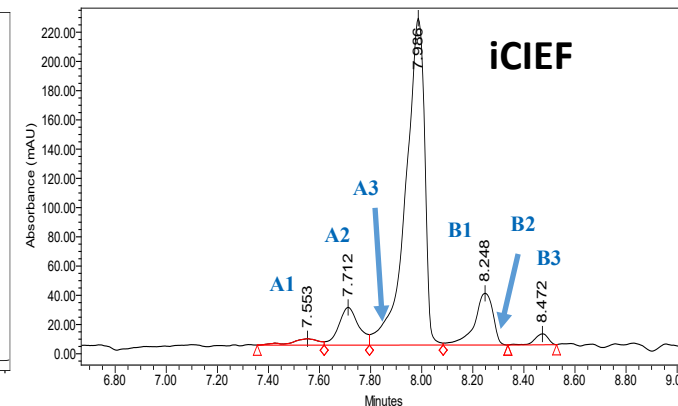
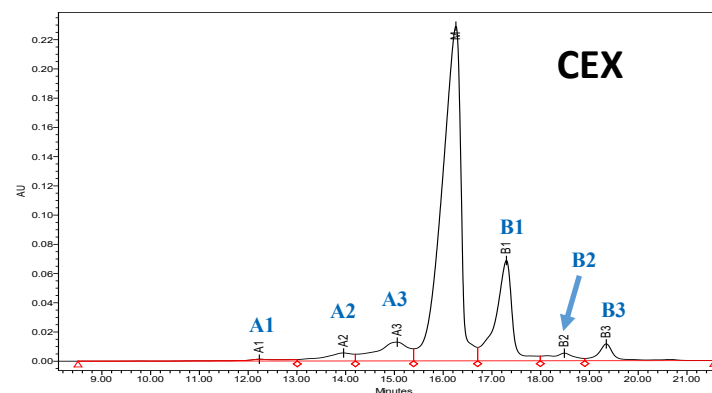
Model Recommended Process Parameters and Chromatography of Validation Run

Buffer A pH=7.0

Buffer B 500 mM NaCl, pH=7.0

Inject amount 100 µg

Stage	Time	Flow rate (mL/min)	A%	B%
1	0	1	100	0
2	20	1	90	10
3	52	1	88	12



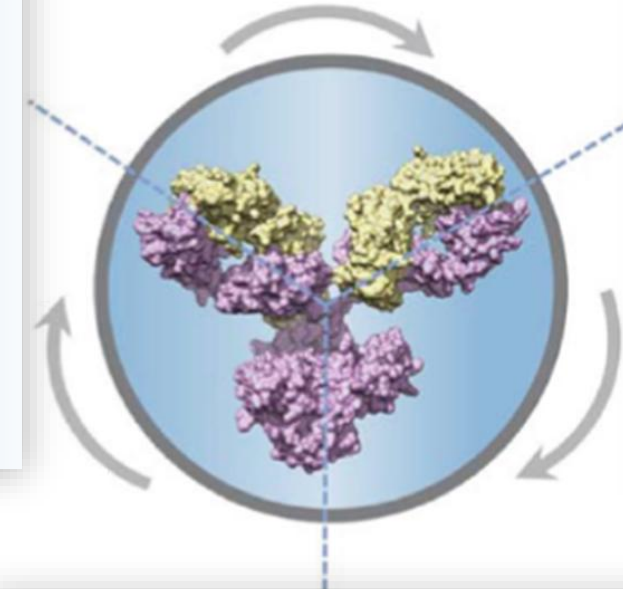
Comprehensive Structure & Function Characterization After Fractionation

Structural Attributes :

- *Intact Mass*
- *Peptide Mapping*
- *Disulfide Mapping*
- *N-/O-Glycan Analysis*

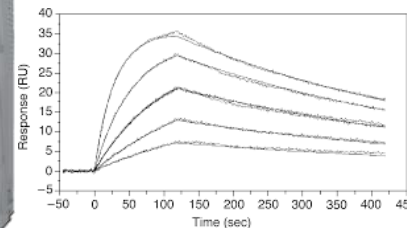
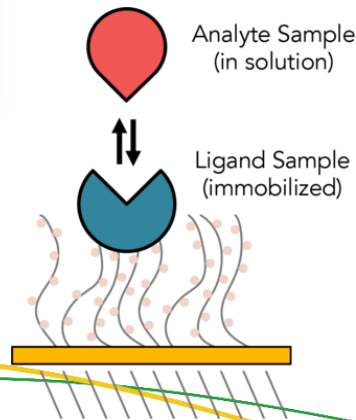
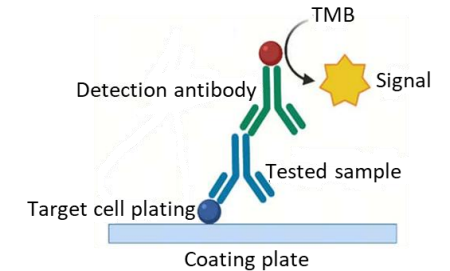
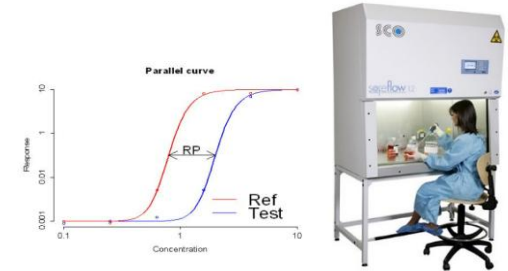
Functional Attributes:

- *Ag-Ab Binding*
- *Biological Activity*
 - *Cell cytotoxicity*
 - *Neutralization*
 - *Cytokine release*
 - *Blockade bioassay*



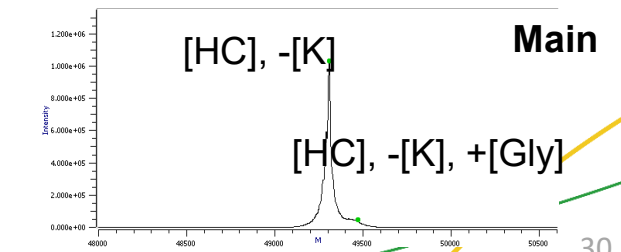
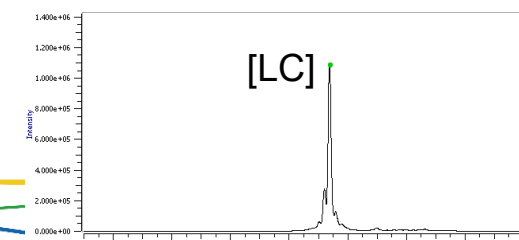
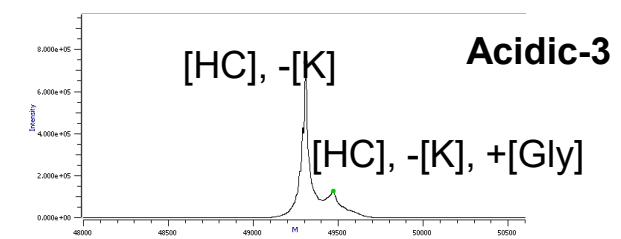
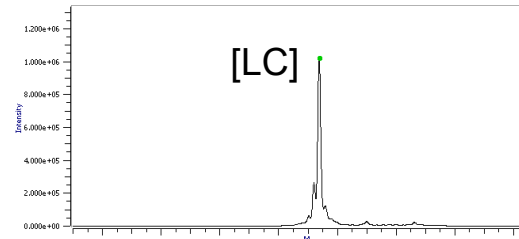
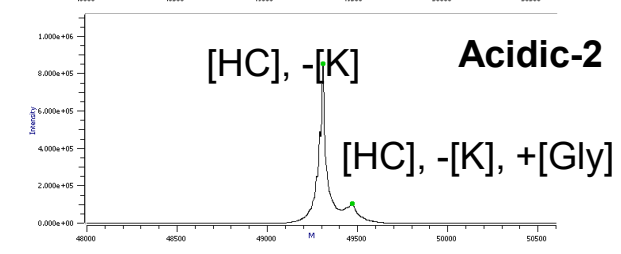
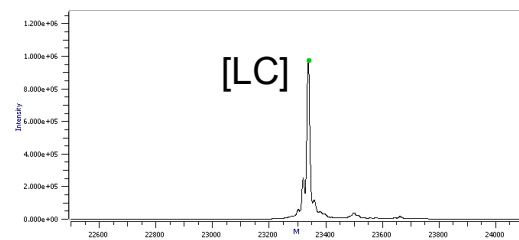
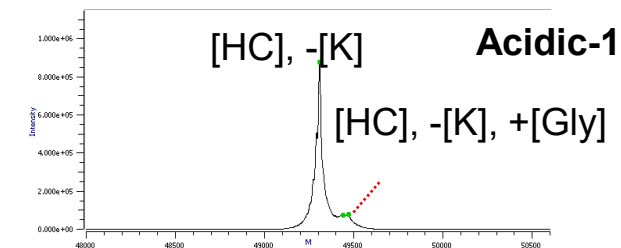
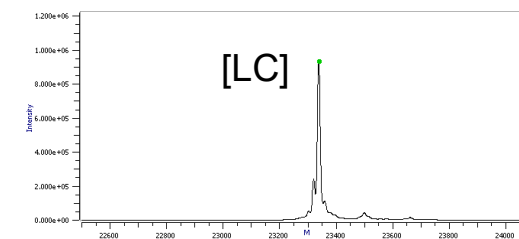
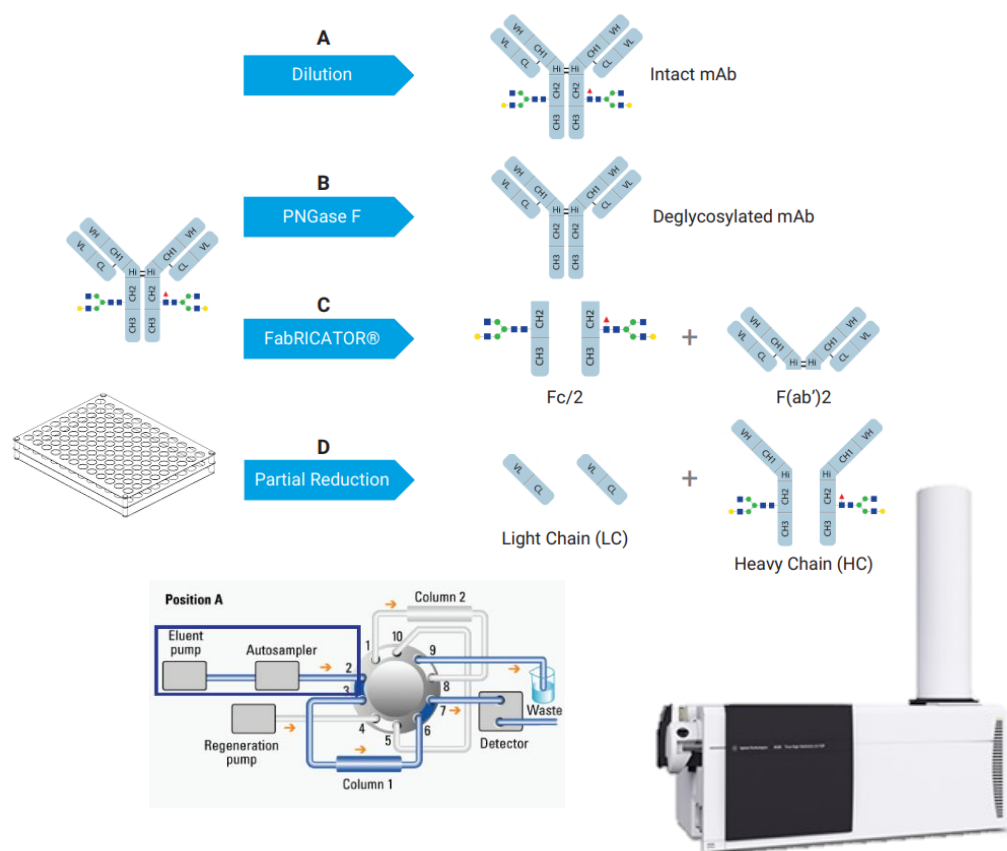
Analysis of effector functions:

- *ADCC, ADCP, CDC, FcRn*



In-depth Charge Variant Characterization After Fractionation -De-glycosylated Reduced Intact Mass

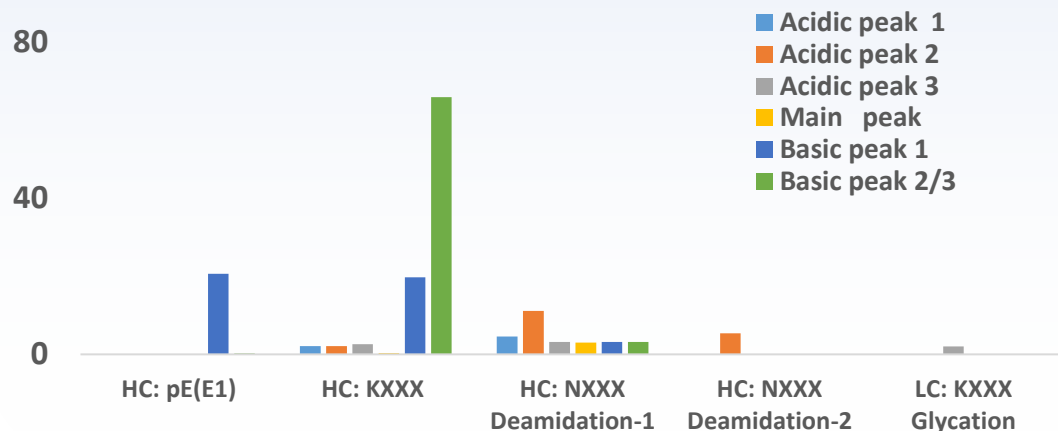
From the results of De-glycosylated Reduced Mass, higher abundance of **glycation** was identified in heavy chain of **Acidic Peak 2 and 3**.



In-depth Charge Variant Characterization After Fractionation

-Peptide Mapping and Glycan Analysis

Peptide Mapping for CEX Fractions



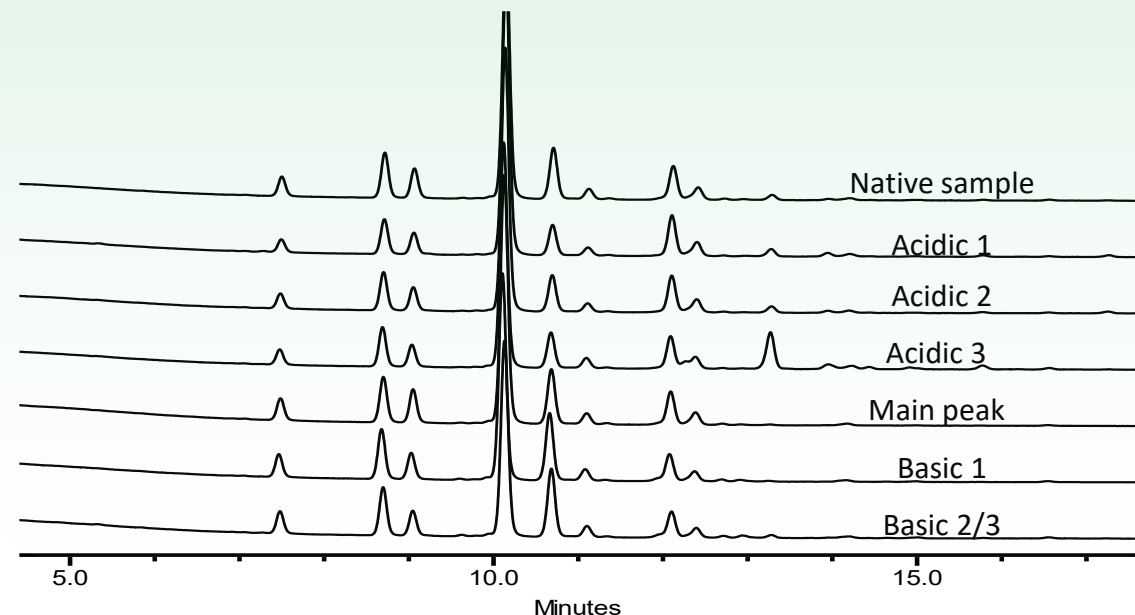
Source of acidic variants

- ☐ Confirm the higher deamidation site-1 NXXX and site-2 in Acidic peak 2.
- ☐ One glycation hotspot in Acidic peak 3

Source of basic variants

- ☐ Confirm lower ratio of glutamine cyclization-pE (Q1)
- ☐ K450 loss in Basic Peak 1 and 2/3.

N-glycan Profile for CEX Fractions

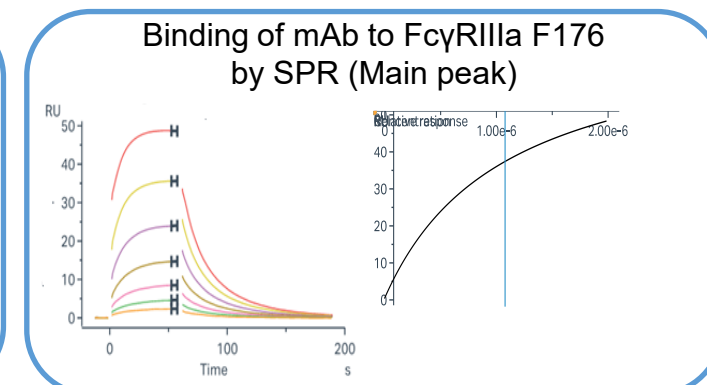
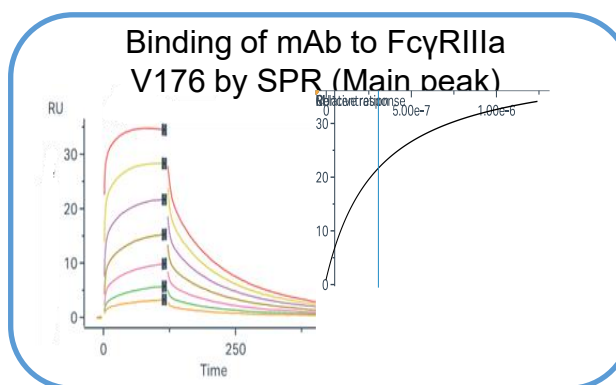
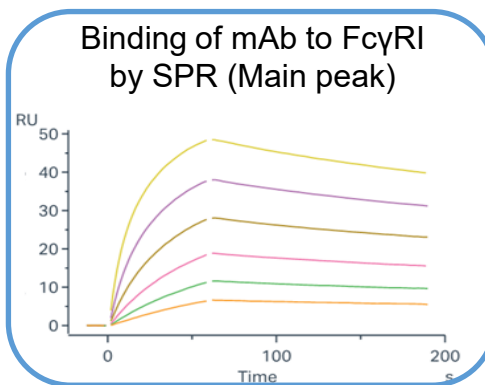
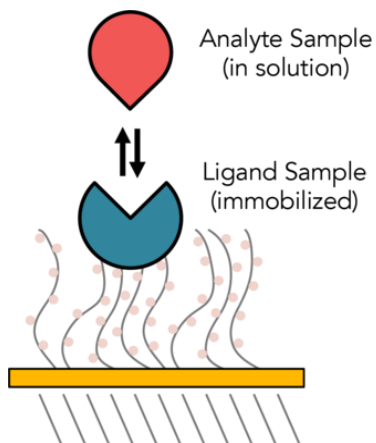


Source of acidic variants

- ☐ Higher sialic acid level (G1FS1-GN) in Acidic peak 3

In-depth Charge Variant Characterization After Fractionation -Effector Function by SPR

Compared with main peak, acidic peaks and basic peaks (fraction Acidic peak 1/2/3, Basic peak 1/2/3) showed comparable binding affinity to different human FcγR receptors



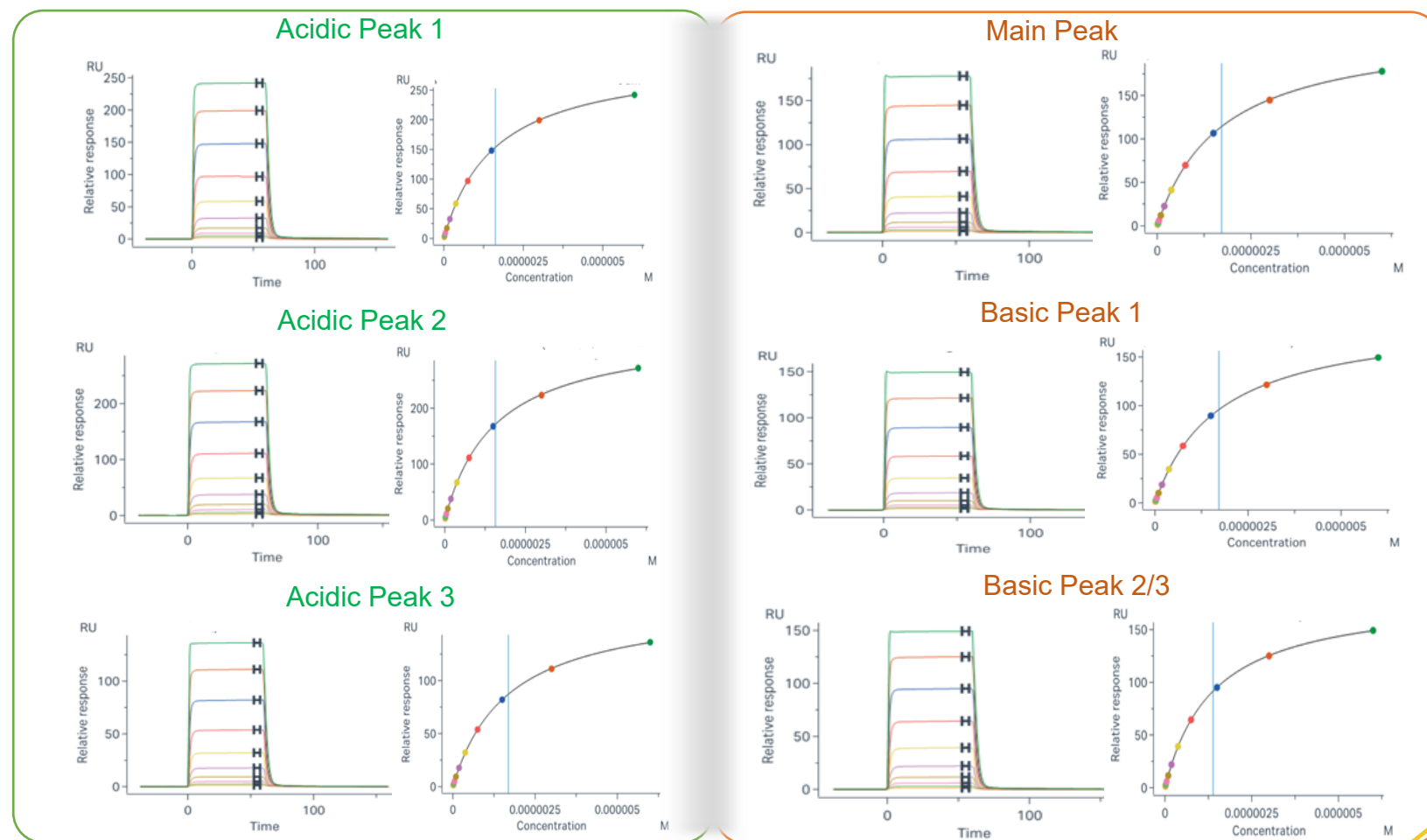
KD (M)	FcγRI	FcγRIIIa R167	FcγRIIIa H167	FcγRIIIa V176	FcγRIIIa F176
Acidic peak 1	3.5E-09 M	6.8E-06 M	2.0E-06 M	3.0E-06 M	1.1E-06 M
Acidic peak 2	3.6E-09 M	6.9E-06 M	1.8E-06 M	3.2E-06 M	1.2E-06 M
Acidic peak 3	3.2E-09 M	7.0E-06 M	1.9E-06 M	3.3E-06 M	1.0E-06 M
Main peak	3.8 E-09 M	6.9E-06 M	1.9E-06 M	3.1E-07 M	1.1E-06 M
Basic peak 1	3.7E-09 M	6.9E-06 M	2.1E-06 M	2.9E-07 M	1.2E-06 M
Basic peak 2/3	3.9E-09 M	7.3E-06 M	1.7E-06 M	3.6E-07 M	1.3E-06 M

In-depth Charge Variant Characterization After Fractionation

-Effector Function by SPR

Compared with main peak, acidic peaks and basic peaks (fraction Acidic peak 1/2/3, Basic peak 1/2/3) showed comparable binding affinity to human FcRn.

FcRn by SPR	KD (M)
Acidic peak 1	1.6E-06
Acidic peak 2	1.6E-06
Acidic peak 3	1.7E-06
Main peak	1.7E-06
Basic peak 1	1.7E-06
Basic peak 2/3	1.7E-06





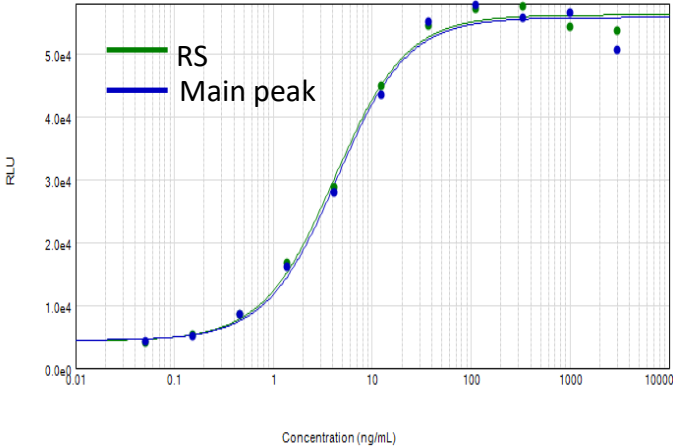
In-depth Charge Variant Characterization After Fractionation

-Effector Function by Cell-based Bioassay

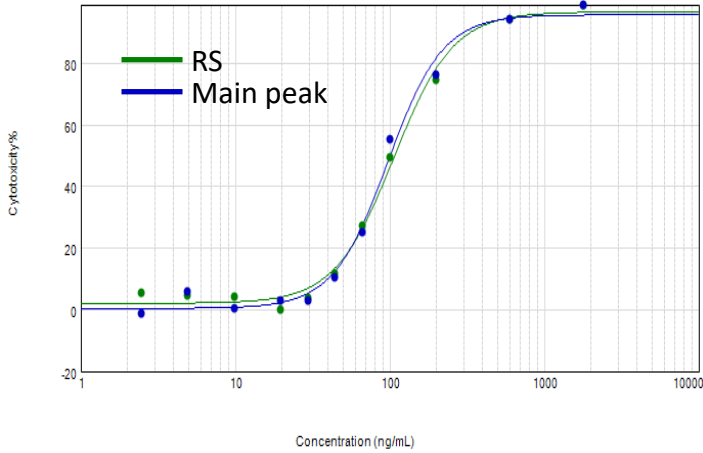
Compared with main peak, acidic peaks and basic peaks (fraction Acidic peak 1/2/3, Basic peak 1/2/3) showed comparable activity in cell based ADCC, CDC, FcRn binding assay

Relative potency	ADCC	CDC	FcRn Binding
Acidic peak 1	92%	99%	107%
Acidic peak 2	103%	106%	102%
Acidic peak 3	89%	101%	108%
Main peak	96%	104%	103%
Basic peak 1	98%	103%	104%
Basic peak 2/3	103%	103%	102%

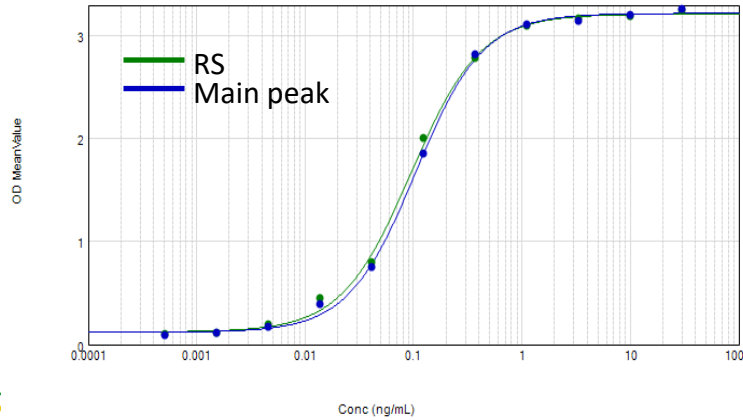
ADCC (Main peak)



CDC (Main peak)



Cell-based Competitive Binding Assay for FcRn (Main peak)





Summary *04*

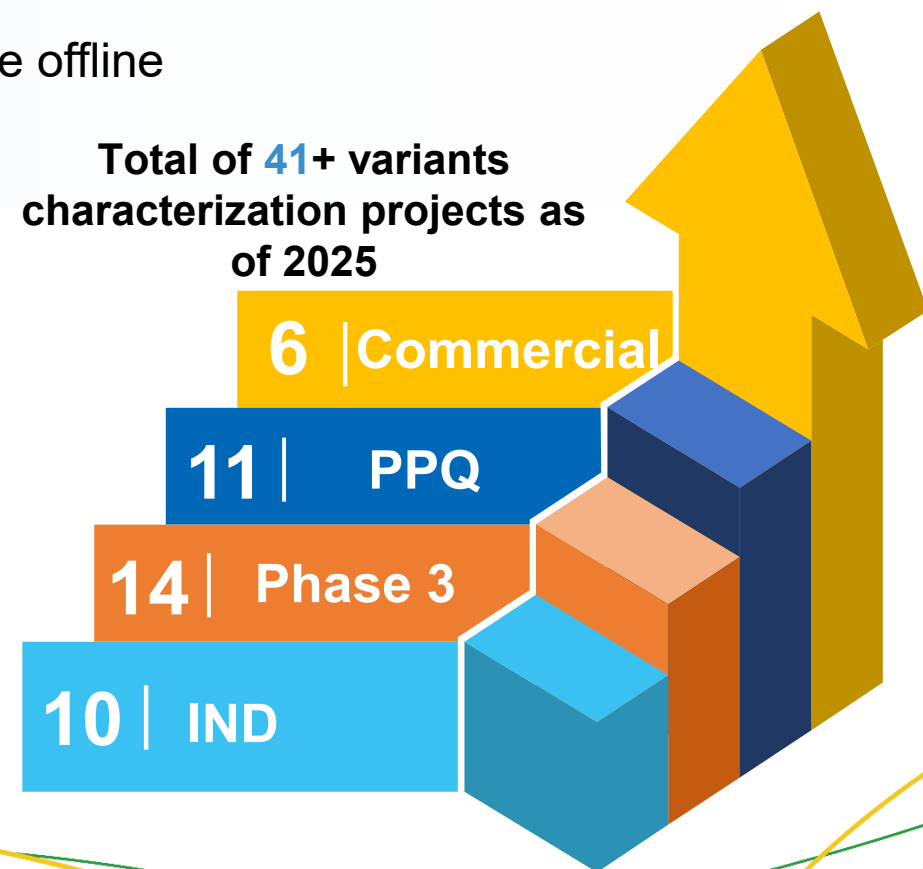
Summary

cIEF-MS has become essential tool for charge variant characterization, which has been routinely applied for analyzing different modalities including monoclonal antibody, bispecific and fusion proteins.

The 'Discovery' assisted approach significantly accelerated the offline fractionation workflow. Generally, 2–3 mg within 2 weeks.



Total of **41+** variants
characterization projects as
of 2025



Wuxi Biologics

- BioDev Wuxi Process Development

CMP Scientific

- James Xia

Agilent technologies

- Clara Chen (China team)
- Jun Li (China team)

Cytiva GoSilico™

- Dong Zhang
- Yijia Guo

CASSS Mass Spec Organizers and Session Chairs

Audiences and Attendees

Thank You