

Integrating Mass Spectrometry-Based Structural Proteomics and Molecular Docking to Analyze Ebola Protein-Host Binding Interfaces

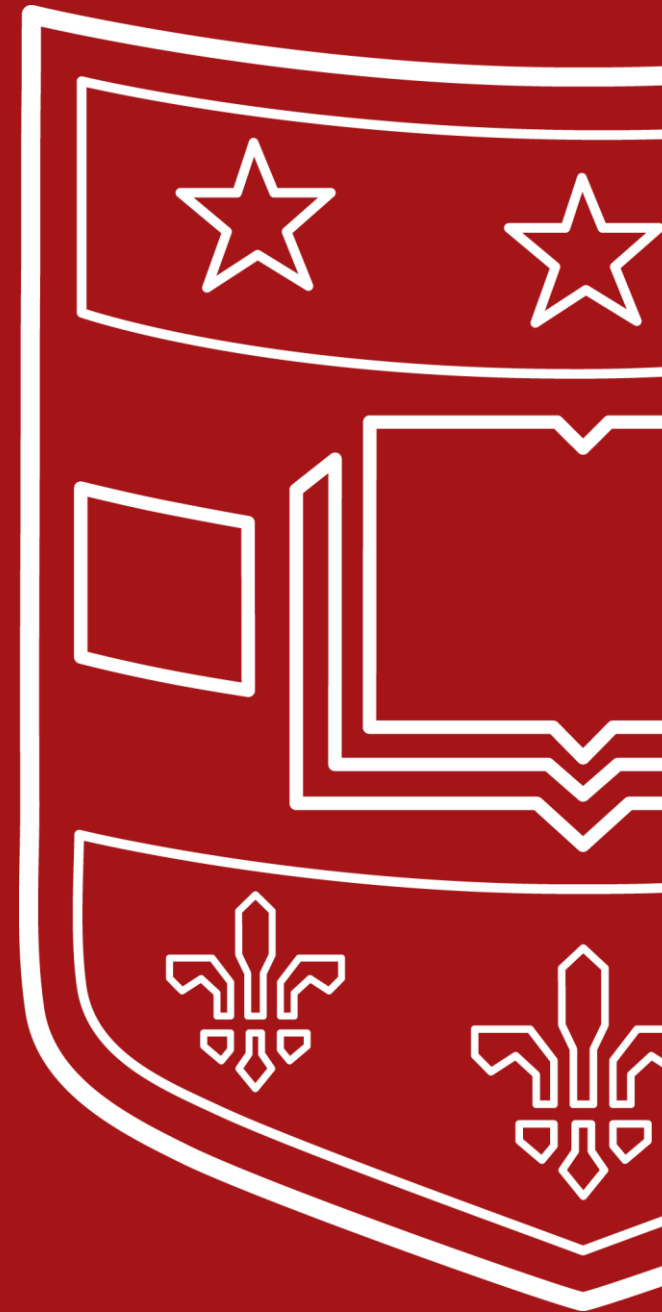
Xinyi (Cynthia) Kuang¹, Grace Uwase², Nicole D Wagner^{1,3}, Daisy W Leung³,
Gaya K Amarasinghe², Michael L Gross¹

CASSS Mass Spec 2025
Costa Mesa, CA

¹Dept of Chemistry, Washington University in St. Louis

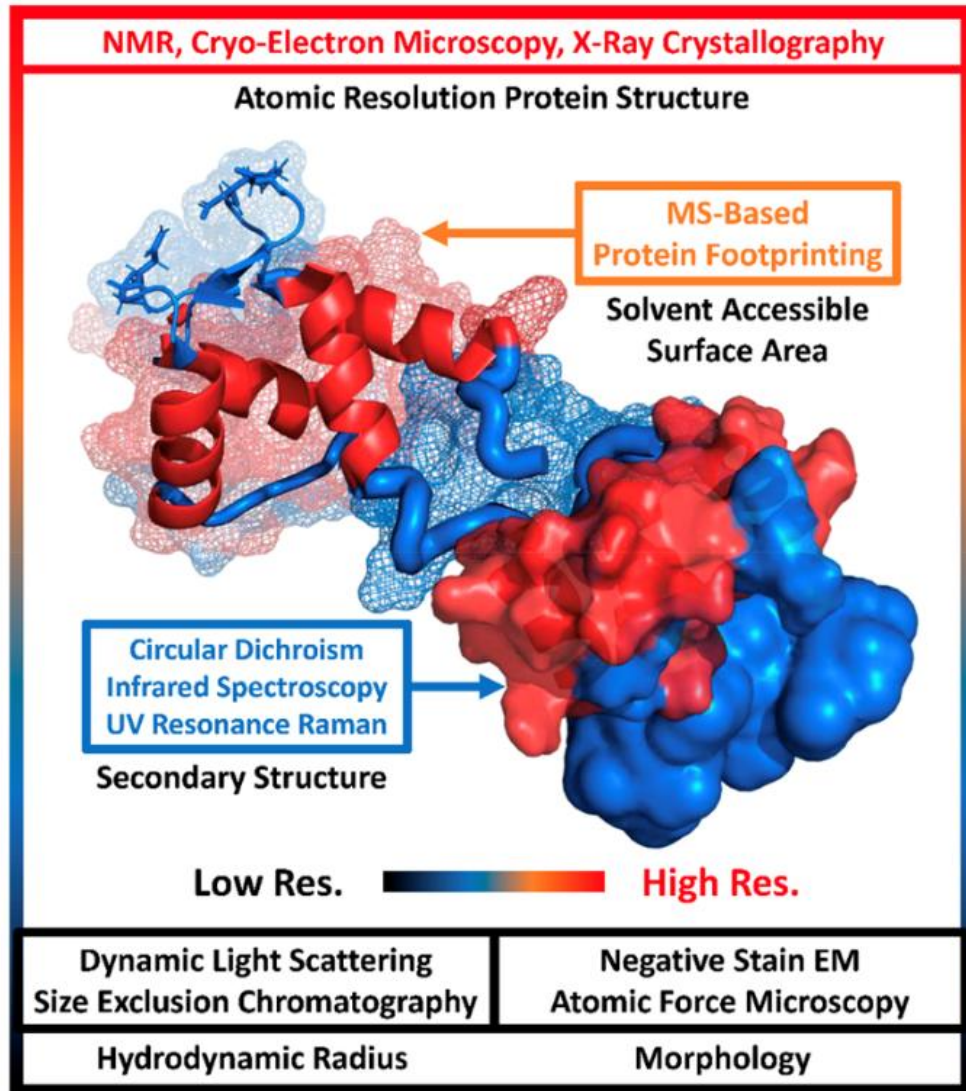
²Dept of Pathology and Immunology, Washington University School of Medicine in St. Louis

³Dept of Internal Medicine, Washington University School of Medicine in St. Louis





Biophysical Tools in Protein Higher Order Structure (HOS)



➤ Protein Higher Order Structure

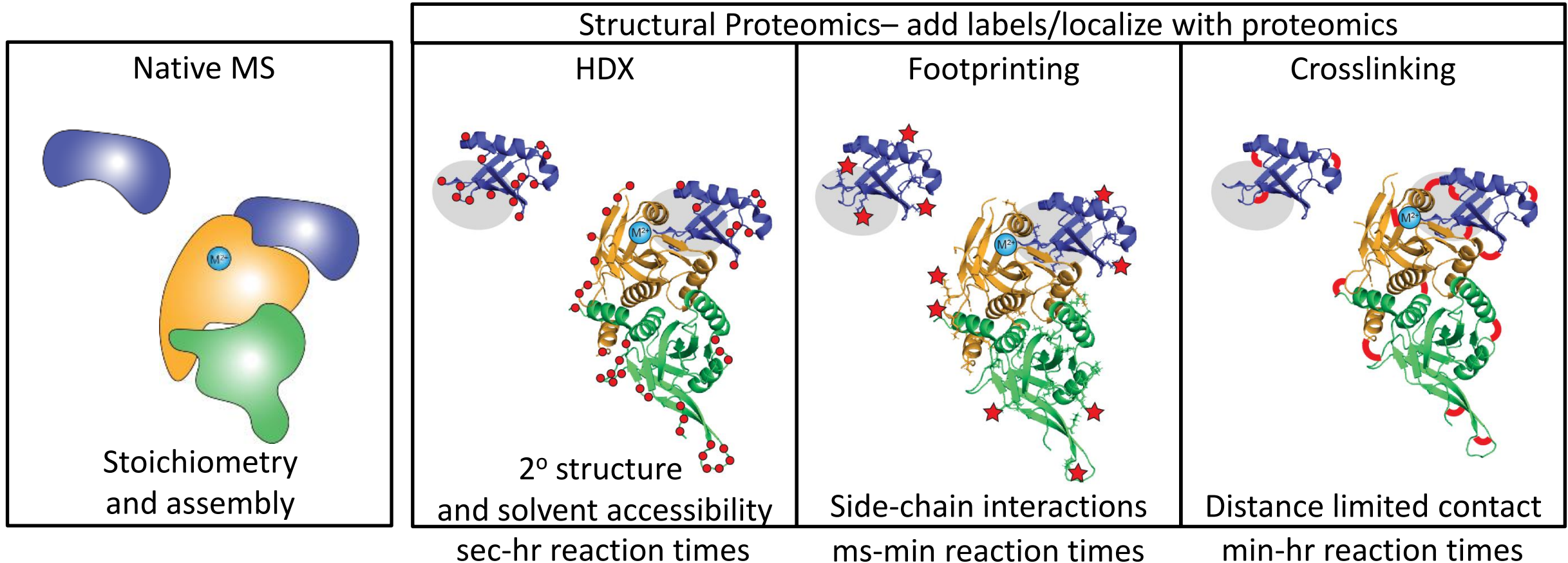
- Secondary to quaternary
- Protein biological function, stability and molecular interactions

➤ Mass Spectrometry-Based Approaches

- Fast and sensitive
- Real-time characterization in solution (under physiological conditions)
- High throughput capability
- Simple sample preparation; requires only ng-μg amounts
- High spatial resolution (peptides & residues)
- ...



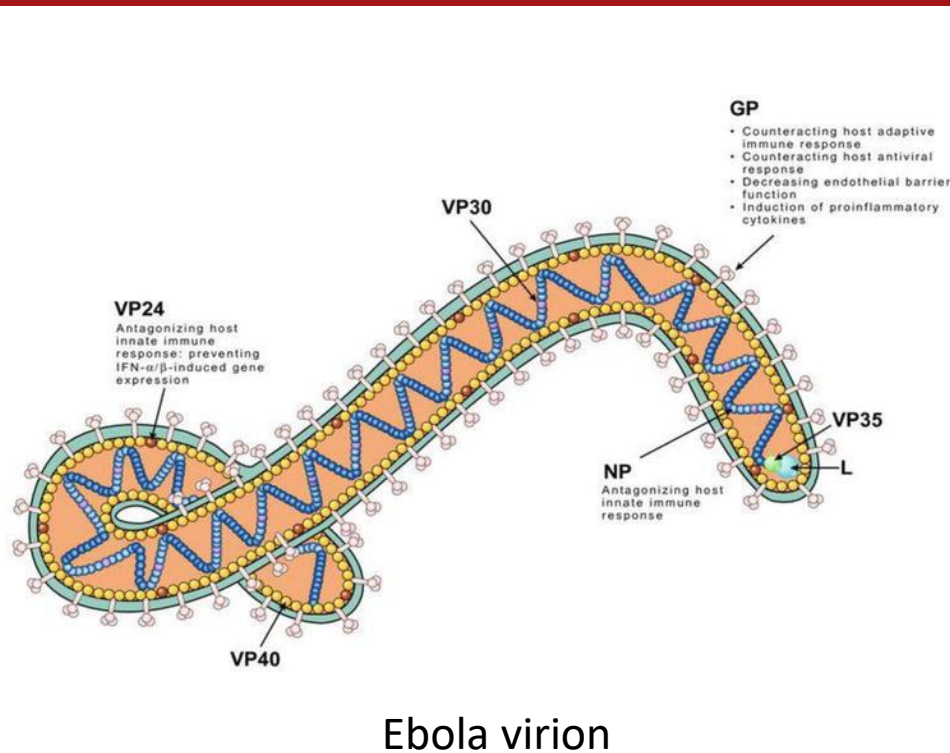
Our suite of mass spectrometry approaches for HOS characterization



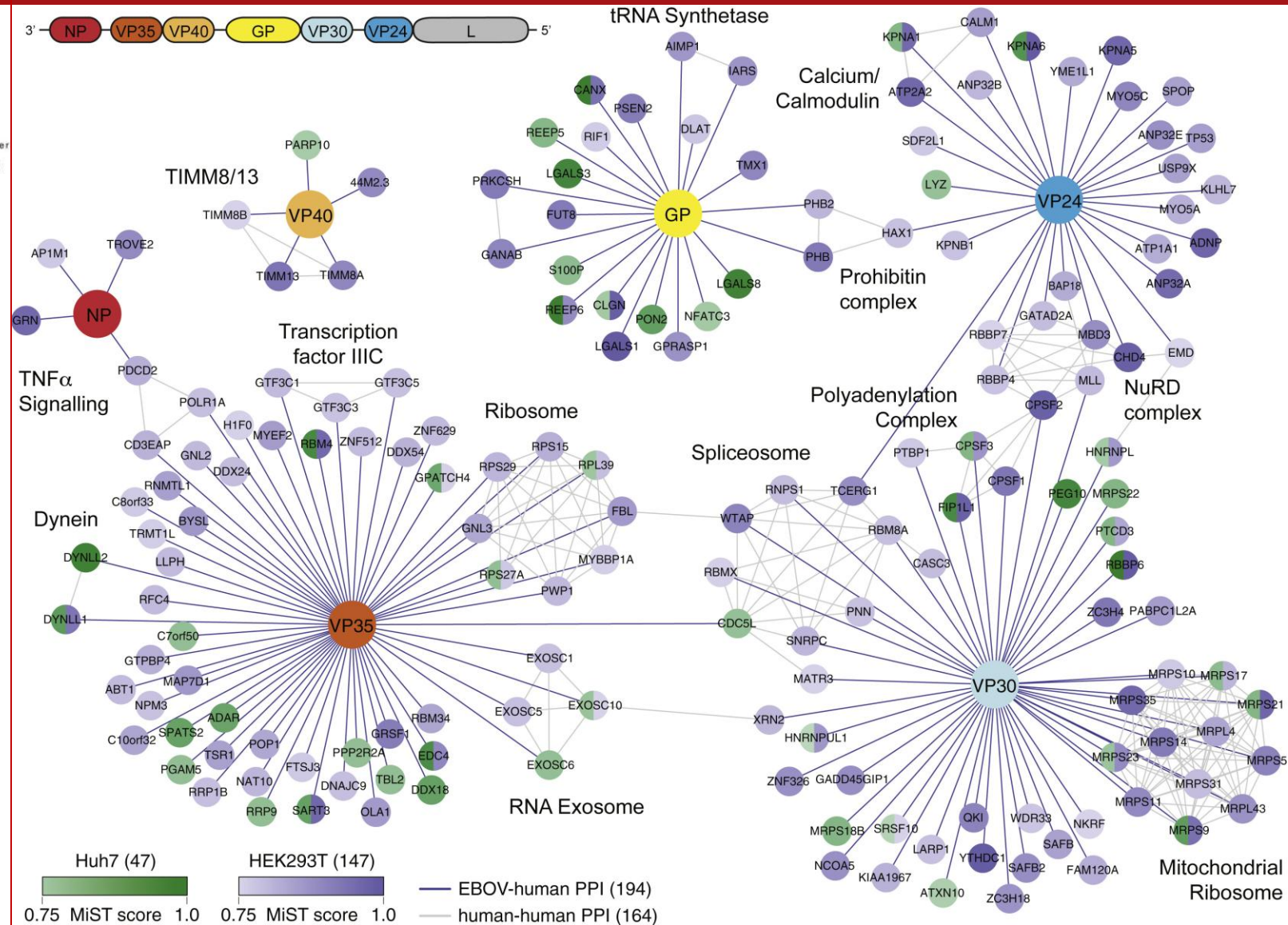
A comprehensive characterization platform is necessary for protein HOS analysis



The biological question we are interested: Ebola Protein-Host E3 ligase



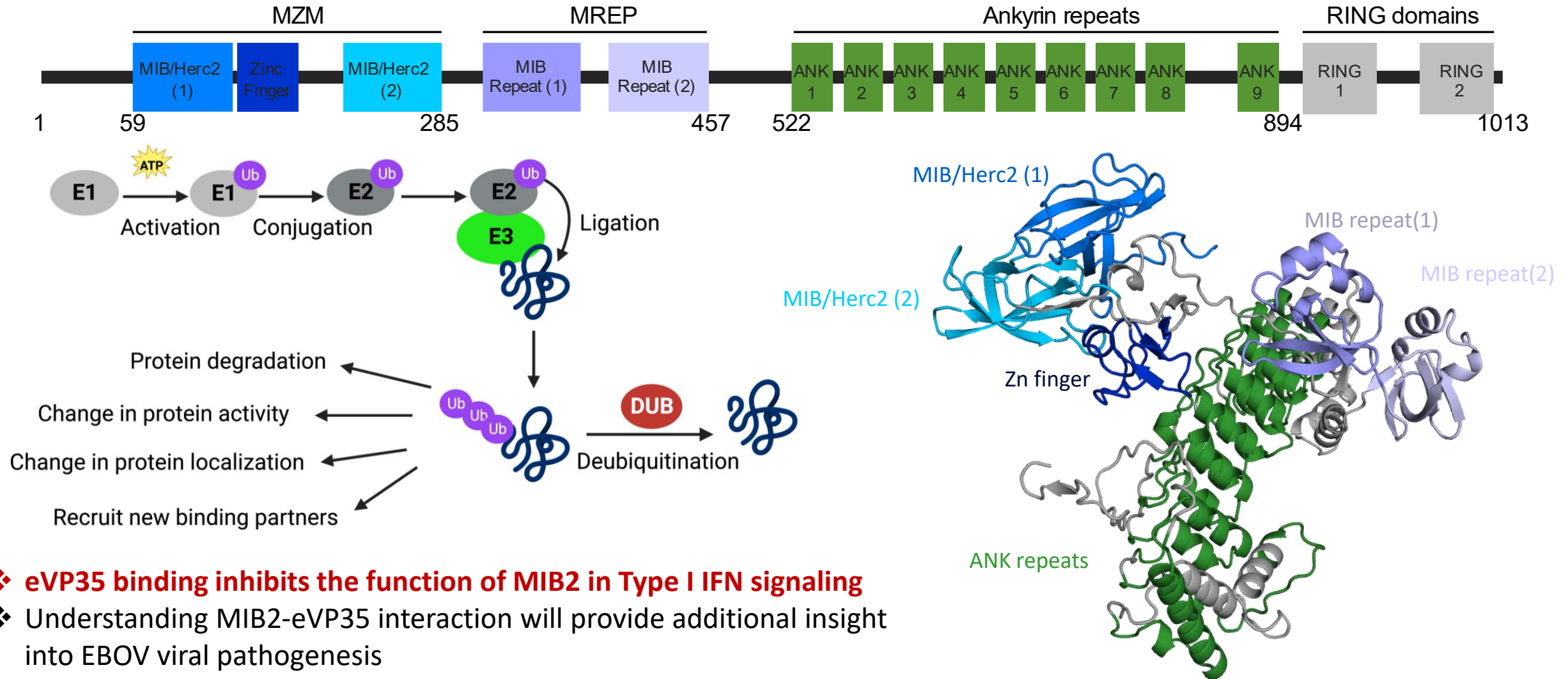
- **eVP35**: multifunctional cofactor essential for viral replication and immune evasion
- **eVP35-E3 ubiquitin ligase MIB2 interaction**





Host E3 ubiquitin ligase: MIB2 (Mindbomb protein 2)

MIB2: Mindbomb protein 2, E3 ubiquitin ligase

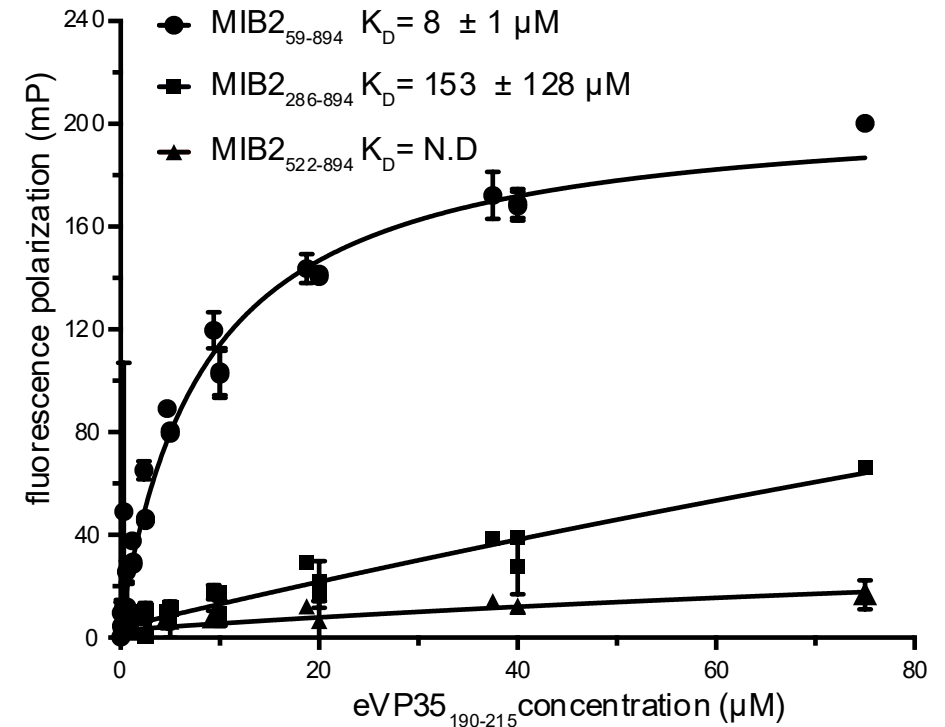


- ❖ **eVP35 binding inhibits the function of MIB2 in Type I IFN signaling**
- ❖ Understanding MIB2-eVP35 interaction will provide additional insight into EBOV viral pathogenesis



Platform: Binding interaction between eVP35-MIB2

	MIB2 construct	Binds eVP35?
	MZM MREP ANK RING	
1-1013		+++
59-894		+++
286-894		+
522-894		-



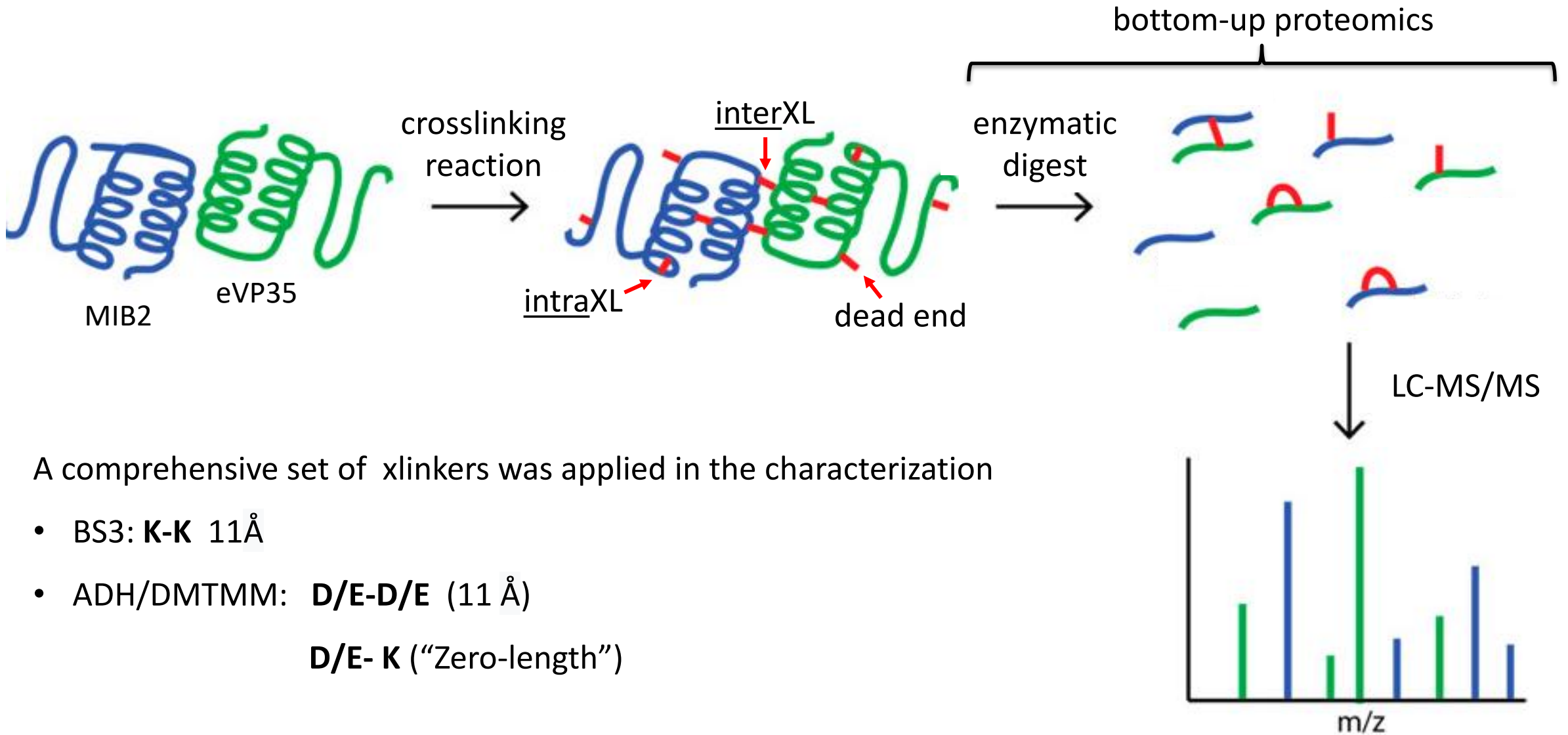
- Truncation experiments characterized MIB2 **MZM** and **MREP** domains are critical for VP35 binding
 - MIB2₅₉₋₈₉₄ construct was selected
- Conformational binding interfaces? Binding induced structural changes?**

Proposed experiment

HDX-MS and XL- MS to localize the eVP35 binding sites on MIB2



Crosslinking (XL)-MS identifies distance-limited proximity

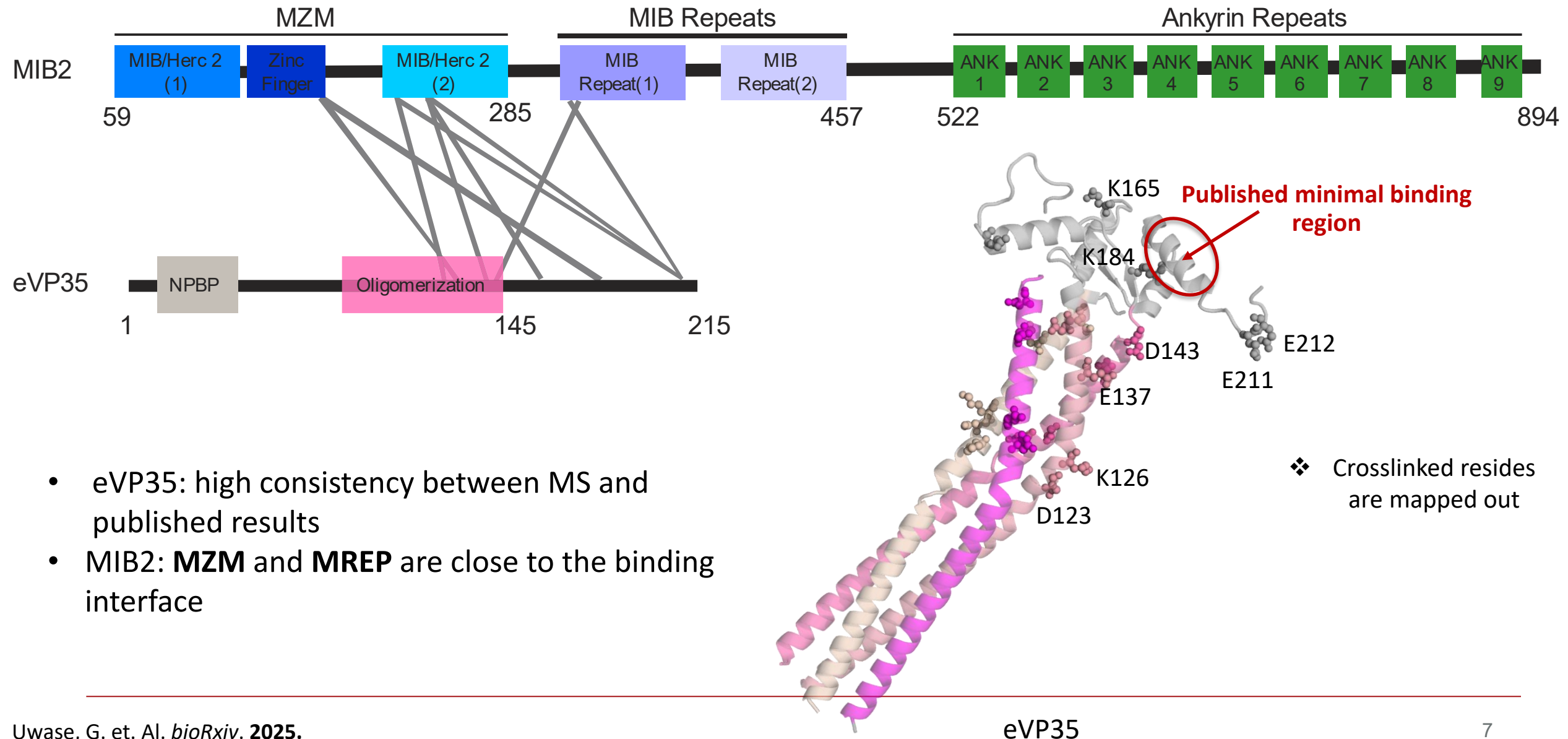


A comprehensive set of xlinkers was applied in the characterization

- BS3: **K-K** 11Å
- ADH/DMTMM: **D/E-D/E** (11 Å)
D/E- K ("Zero-length")

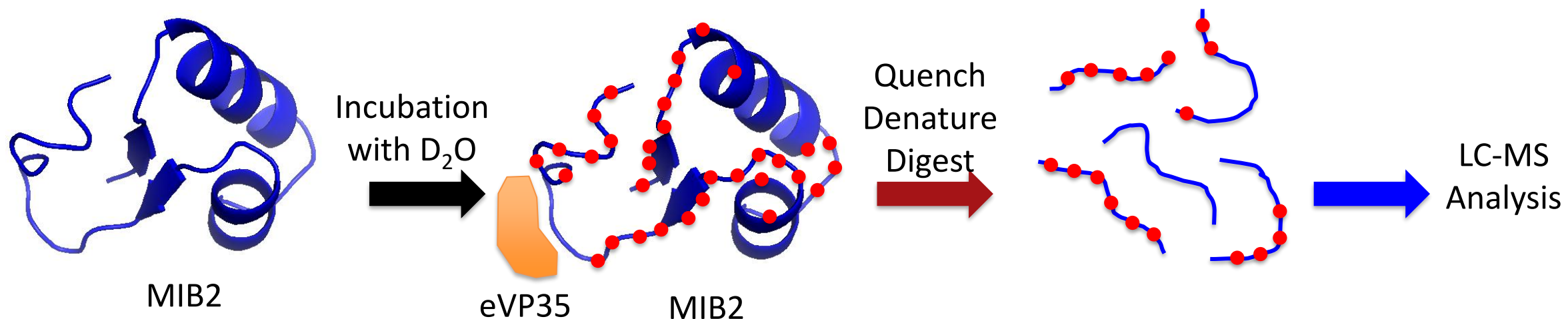


MIB2 MZM and MREP domains crosslink to eVP35 near the identified minimal binding site

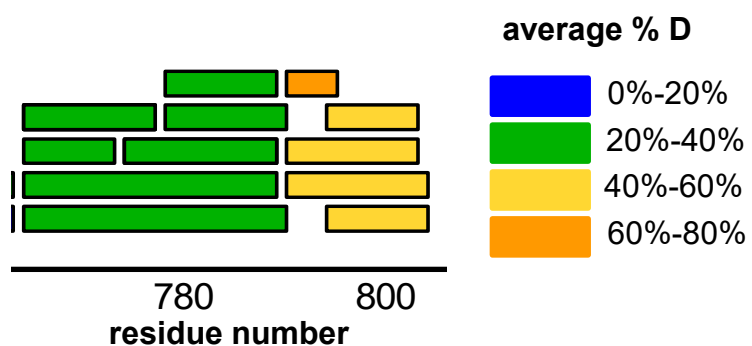




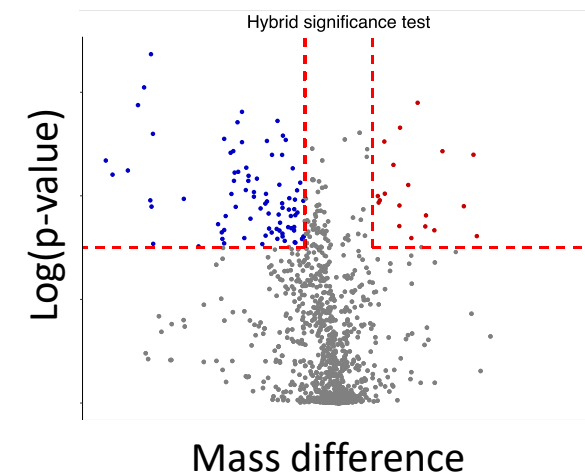
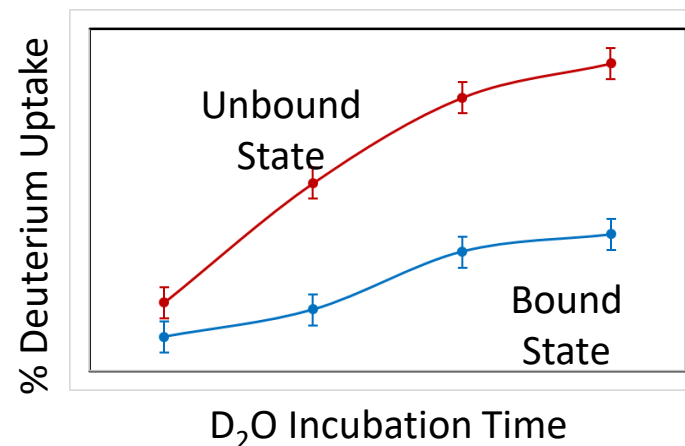
HDX-MS provides insight into second structure and solvent accessibility



Reports on regions of flexibility/dynamics

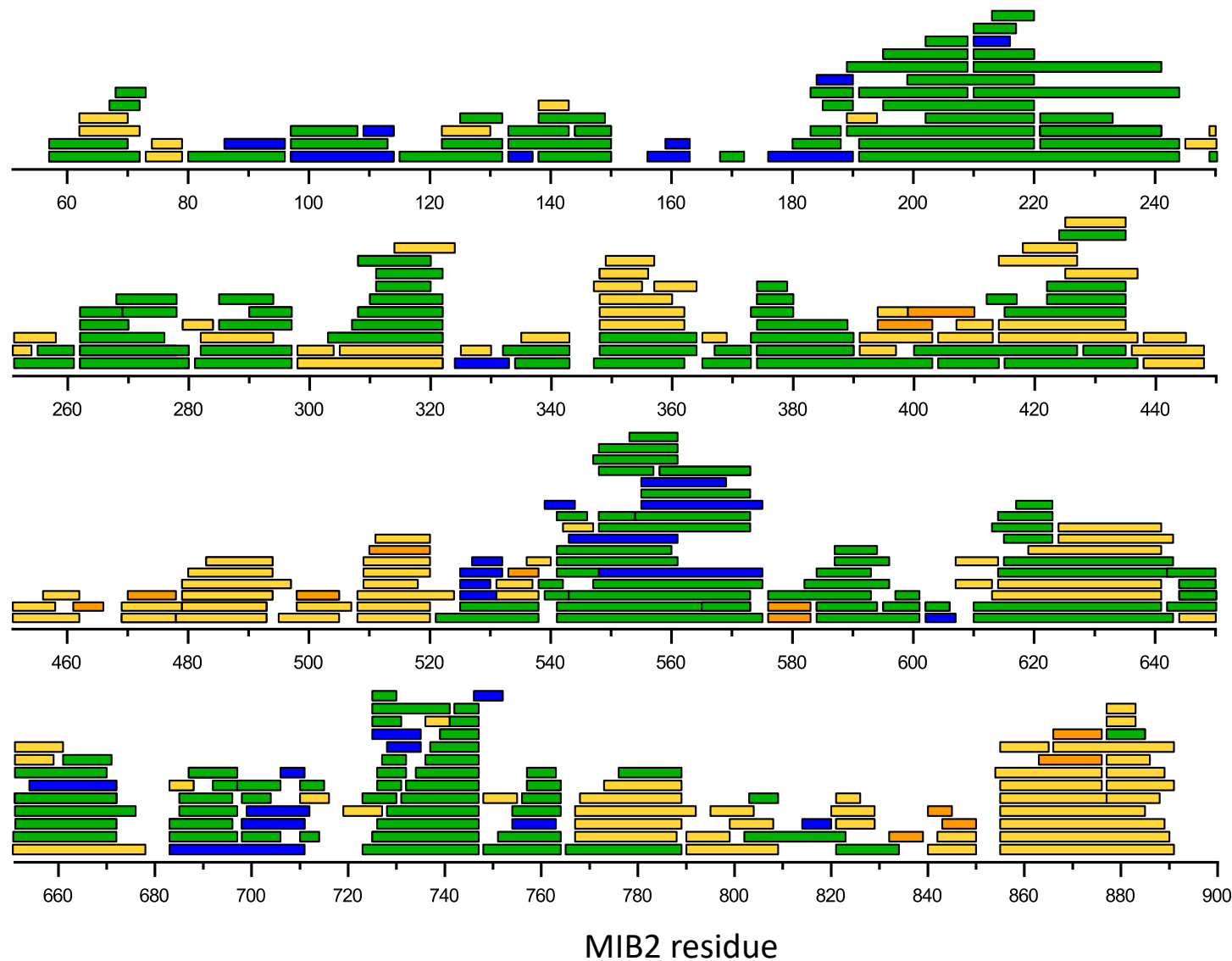


Identifies statistically significant differences





HDX-MS reveals regions of stability and flexibility within MIB2



- Digestion: Fungal XIII-pepsin at 4C
- 376 peptides
- 96% coverage
- Redundancy: 6
- Avg peptide length: 12.5



average % D for all time points

0%-20%

20%-40%

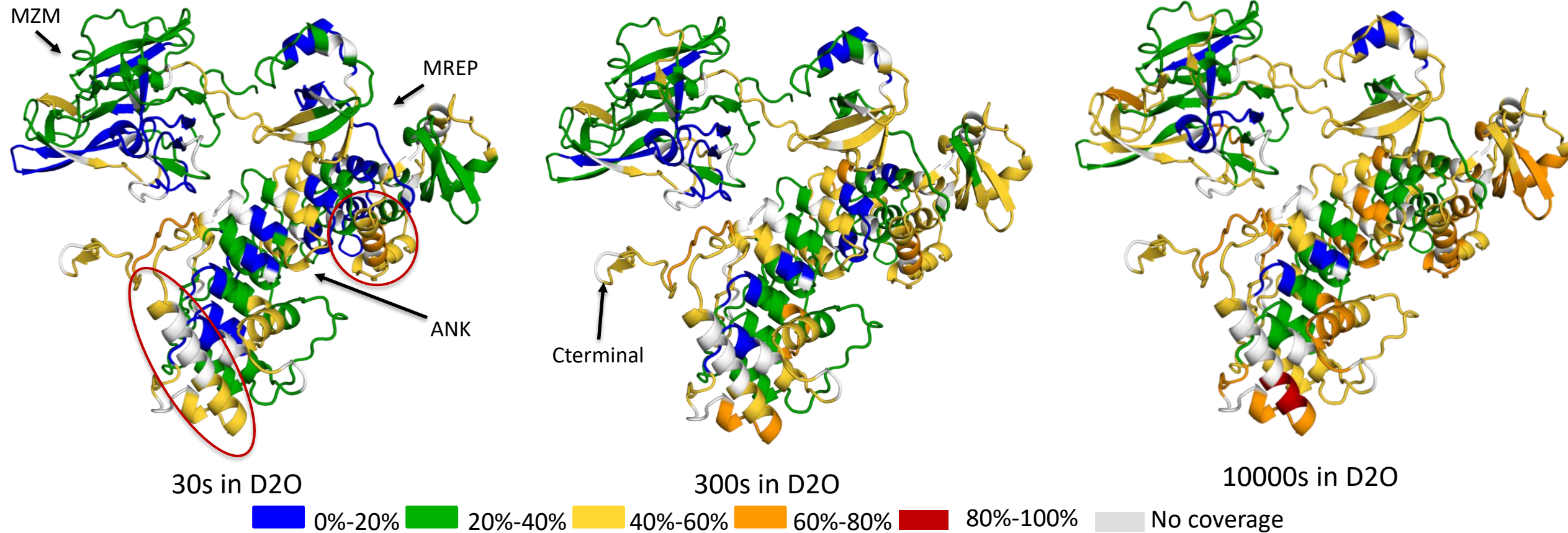
40%-60%

60%-80%

No coverage



HDX-MS reveals MIB2 regions of stability and flexibility within MIB2



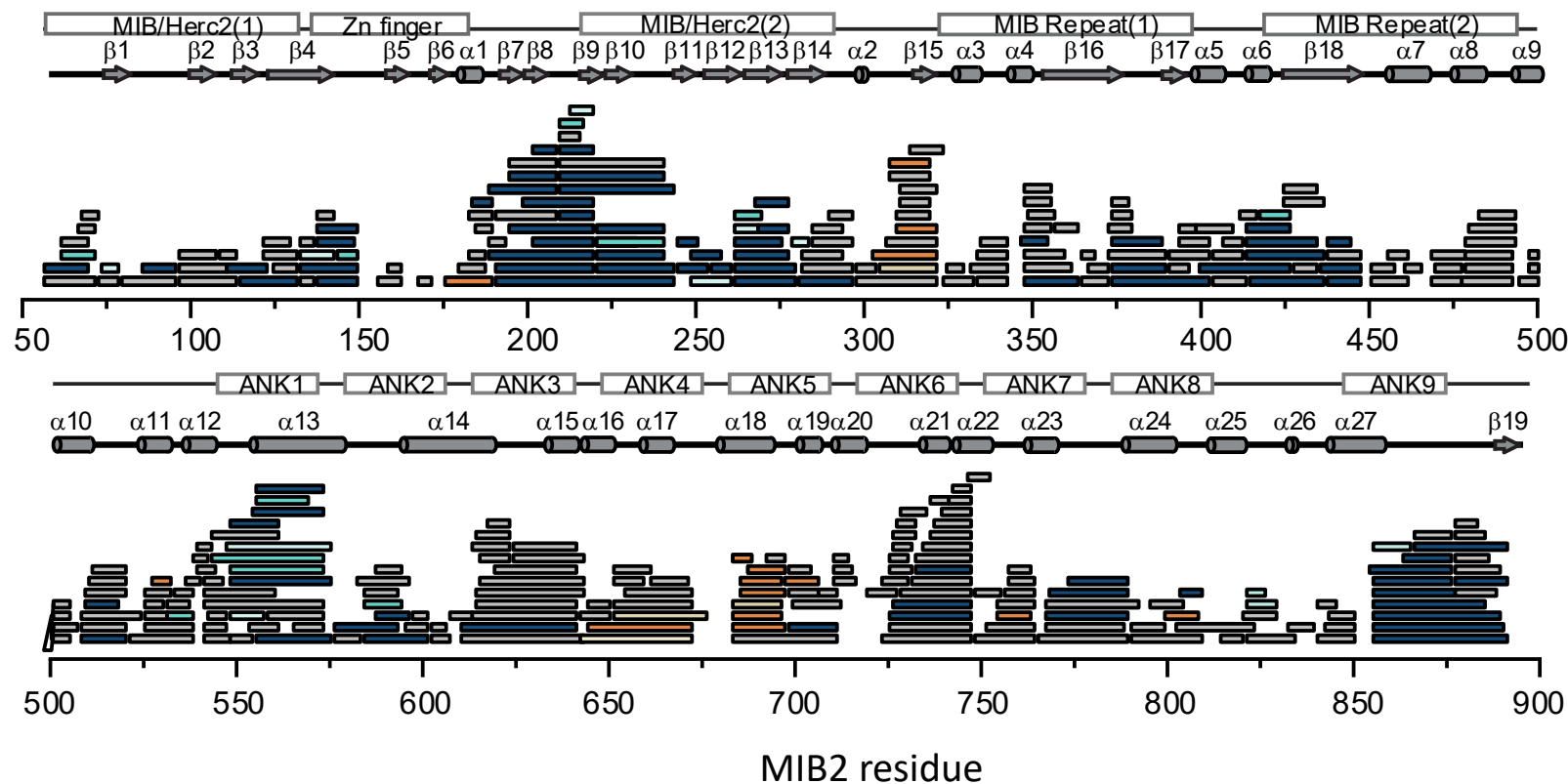
- MZM exhibits relative stability
- Some of the long helices predicted by AlphaFold show > 40% deuterium uptake even at 30 sec of HDX
- HDX results complement the AlphaFold structure by revealing dynamic features of MIB2.



Differential HDX reveals potential eVP35 binding sites and remote conformational changes of MIB2

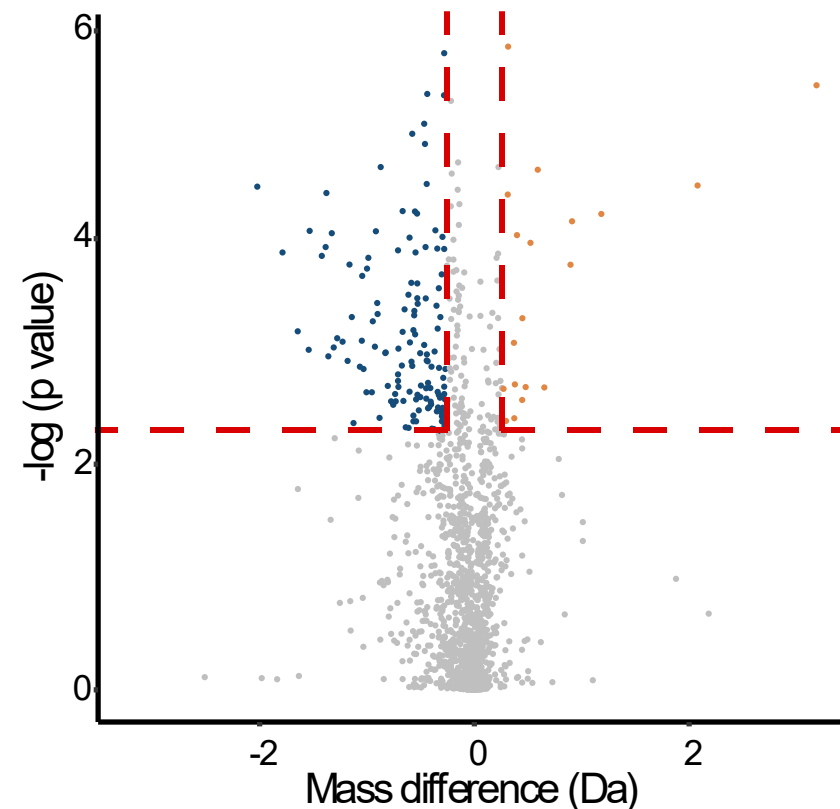
Differential HDX between MIB2 +/- eVP35

Differential HDX peptide map



decreased HDX ■ P < 0.005 ■ P < 0.01 ■ P < 0.05 • Decreased HDX: binding sites or more hydrogen bonds
increased HDX ■ P < 0.005 ■ P < 0.01 ■ P < 0.05 • Increased HDX: gain flexibility or dynamics
non-significant ■

Volcano plot

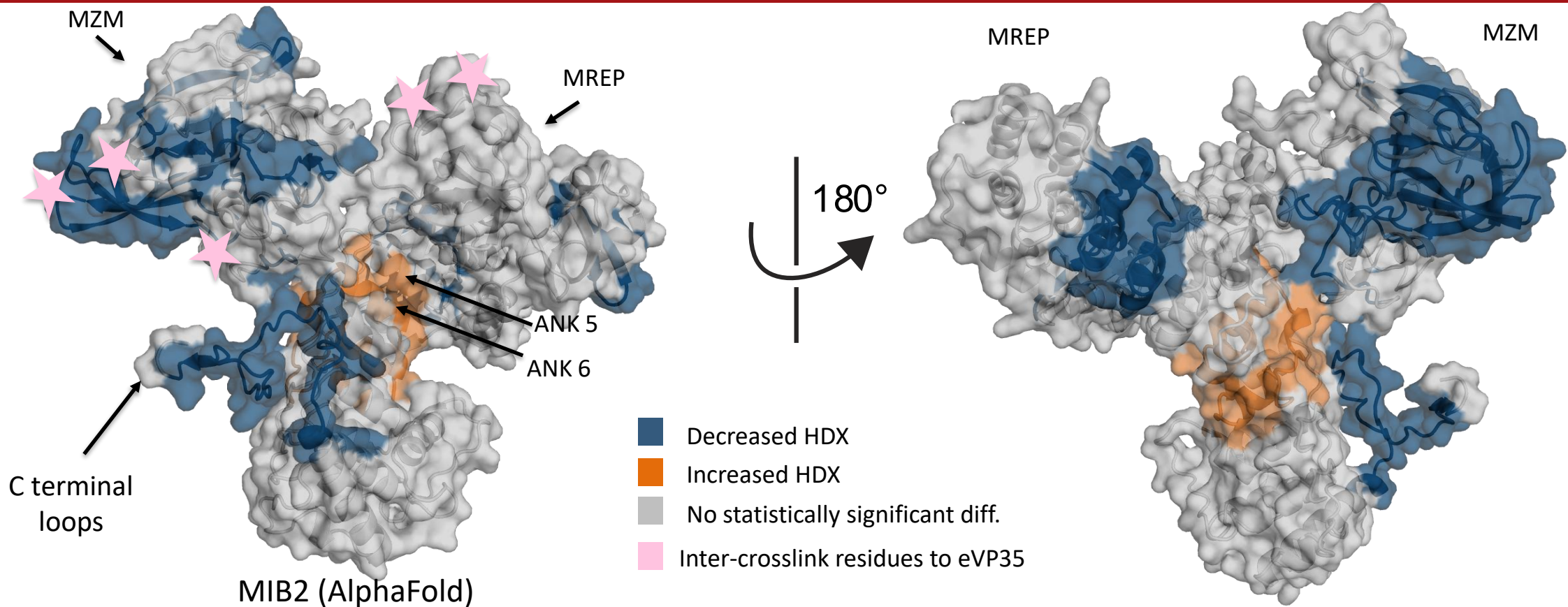


P-value cutoff: 0.005

Global significance limit: 0.25 Da



Differential HDX mapped onto an AlphaFold structure reveals discontinuous interaction surface



- Protected regions form a **discontinuous surface** (consistent with interXL)

Possibilities:

(i) binding induced conformational changes?

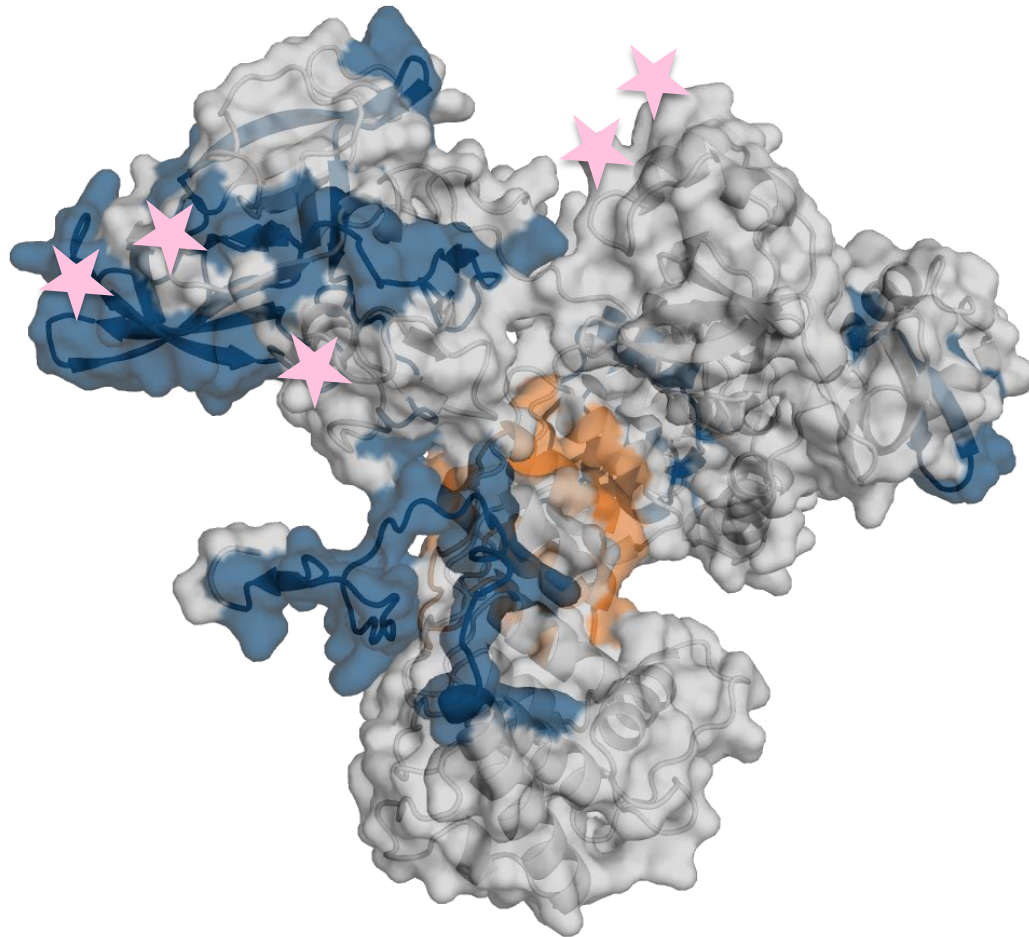
(ii) Or this structure is inconsistent in this context (especially domain-orientation)? (consistent with interXL)

(iii) MZM and MREP appear reoriented upon eVP35 binding?

- ANK 5-6 shows increased flexibility or dynamic motion upon binding



Conclusions and Questions remained

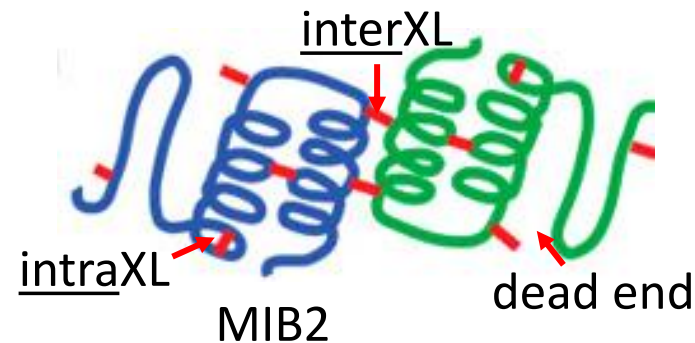


MIB2 (AlphaFold)

- Decreased HDX
- Increased HDX
- No statistically significant diff.
- Inter-crosslink residues to eVP35

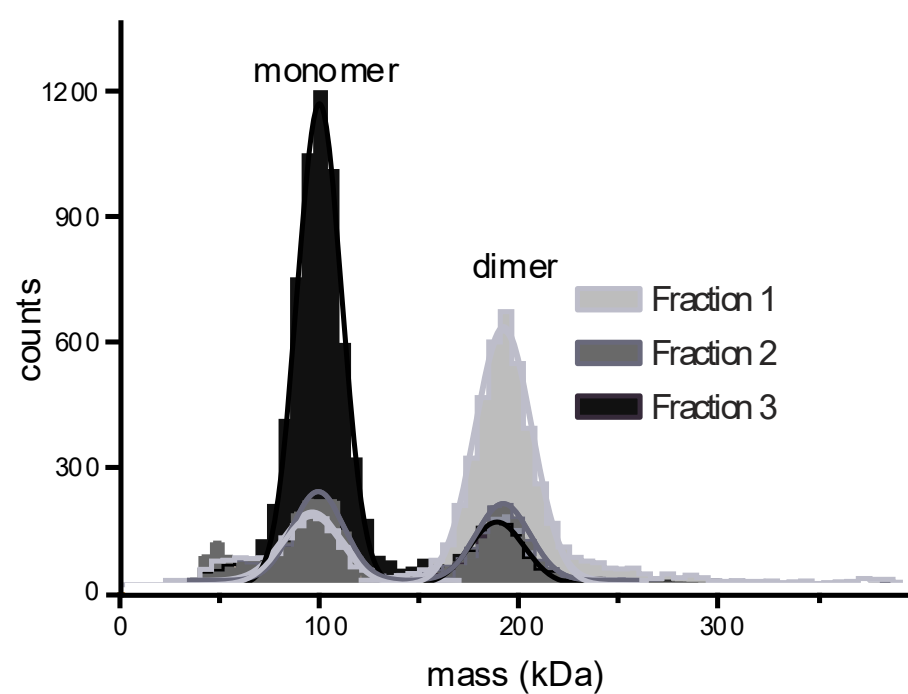
- HDX-MS and XL-MS results report a potential binding site but also additional structural perturbations
- eVP35 binds MIB2 MZM and MREP domains
- MS results complements AlphaFold, revealing dynamics and alternative domain orientations.
- Inspiration: MIB2 domains orientation?

intramolecular XLs within MIB2 inform on domains proximity

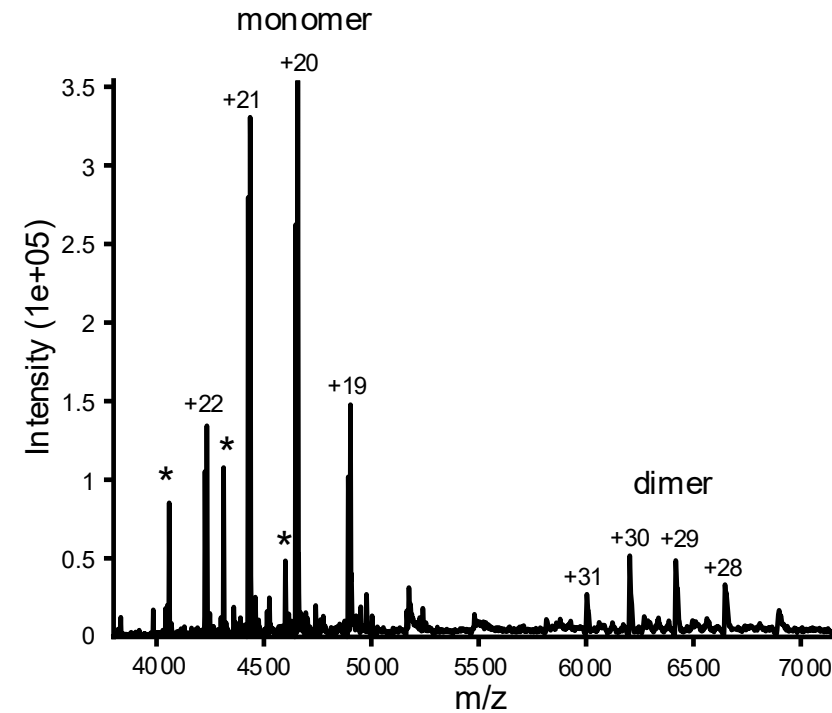




Challenge: MIB2 adopts monomeric and dimeric forms



Mass photometry



Native MS

Crosslink types:

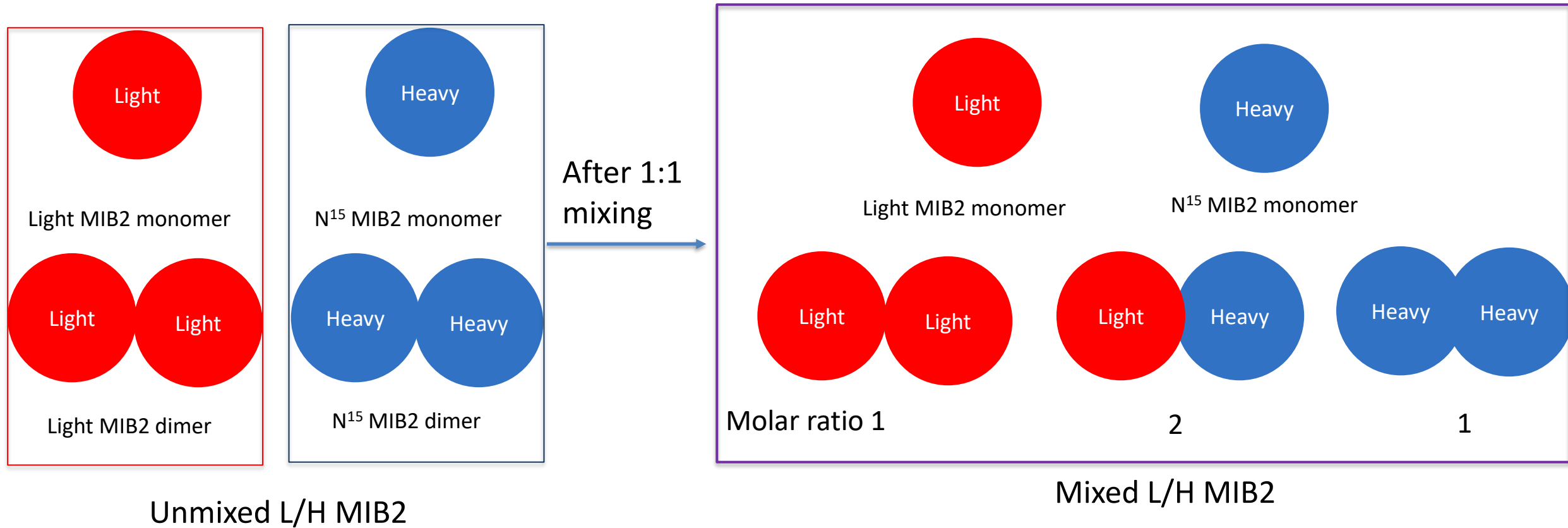
1. intramolecular xlink (within monomer or one subunit of dimer)
2. oligomeric xlink (between two subunits of dimer)
3. mixture of two



How to differentiate them?



Solution: XL-MS of light/heavy mixtures of MIB2 enables differentiation



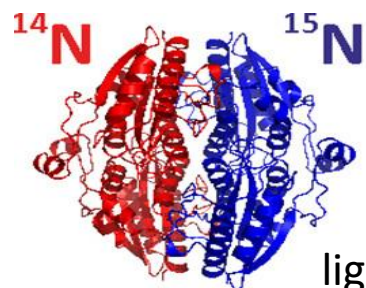
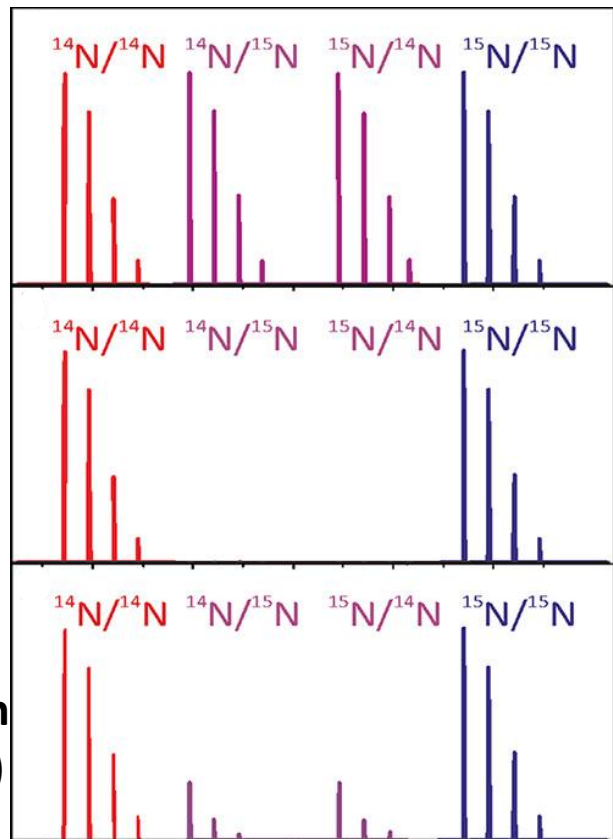


XL-MS of light/heavy mixtures of MIB2 enables differentiation

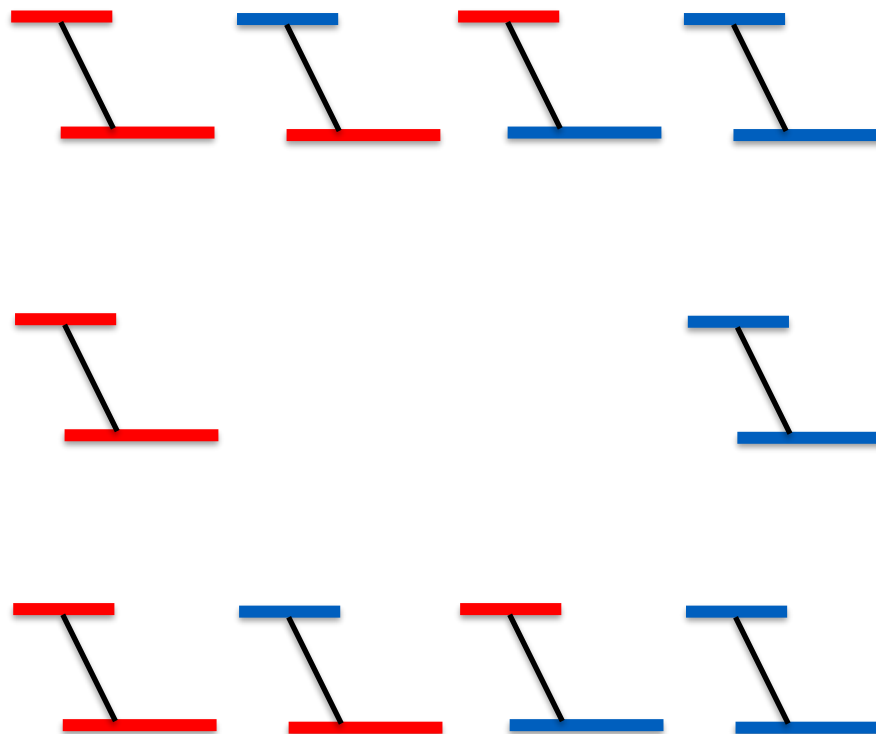
Oligomeric XL

intraXL

Mixture
(if dimer has much
lower abundance)

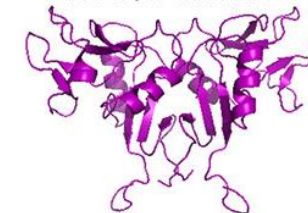


light-heavy MIB2 dimer

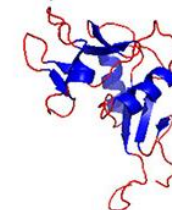


Crosslinked peptides

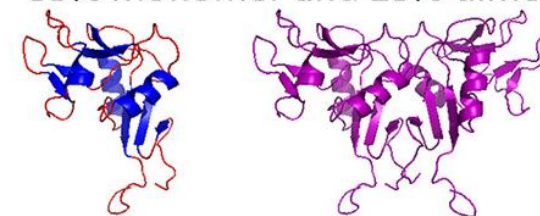
100% dimer



100% monomer



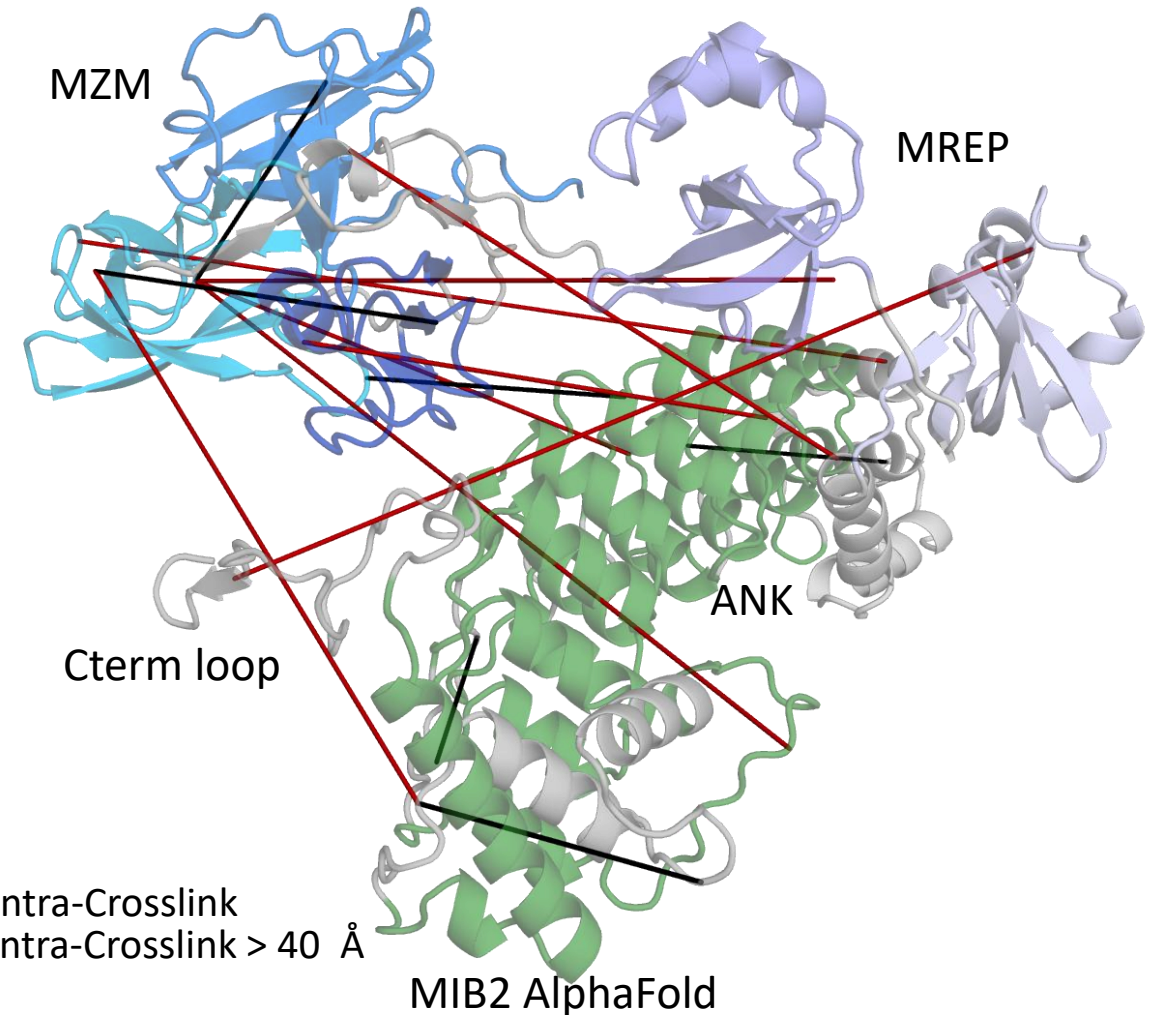
85% monomer and 15% dimer





MIB2 Intra-xlinks contradict AlphaFold predicted structure

Crosslink sites		C α -C α distance on alphafold (Å)	Theoretical range of C α -C α distance (Å)
XL site 1	XL site2		
D131	E228	29.9	9-30
K163	K228	34.5	9-30
E228	K320	69.8	6-16
D293	E457	34.5	9-30
K181	E603	47.5	6-16
K220	D820	61.7	6-16
E228	D583	44.9	9-30
E228	D778	80.4	9-30
E236	E508	82.5	9-30
E251	E645	25.1	9-30
D489	E550	25.8	9-30
K402	E889	76.5	6-16
K689	K725	13.2	9-30
E803	D820	23.8	9-30

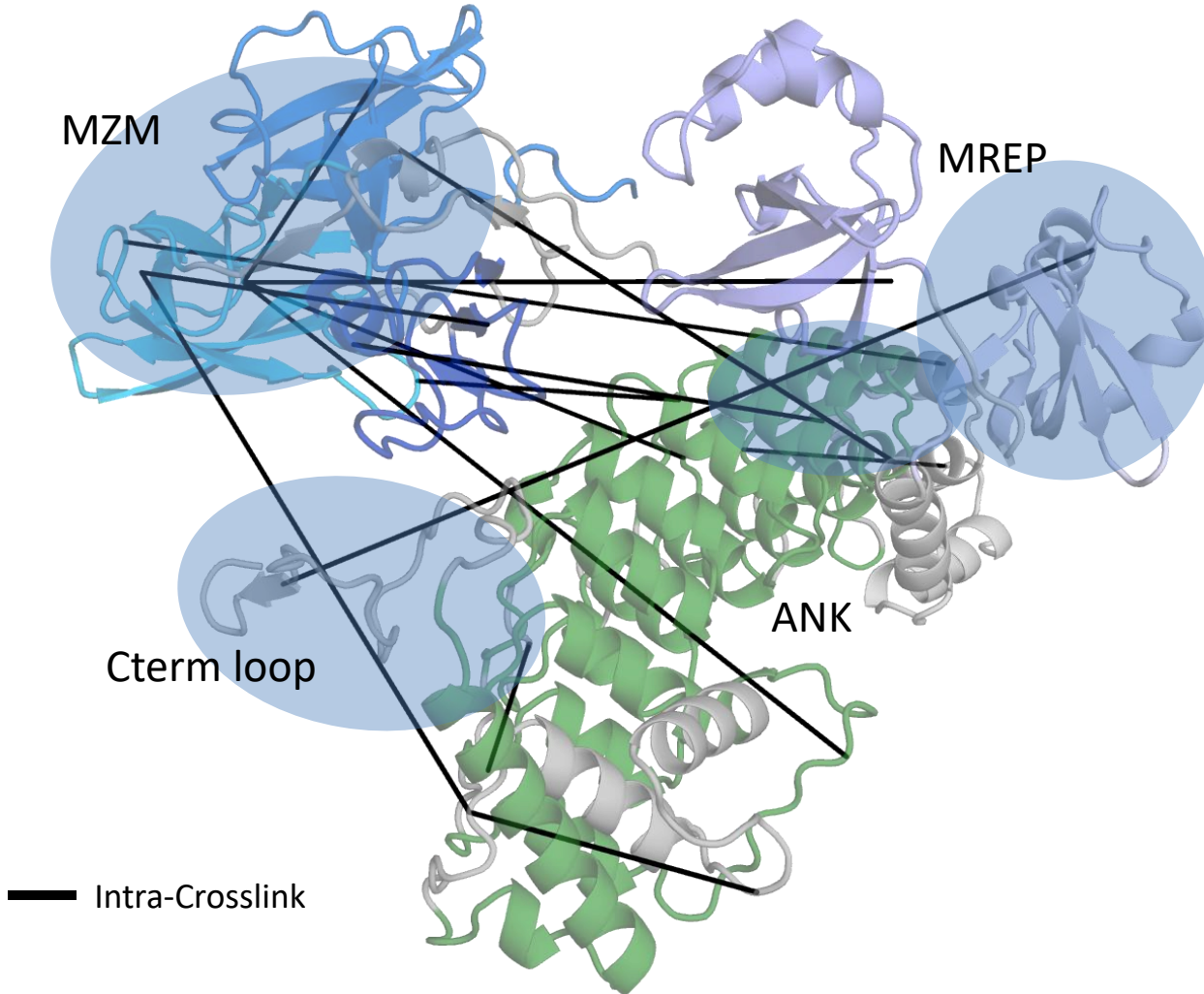


- ❖ Most of the intra-xlinks observed are not reasonable for the alphafold structure
- ❖ Close proximity between MZM-MREP, MZM-ANK, and MREP - Cterm loop

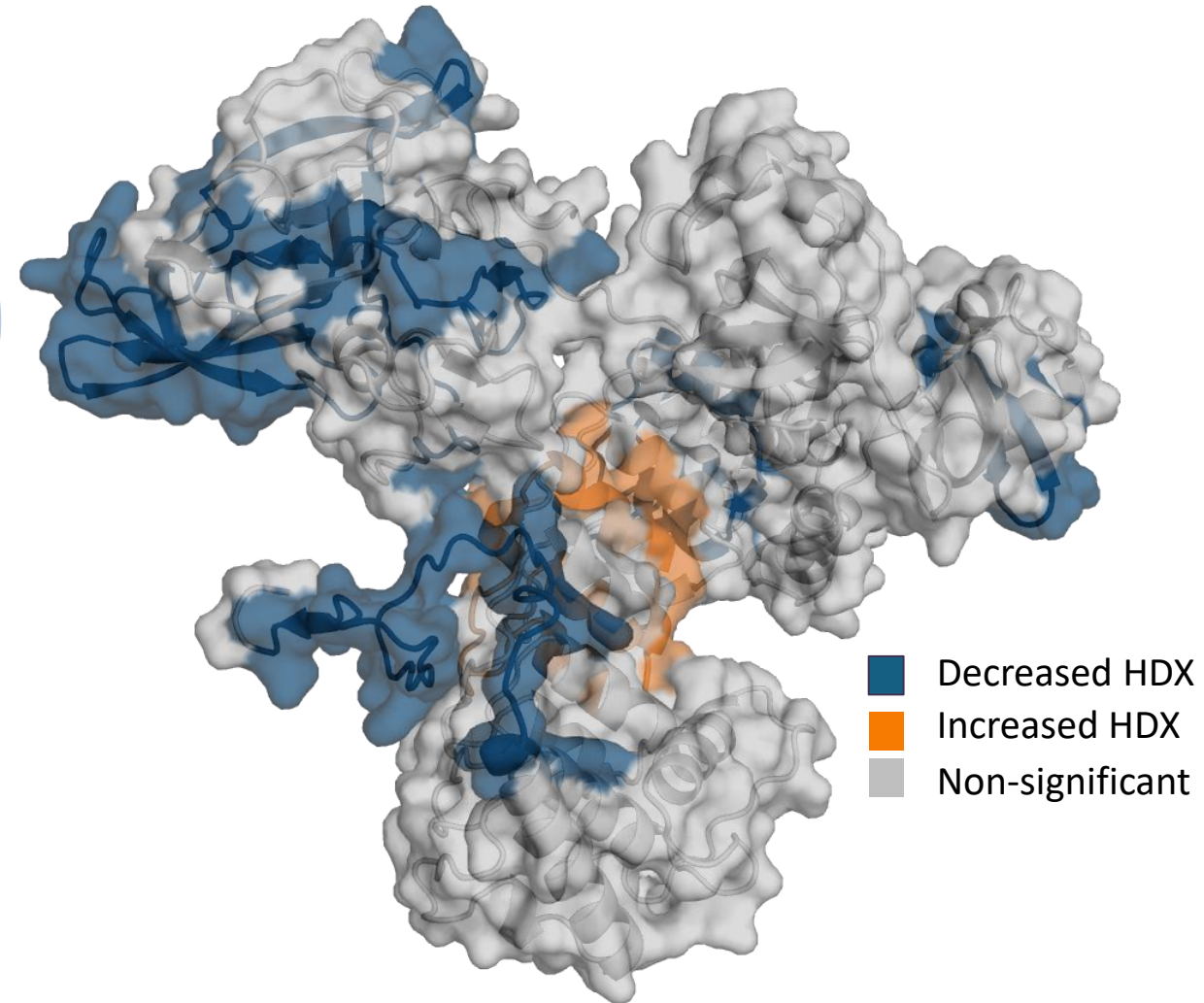


Intra-xlinks indicate the HDX “protected” regions are in close proximity

Summary of intraXL



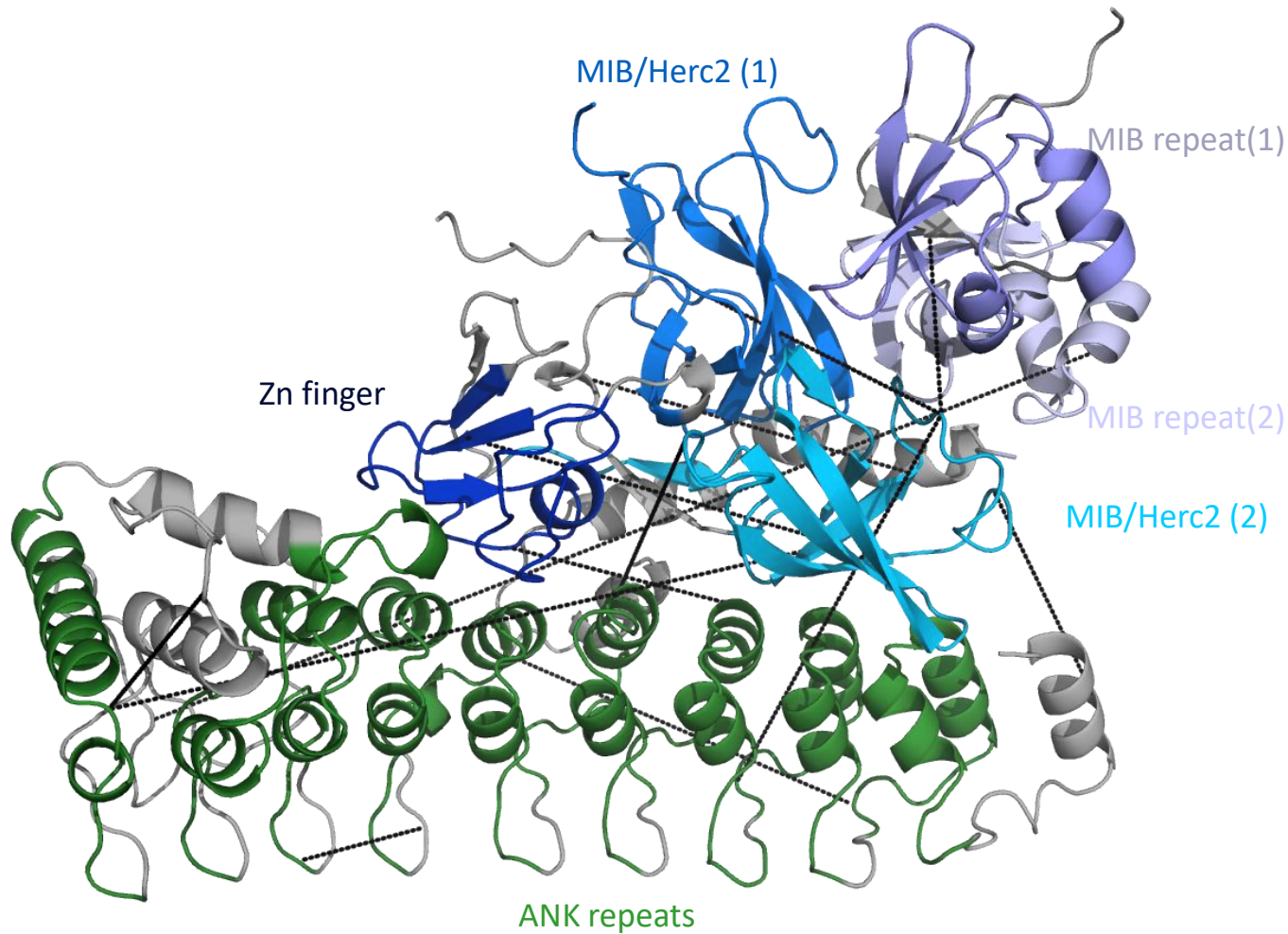
Summary of HDX



❖ Decreased HDX regions seem in close proximity

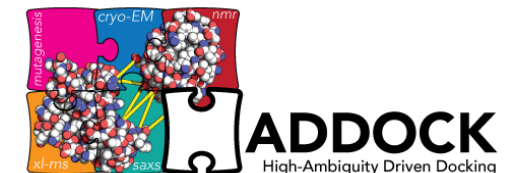


Experimentally derived MIB2 structure



MIB2 docking representative model

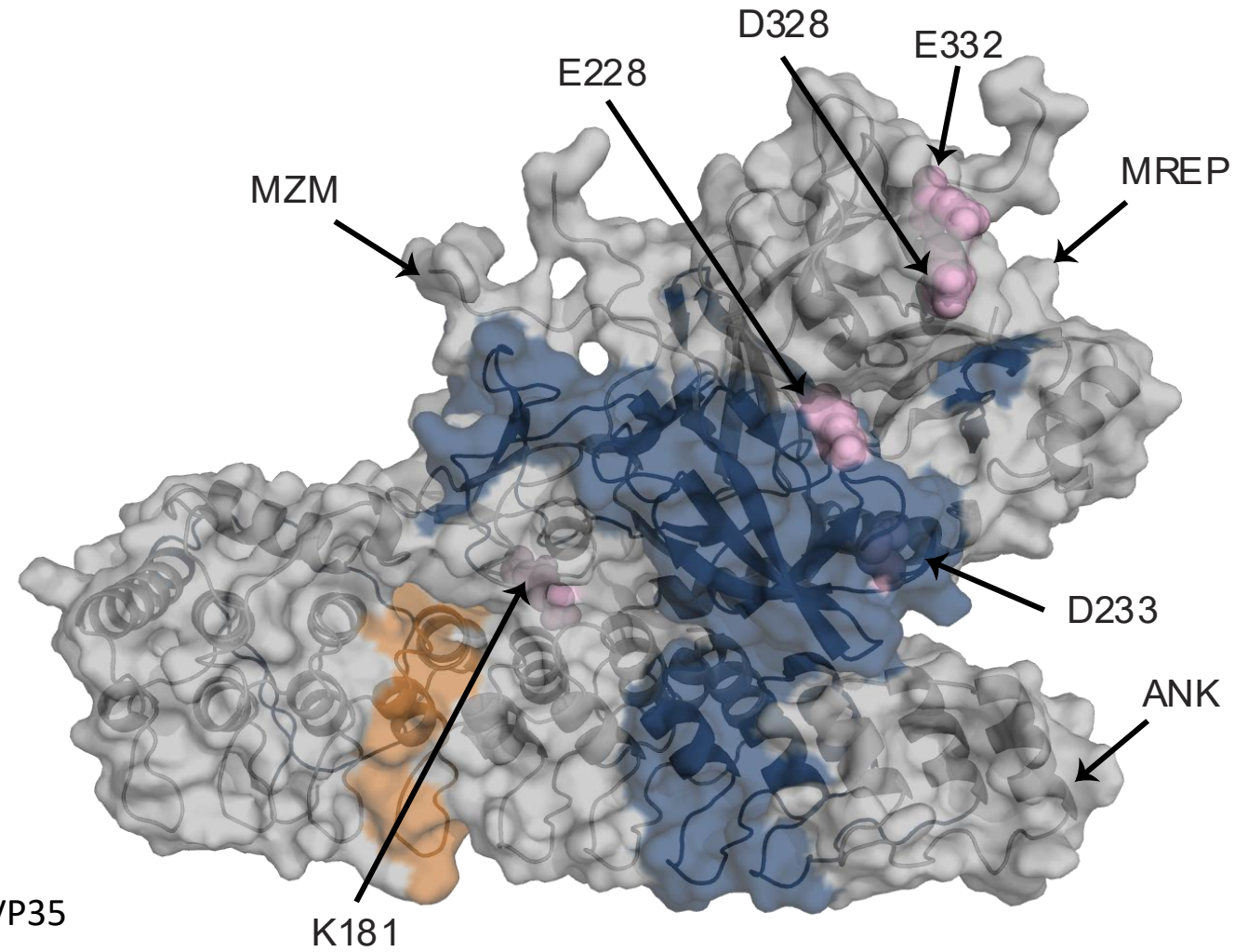
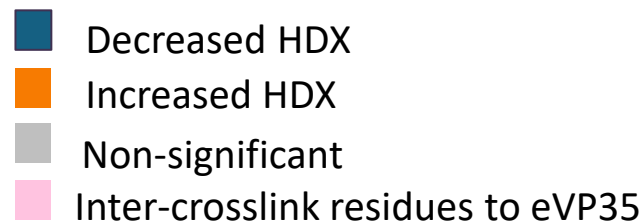
- HADDOCK was used for docking and **intra-crosslink were imported as distance restraints**
- Ambiguous interaction restraints were applied
- Model suggests a more **compact structure** --- contrasting AlphaFold extended structure
- **MZM and MREP domains are positioned closely together and adjacent to the ANK**
- Docking does not account for loop flexibility or consider conformational changes, potentially introducing artifacts due to enforced rigidity.





Experimentally derived MIB2 structure

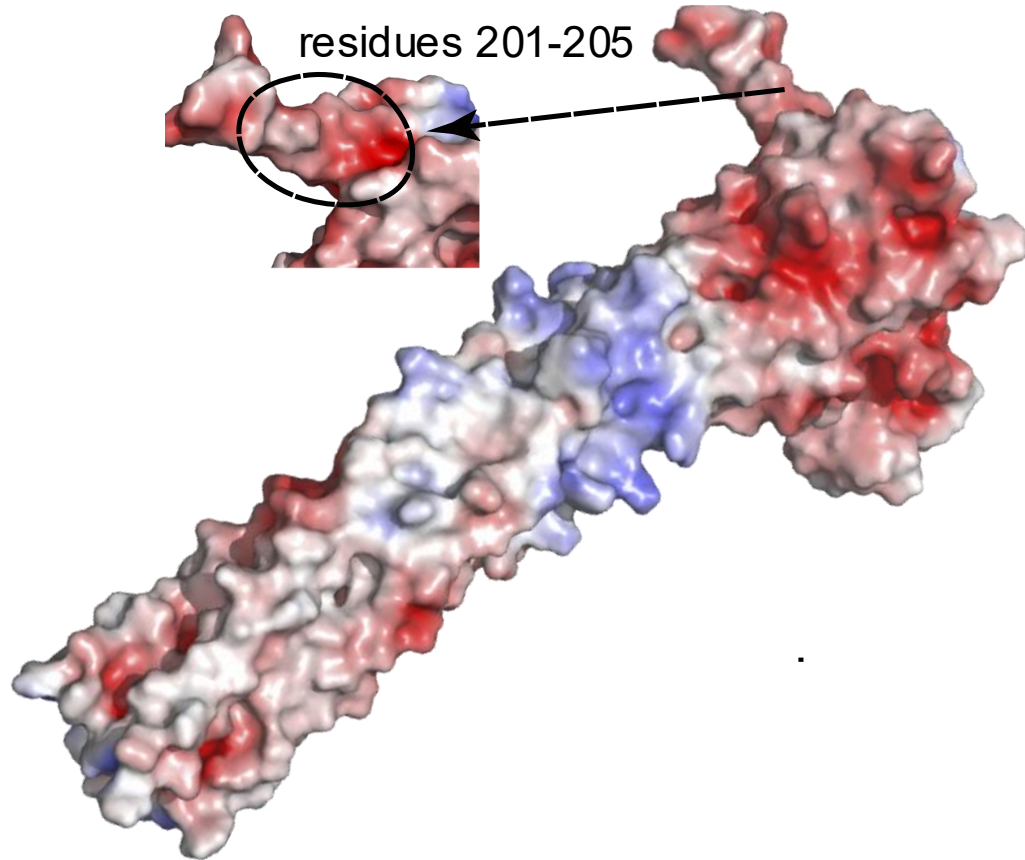
- **The representative model aligns well with HDX and inter-crosslinks results**
- HDX protected regions form a continuous surface (primary protection region)
- MIB2 residues inter-crosslinked to eVP35 are spatially proximal and overlap with the primary protection region



MIB2 docking representative model

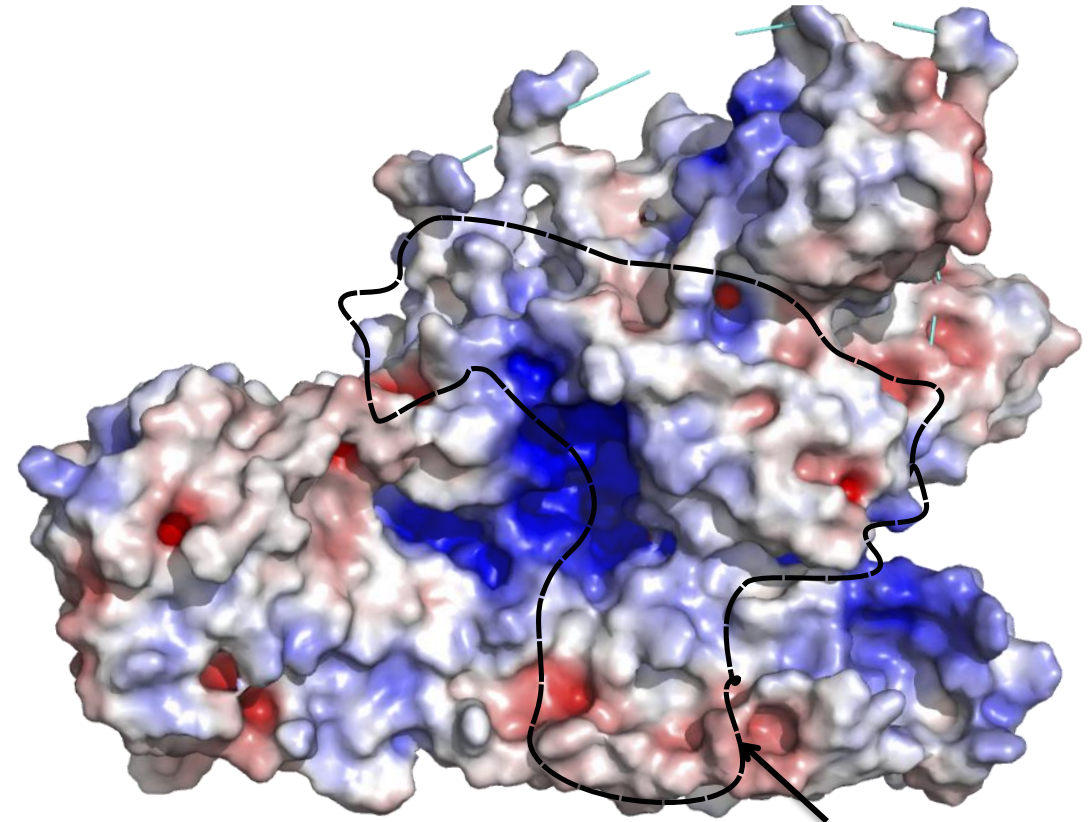


How to validate the docking model?



residues 201-205

eVP35



Primary protected region

MIB2 docking model

This electrostatic complementarity at the interface supports the plausibility of the proposed domains orientation.

The model is not expected to represent MIB2 definitive conformations.

Negatively charged

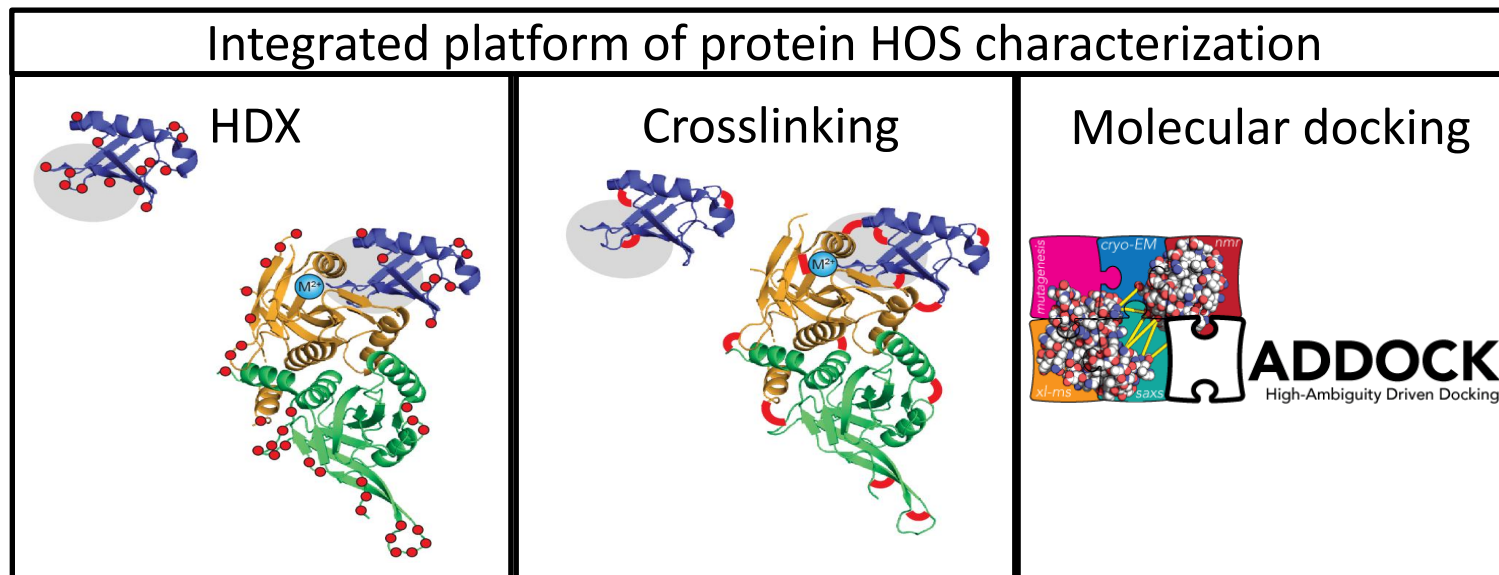


Positively charged



Conclusions and Perspectives

1. Characterized successfully the MIB2-eVP35 binding interface and achieved an experimentally-derived MIB2 structure.
2. Allowed additional structural understanding of Ebola viral pathogenesis.
3. Established an integrated platform for protein HOS and protein-protein interface characterization, particularly for multi-domain, flexible protein like MIB2 that eludes AI-based docking alone.
4. Provided approaches that would aid the design of protein therapeutics





Acknowledgement

Gross Lab

- Michael L. Gross (PI)
- Nicole D. Wagner
- Henry Rohrs
- Don Rempel
- George Mathai
- Tarang Jadav
- Sanjeev Kumar
- Jusung Lee
- Kamesh Mula
- Nuwani Weerasinghe
- Hsin-Chieh Yang
- Nolan McLaughlin
- Cole Hediger
- Xinzhu Li
- Polen Yunus

Amarasinghe & Leung Lab

- Gaya K. Amarasinghe (PI)
- Daisy W. Leung (PI)
- Grace Uwase



National Institutes
of Health

5P01AI120943



Protein Metrics



Thank You for your attention!