

# Rapid Online Buffer Exchange with the **DynaChip X1** Platform for Complex Biological Analysis



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CEO & Co-Founder



**CASSS MS: September 25<sup>th</sup>, 2025**

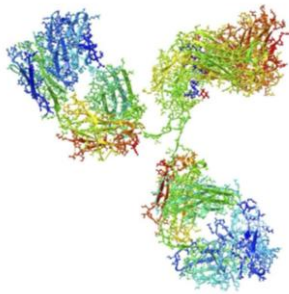
## Overview

- Background
- DynaChip X1 Platform
- Results and Applications

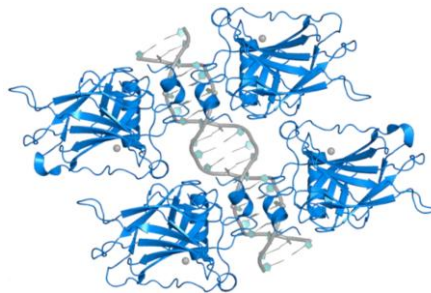
# Structural analysis is the key to biological insight for therapeutics



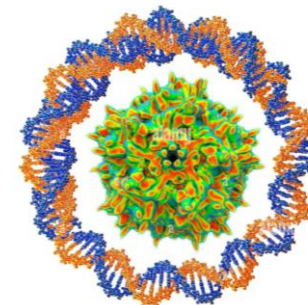
*Native Mass Spec & CDMS are emerging as a powerful tools for structural analysis of complex biomolecules*



**mAbs**



**Disordered Proteins (IDPs)**

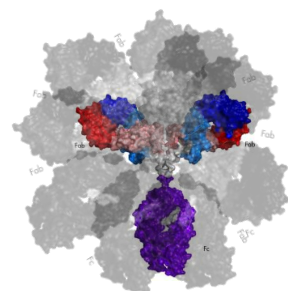


**AAVs**

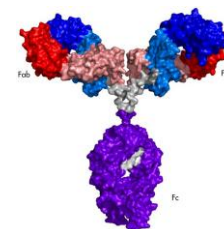
# Mass Spec Challenges

## 1) Sample Prep—Cleaning the sample

Hours per sample to remove buffers, salts, detergents, etc.



Dirty



Clean

## 2) Sample Introduction—Injection into the mass spec

Days to weeks fixing “plumbing issues”



Bubbles



Clogs



Leaks

# DynaChip X1 enables fast and simple mass spec

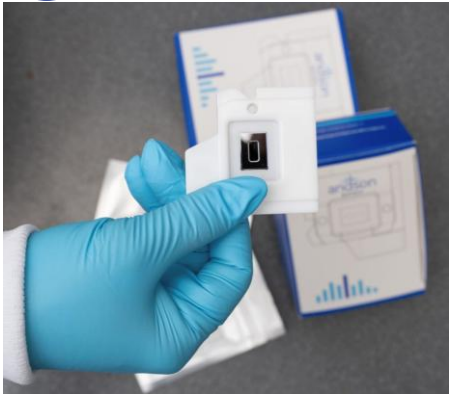


- **Sample cleaning** – High efficiency; Non-destructive
- **Sample injection** – Push-button operation; Results in minutes

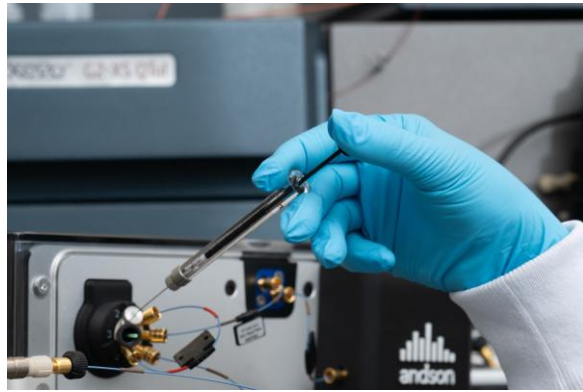
# DynaChip X1 Workflow

*Enables both on-demand and automated analysis*

## 1 Load Chip



## 2 Inject Sample

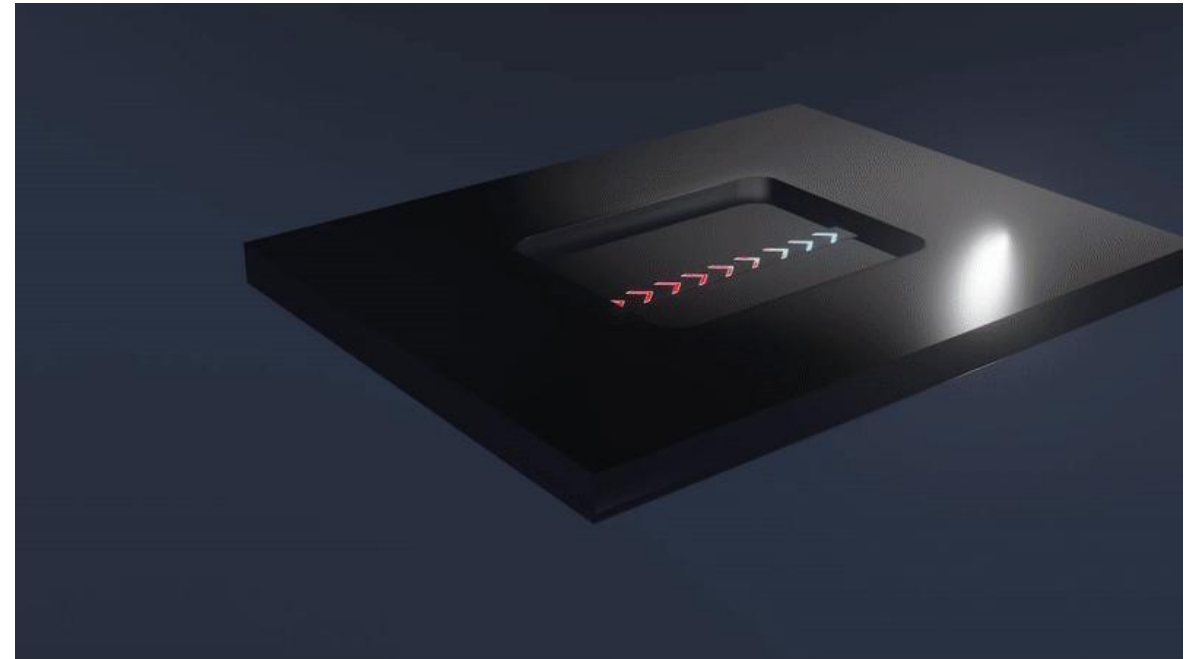


**On-Demand**



**Automated**

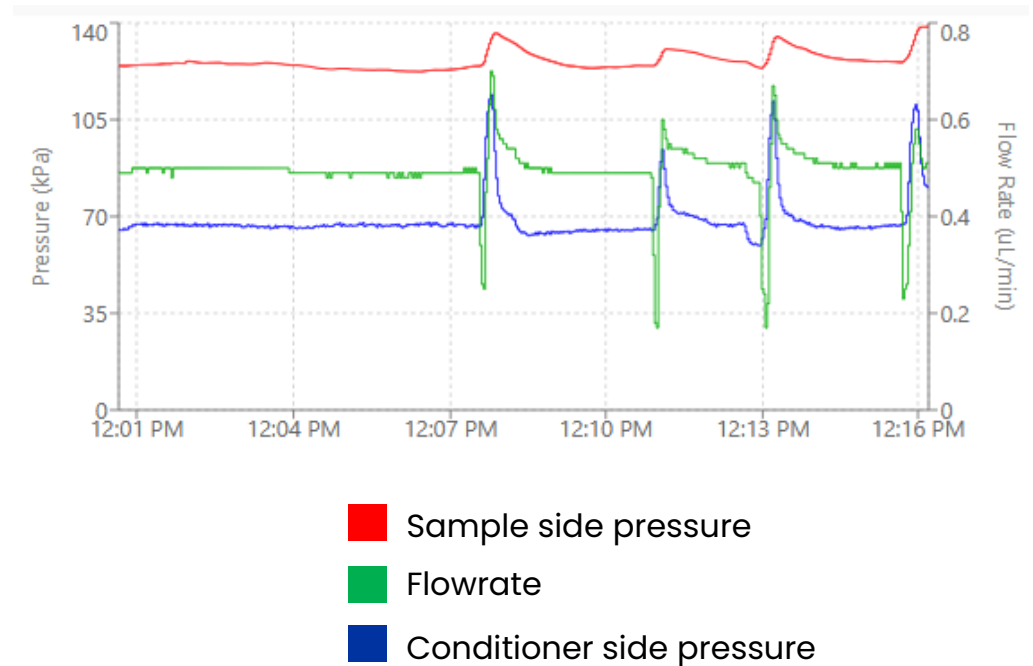
## 3 Results in Minutes



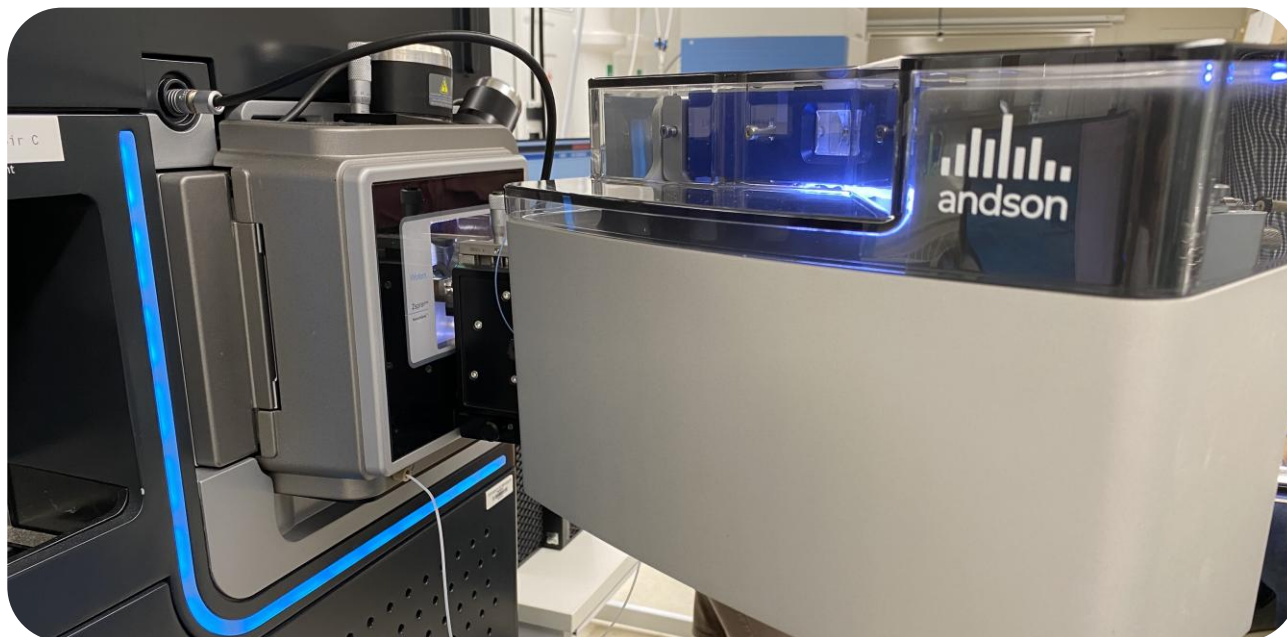
# Fighting Bubbles, Clogs, and Leaks

- Real-time readouts: pressure/flow rate
- Automated alerts of system instability
- Identify “where” the issue is
  - Emitter
  - DynaChip Cartridge
  - Tubing and/or fitting
  - Other

**Real-Time Readout –  
Fused silica clog in union**



# Universal Compatibility



**Roll-up integration with most  
commercial nanoESI interfaces**

**andson**  
BIOTECH

**Thermo** Waters™  
SCIENTIFIC

 **Agilent Technologies**

**M**egadalton  
**S**olutions

**BRUKER**

**SCIEX**  
Coming soon

**TrueMass**  
Spectrometry

# DynaChip X1 Platform

Fundamentals & Capabilities

# Selected List of Reagents Removed

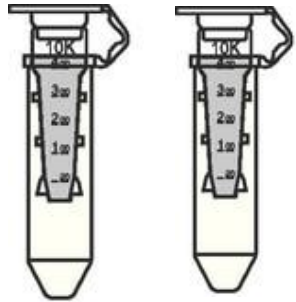
| Reagent            | Concentration Tested |
|--------------------|----------------------|
| ★ Ammonium sulfate | 500mM                |
| ★ NaCl             | 3M                   |
| KCl                | 2.7mM                |
| Sodium Phosphate   | 20mM                 |
| ★ Urea             | 8M                   |
| Glycine            | 250mM                |
| Histidine-HCl      | 20mM                 |
| Imidazole          | 17.5mM               |
| ★ PBS              | 1X                   |
| Tris               | 80mM                 |
| HEPES              | 10mM                 |
| Sucrose            | 25mM                 |

| Reagent          | Concentration Tested |
|------------------|----------------------|
| Hexylene glycol  | 1%                   |
| Formamide        | 70%                  |
| Pluronic F-68    | 0.1%                 |
| ASB-14           | 300uM                |
| C8E4 Detergent   | 2X CMC               |
| MEGA-8           | 1mM                  |
| LADO             | 0.05%                |
| Lauryl Maltose   | 0.01%                |
| Neopentyl glycol |                      |
| Cholesteryl      | 0.001%               |
| Hemisuccinate    |                      |
| <b>And More</b>  |                      |

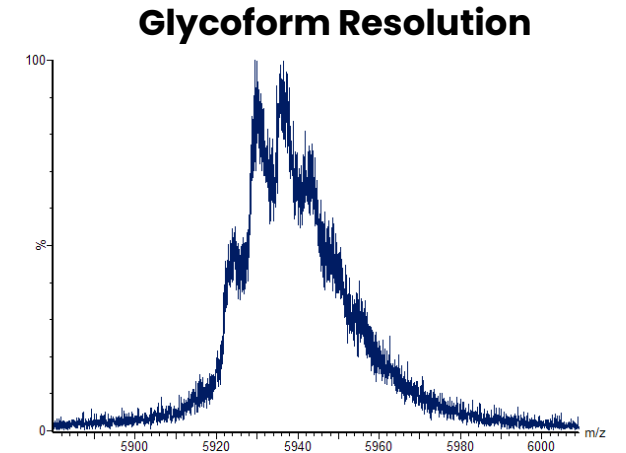
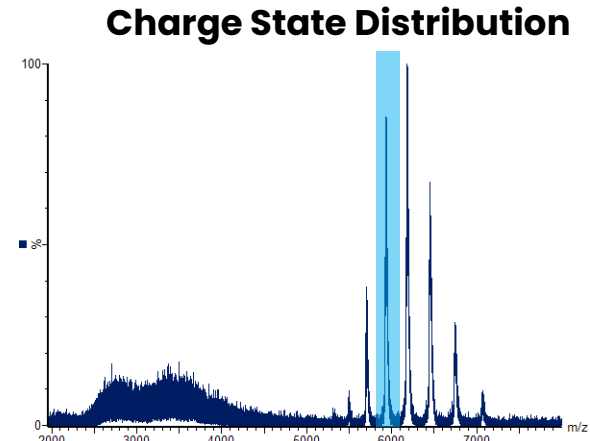
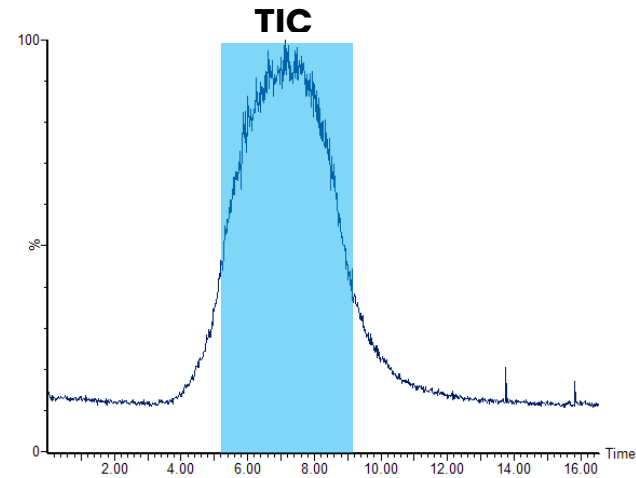
★ = Data shown in this presentation

# DynaChip X1 vs. Spin Filter

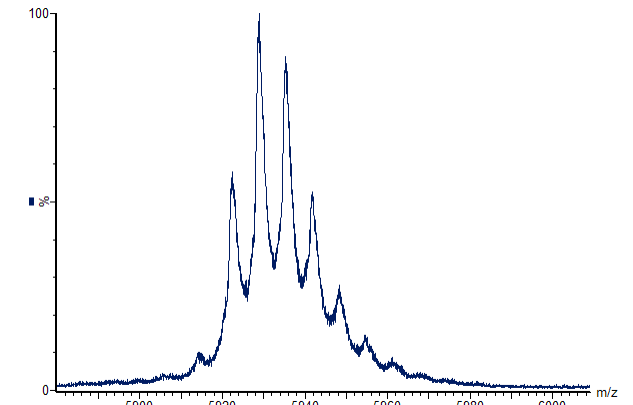
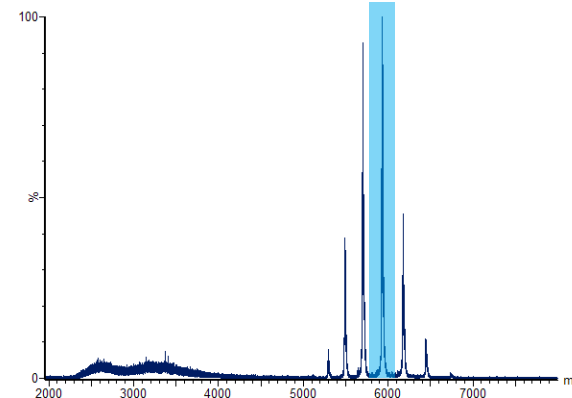
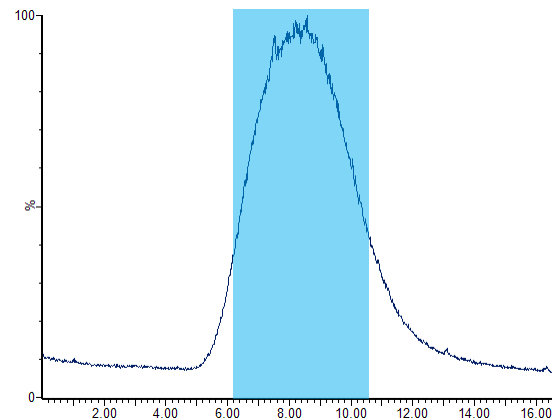
*Online clean up of 1x PBS — NIST mAb*



**2X Spin Filter (30kDa):  
60 min total**



**DynaChip X1:  
15 min total run time**



Data Acquired on: Waters Xevo G2-S Q-TOF

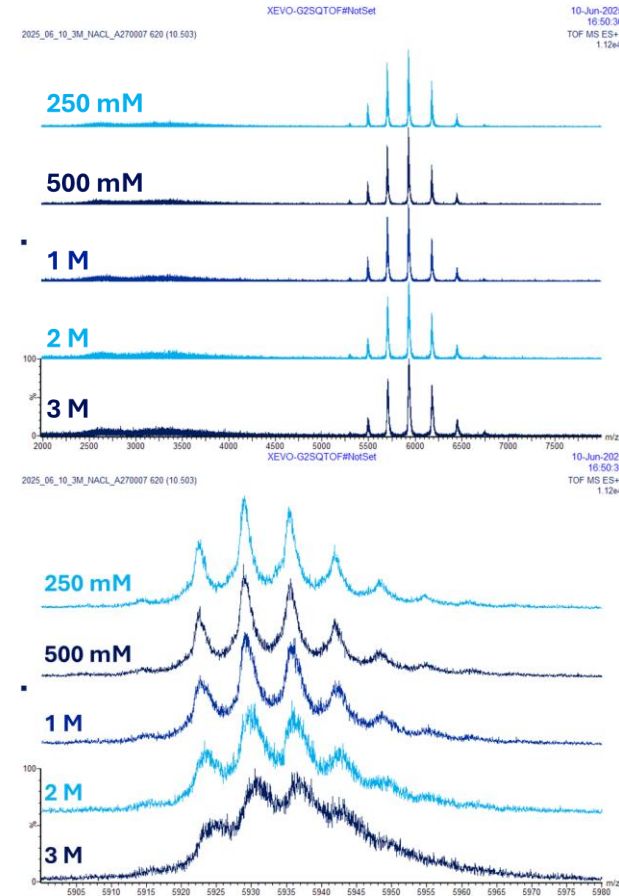
# Exploring DynaChip Buffer Exchange Limits

*Removal of 250 mM – 3 M NaCl for Glycoform Resolved NIST mAb*

**1.286g NaCl in 7.33 mL  
water**



**49% saturated  
solution**



**Full Spectrum**

**Single charge  
state zoom**

- 0.1 mg/ml NIST mAb exchanged out of NaCl solutions

Data Acquired on: Waters Xevo G2-S Q-TOF

# NIST mAb "Refolds" after Removal of 8M Urea

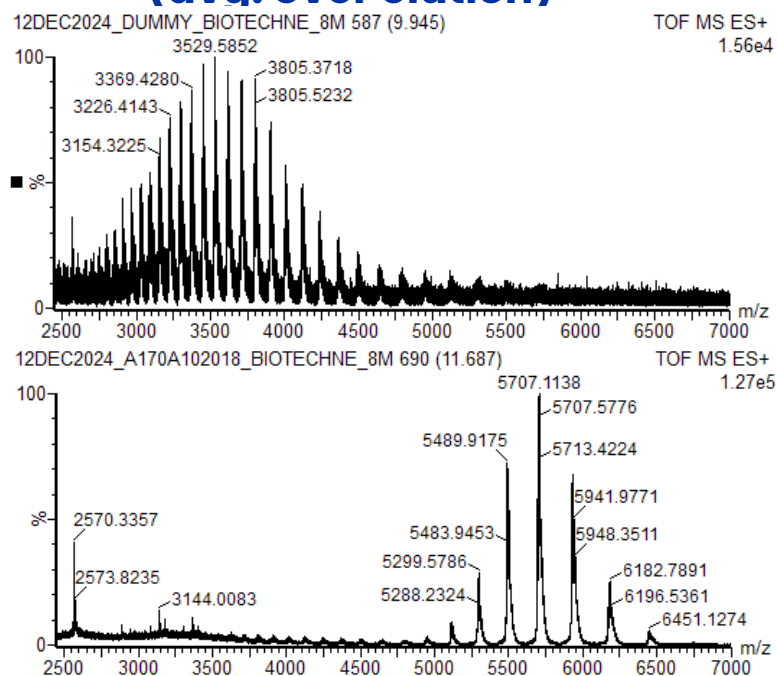


Direct  
Infusion

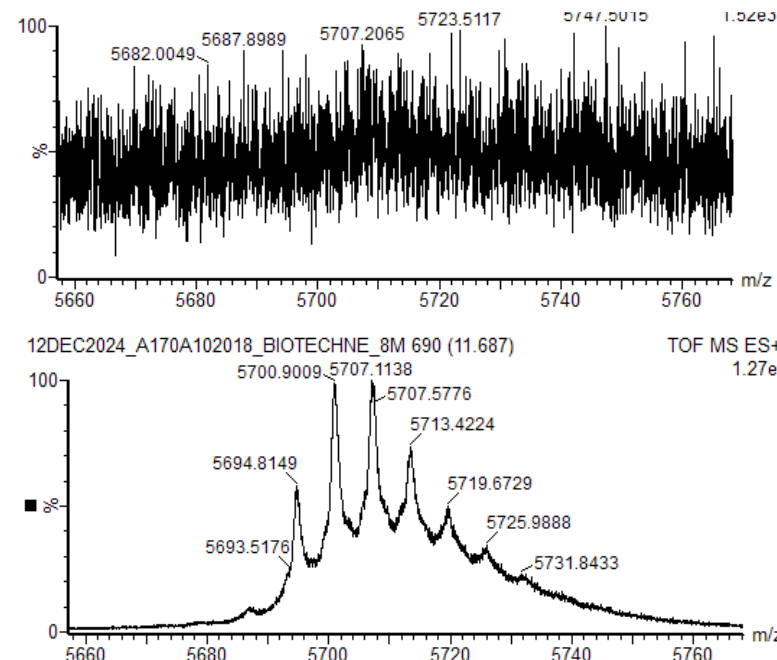


DynaChip XI

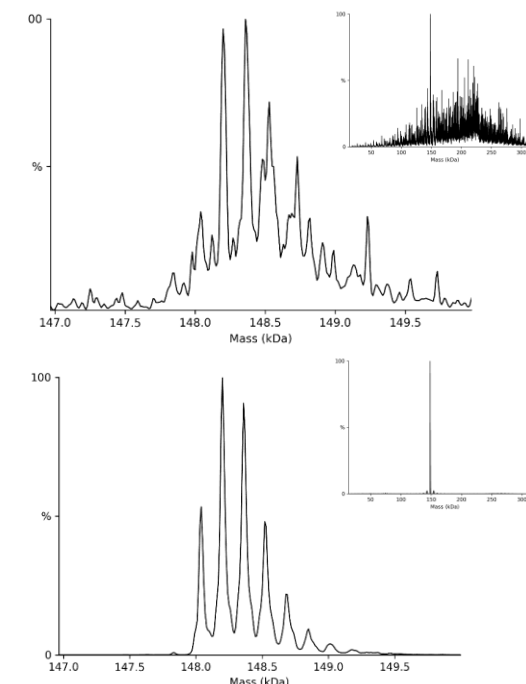
## Spectra (avg. over elution)



## +27 Charge State



## Deconvolution



- Glycoform resolution achieved following **DynaChip XI** buffer exchange
- Maintenance of **native confirmation** after sample cleanup
- Clean deconvolution following **DynaChip XI** sample treatment

biotechne®

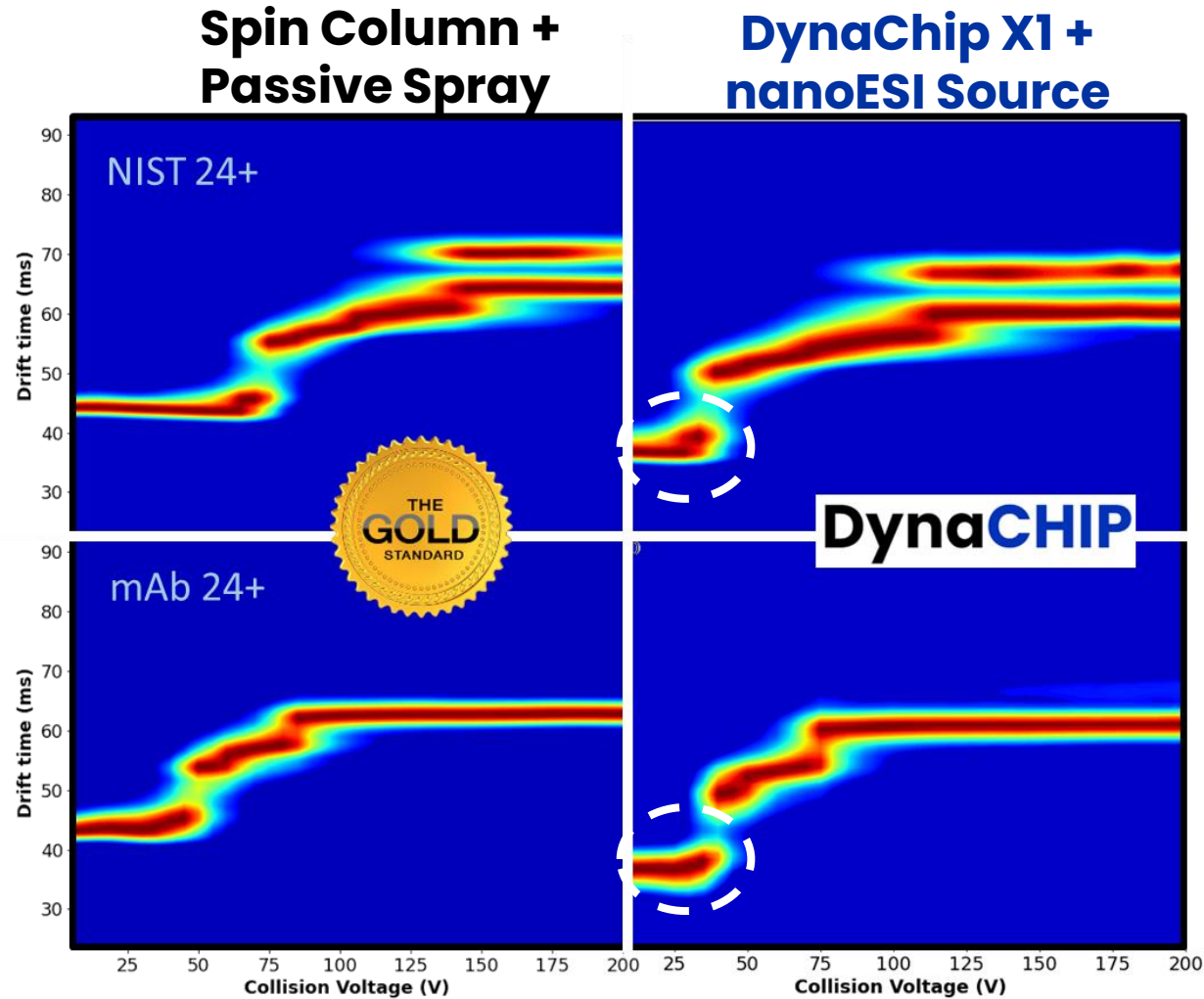
proteinsimple

Data Acquired on: Waters Xevo G2-S Q-TOF

# Results and Applications

mAbs & ADCs

# Native MS – Collision Induced Unfolding

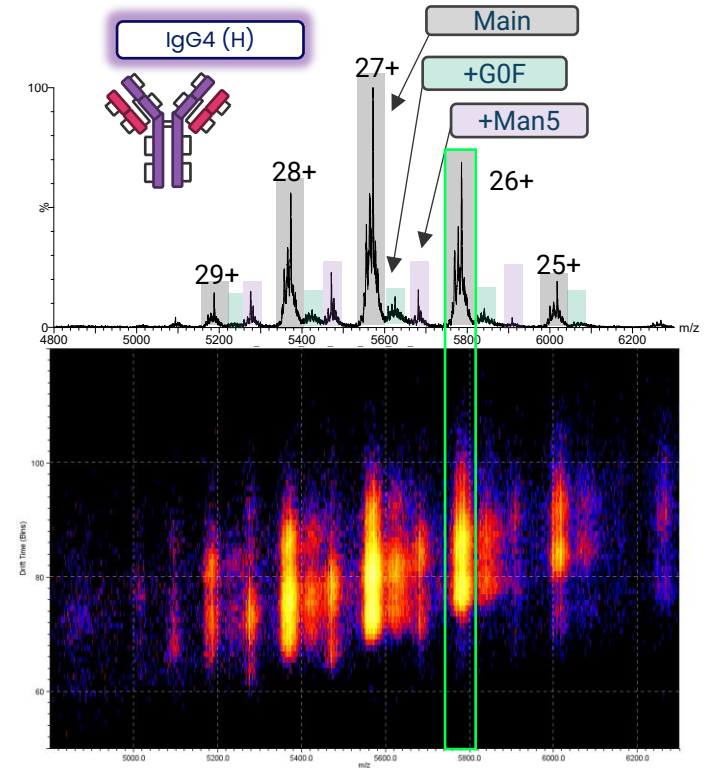
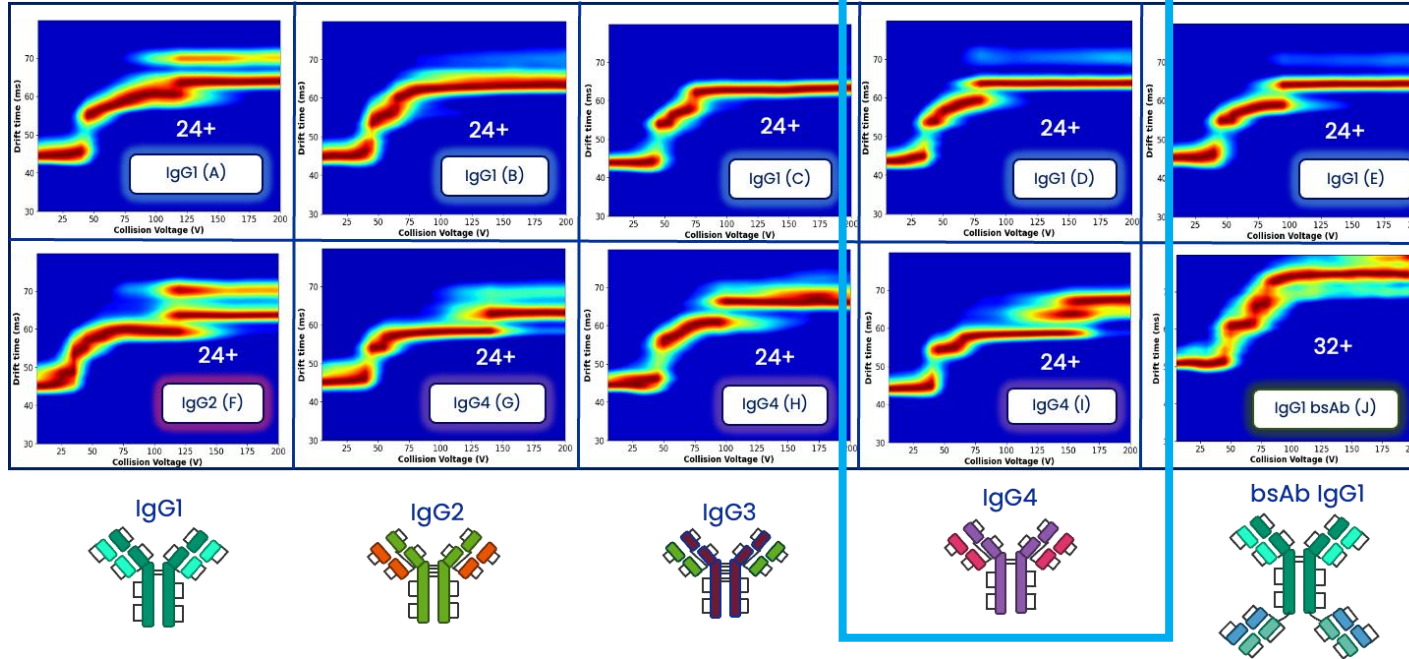


- Maintained **ground state transition** with DynaChip flow-through buffer exchange
- **Simplified native workflow:** commercial ion sources, no prior buffer exchange

CIU Voltage Ramp: 5V steps @ 18 seconds (40 steps)  
= 12 min CIU collections

# Native MS – Collision Induced Unfolding & Ion Mobility

## Ion Mobility

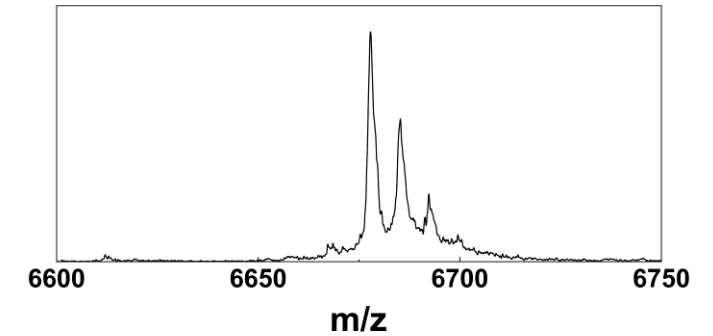
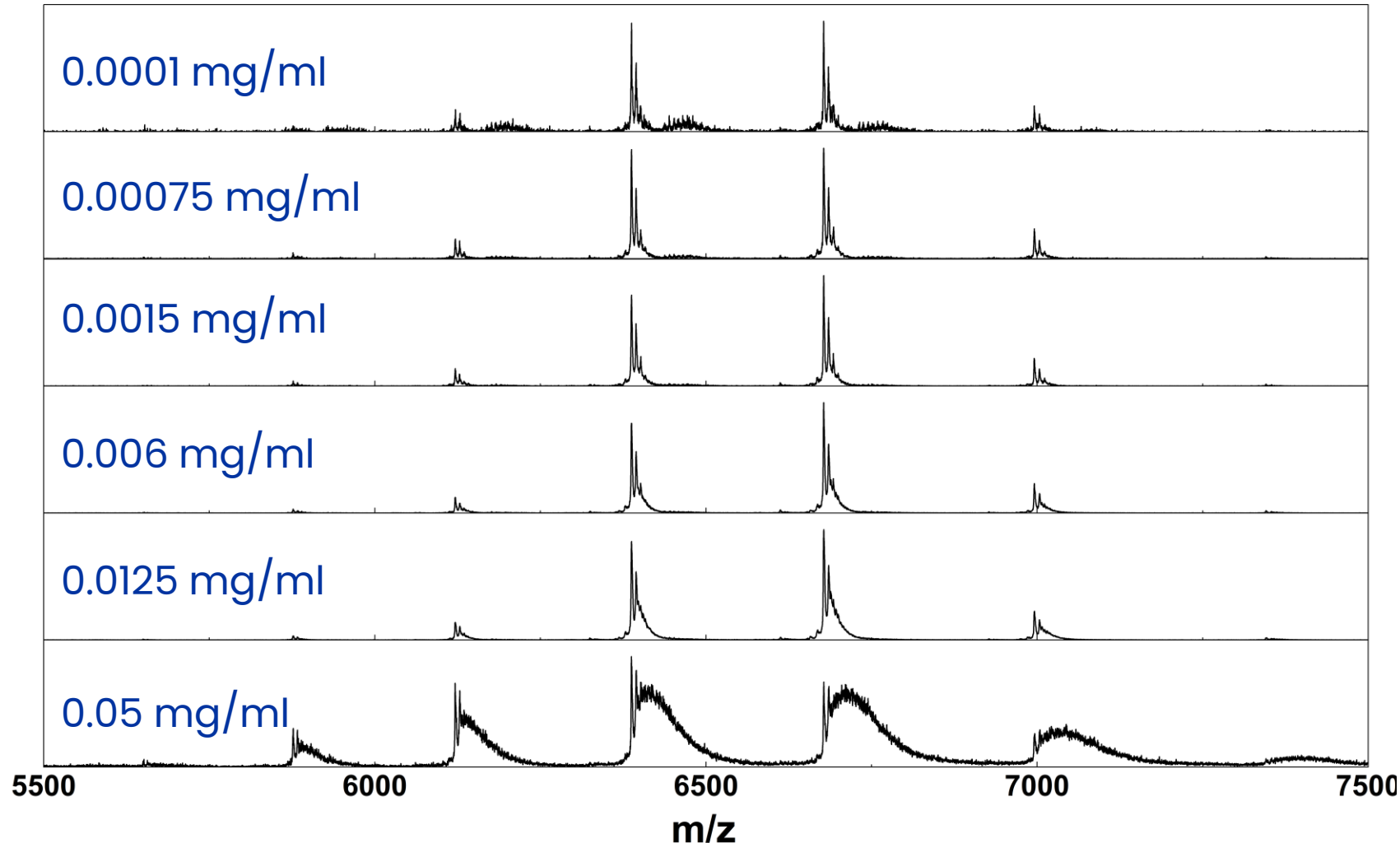


- CIU analysis of multiple mAb classes (above)
- CIU of forced degradation on Ig2 (*not shown*)

- CIU, IM-MS captured significant impact of Mannose-5 on stability
- Not captured as clearly in PTM analysis via LC-MS

# Direct from Crude Cell-Culture Media mAb

Highly sensitive analysis from day 5 culture media – **2.3 mg/mL**



- ✓ 3 bioreactor conditions
- ✓ 0-14 or 0-21 days
- ✓ Biological replicates
- ✓ **Over 100 samples run in ~ 2 days**

In collaboration with:



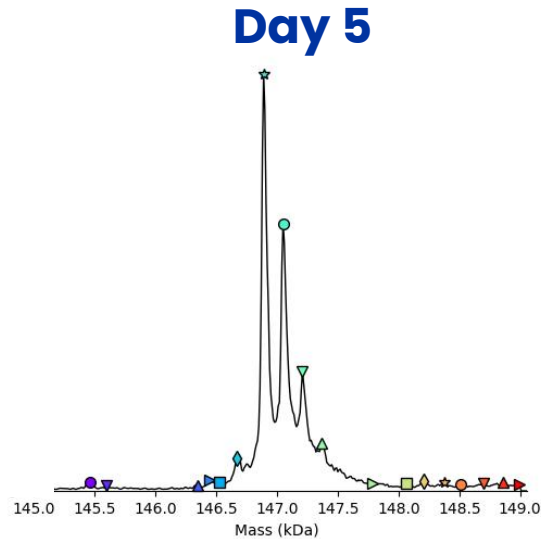
The University of Manchester

Data Acquired on: Waters Cyclic IMS

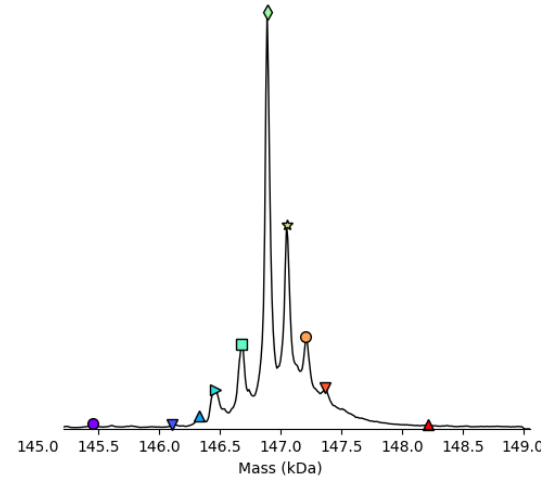
# Direct from Crude Cell-Culture Media mAb

*High-throughput mAb product monitoring – Ambr250 bioreactors*

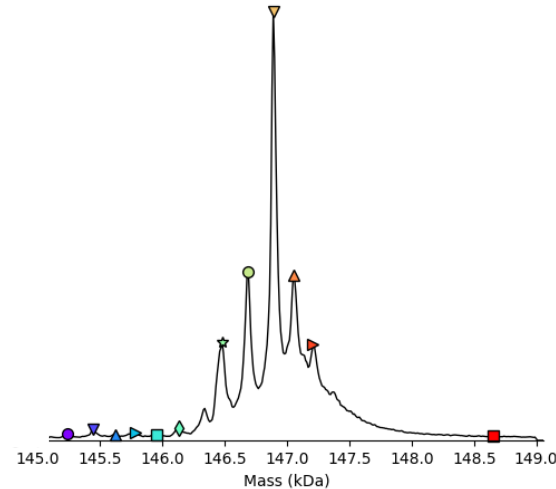
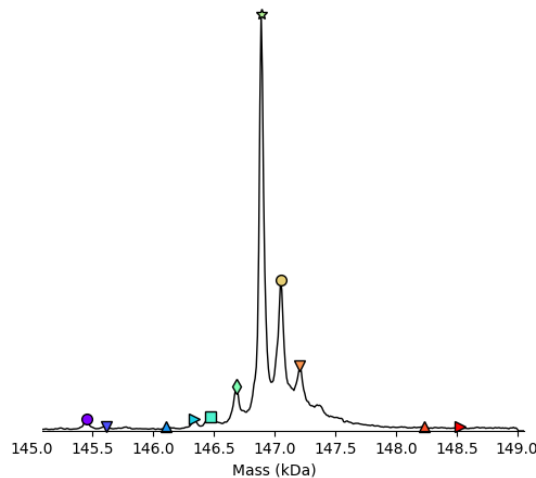
Control



Day 14



Intensified  
Fed-Batch



- ✓ Glycan Profile
- ✓ Condition comparison
- ✓ 10 minute per sample

In collaboration with:

MANCHESTER  
1824

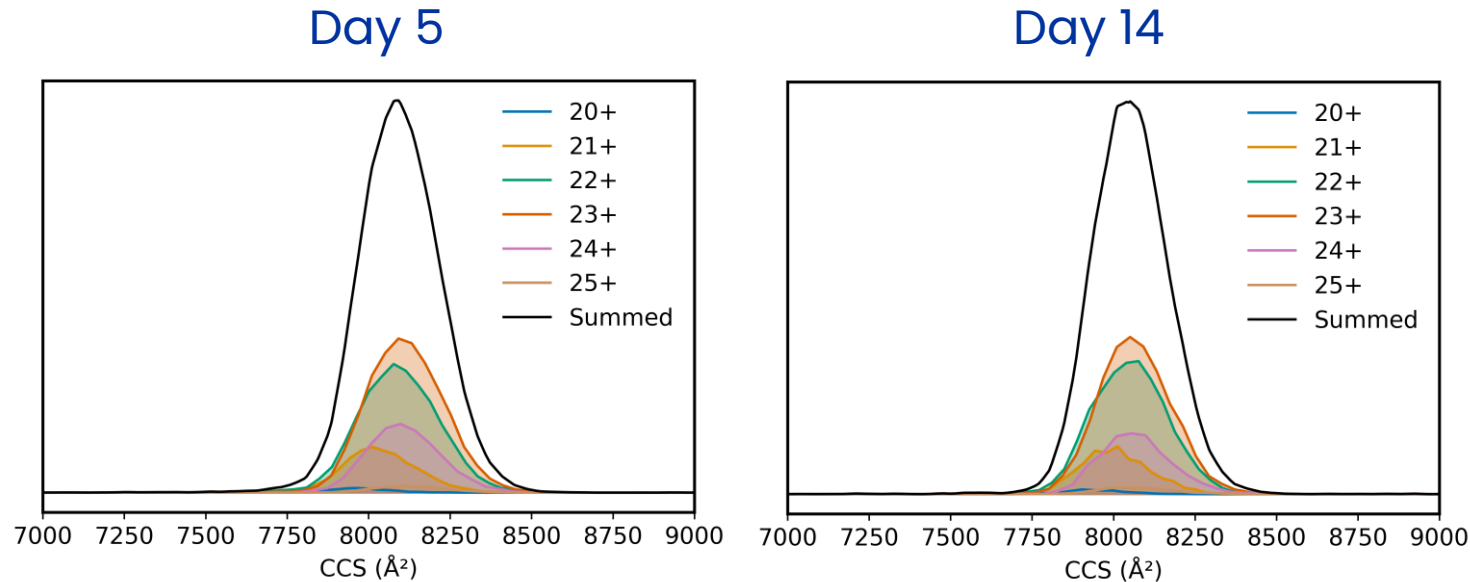
The University of Manchester

Data Acquired on: Waters Cyclic IMS

# Direct from Crude Cell-Culture Media mAb

High-throughput mAb product monitoring – Ambr250 bioreactors

Collision  
Cross  
Section  
Distribution



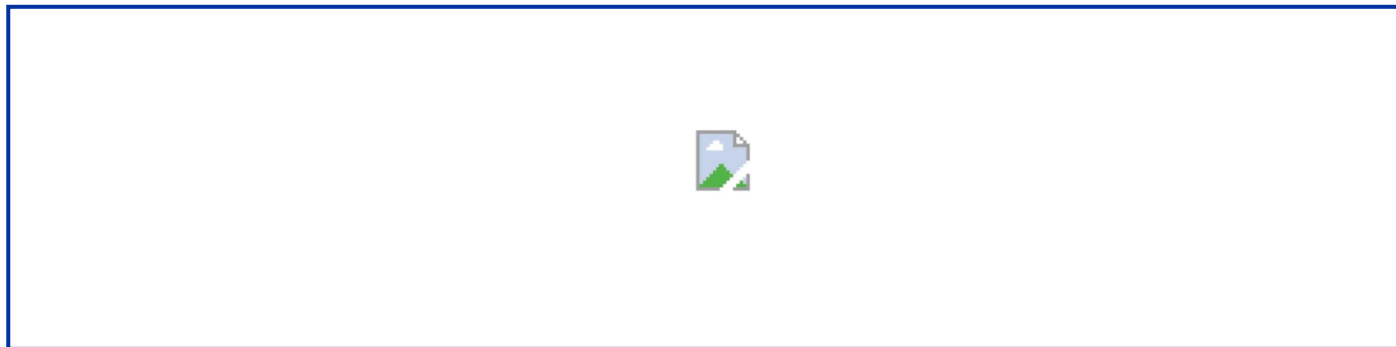
✓ **Collision Cross Section Distribution**

✓ **CIU**

✓ High level structural insights

✓ 10 minute per sample

CIU



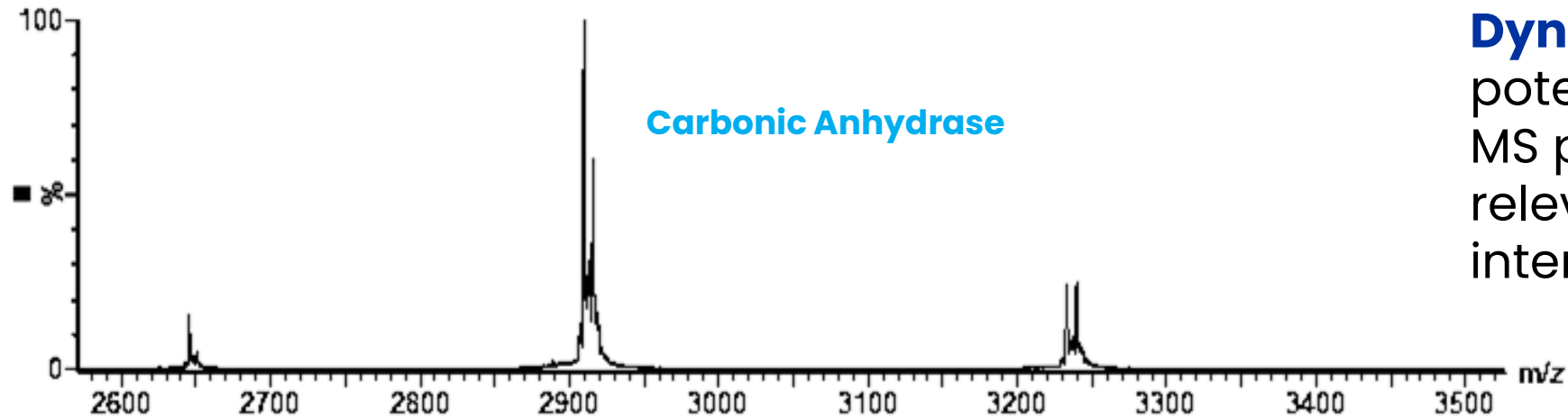
In collaboration with:

MANCHESTER  
1824

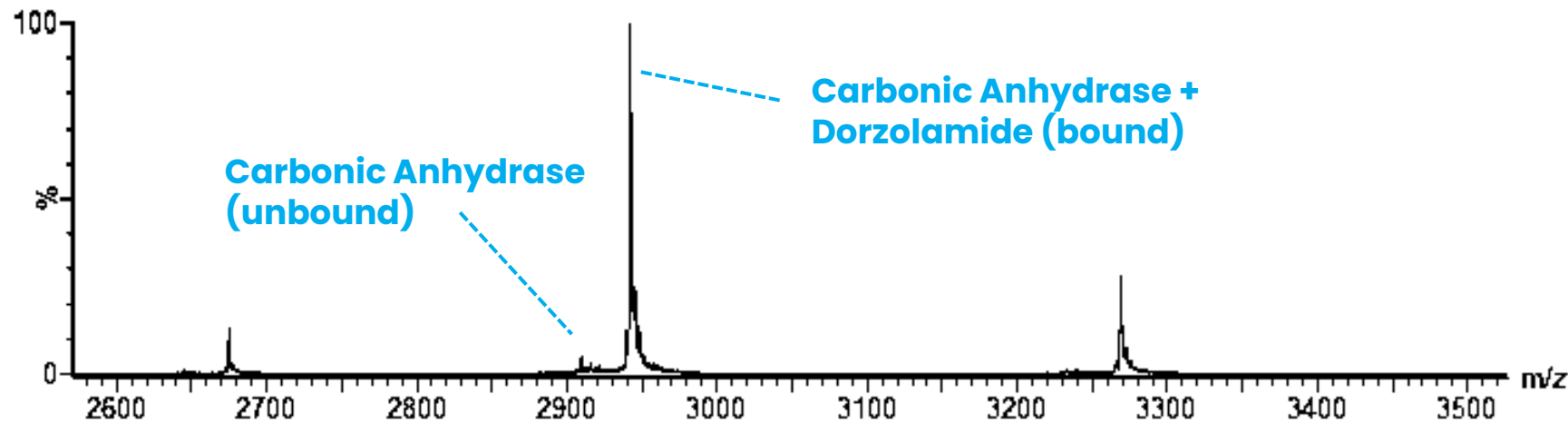
The University of Manchester

Data Acquired on: Waters Cyclic IMS

# Protein-Ligand Interactions



**DynaChip X1** has the potential to provide native MS platform for biologically relevant protein-ligand interactions.

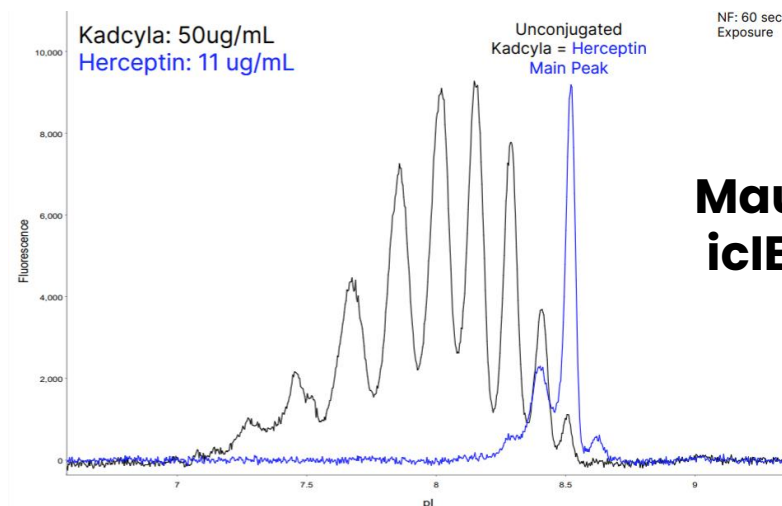


Data Acquired on: Waters Xevo G2-S Q-TOF

# ADCs—Drug Antibody Ratio (DAR) Analysis

*Compatibility with 3rd Party Systems like MauriceFlex*

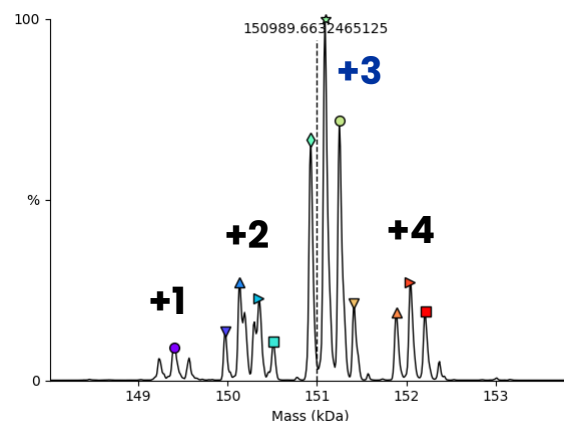
Acidic peaks have increasing lysine conjugation or increasing drug to antibody ratio (DAR)



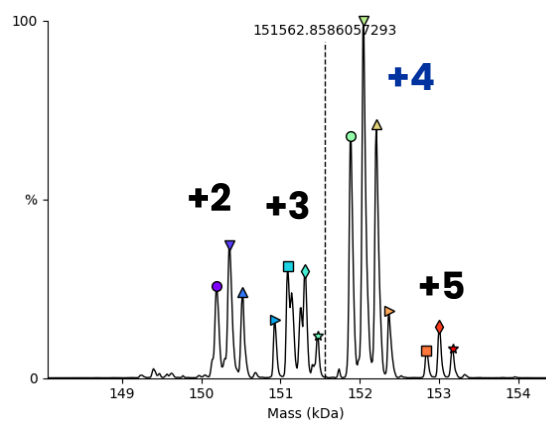
**MauriceFlex  
icIEF Profile**

## Representative Deconvolutions

★ **Fraction C12**



★ **Fraction C11**



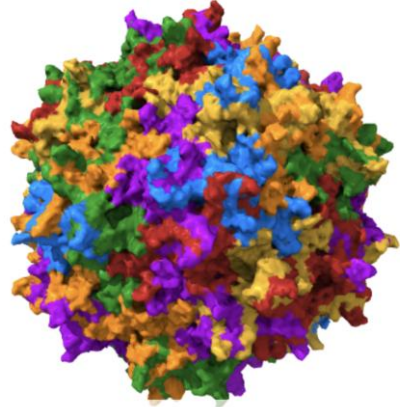
pI ↑

| Example Fraction     | Average DAR |
|----------------------|-------------|
| Herceptin (Unloaded) | 0.00        |
| B12                  | 2.52        |
| ★ C12                | 2.88        |
| ★ C11                | 3.48        |
| C9                   | 4.35        |
| C7                   | 4.70        |
| C6                   | 5.07        |

# Results and Applications

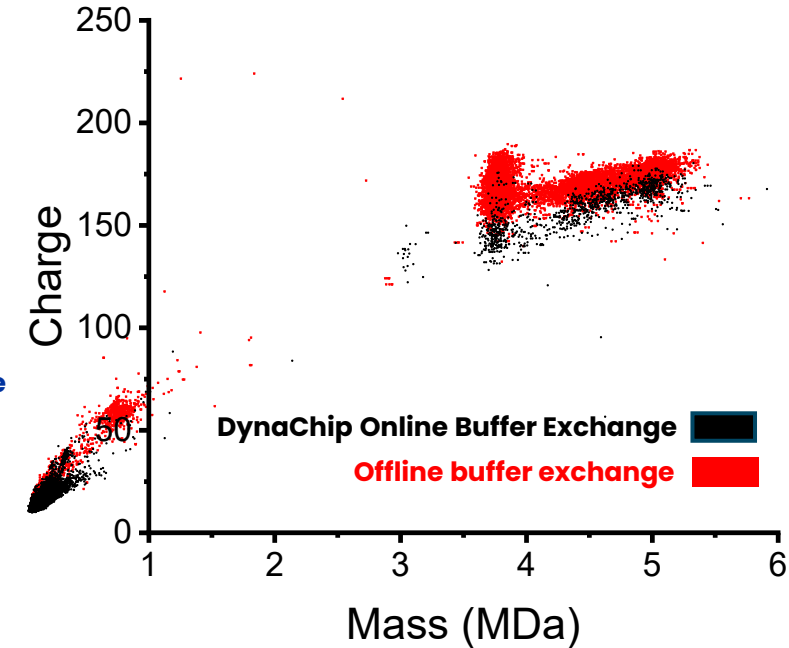
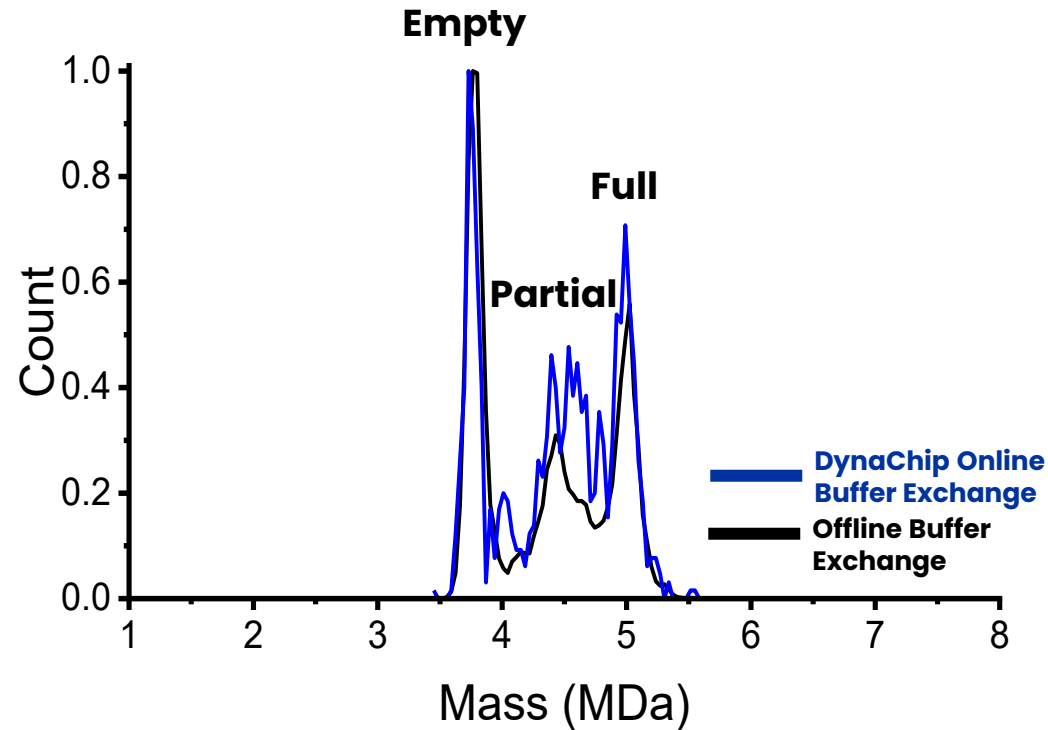
Large Molecules & CDMS

# Full/Empty AAV Capsid Ratio with Online DynaChip X1



**AAV8**

scDNA HEK production

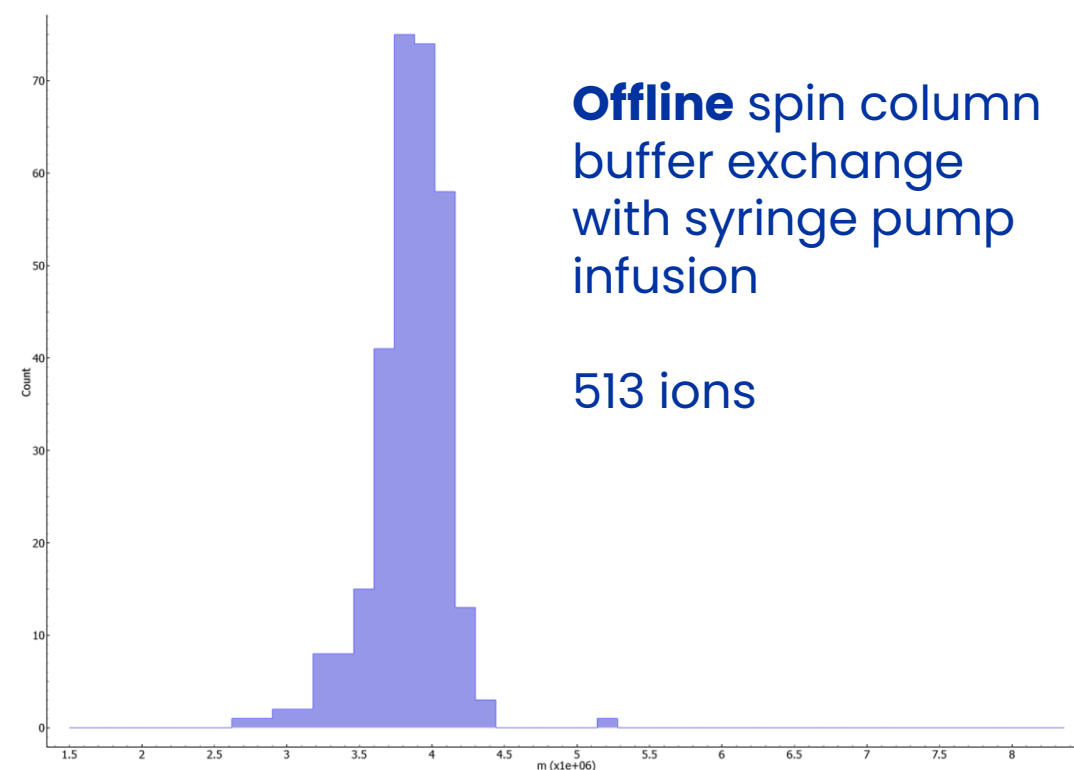
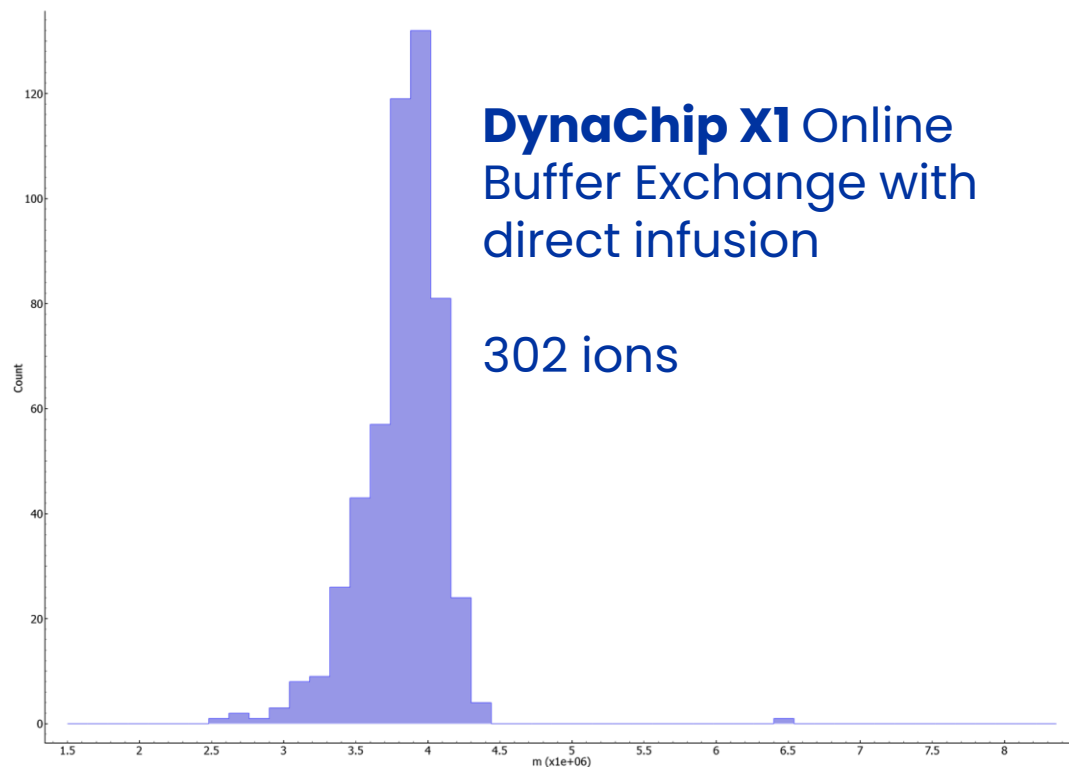


| Prep                        | Structure      | Mass (MDa)   | FWHM (MDa)   |
|-----------------------------|----------------|--------------|--------------|
| Offline<br>(2x spin column) | Empty          | 3.777        | 0.163        |
|                             | Partial        | 4.481        | 0.135        |
|                             | Full           | 5.008        | 0.101        |
| <b>DynaChip</b>             | <b>Empty</b>   | <b>3.754</b> | <b>0.122</b> |
|                             | <b>Partial</b> | <b>4.537</b> | <b>0.220</b> |
|                             | <b>Full</b>    | <b>4.996</b> | <b>0.100</b> |

**DynaChip X1** streamlines CDMS workflow with online buffer exchange and direct nanoESI integration

# CDMS – AAV 5

*Enabling online AAV analysis from small sample volumes*



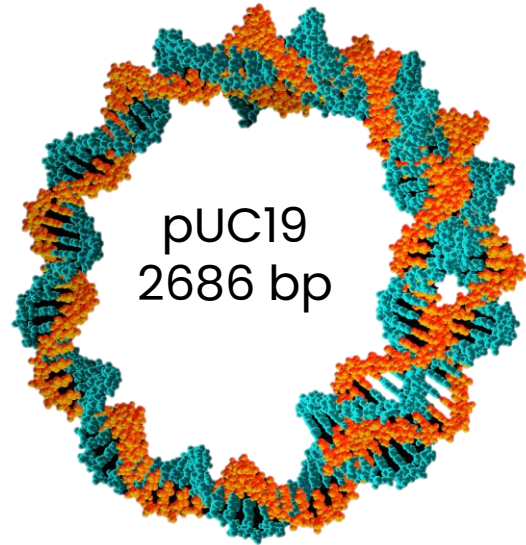
In collaboration with:



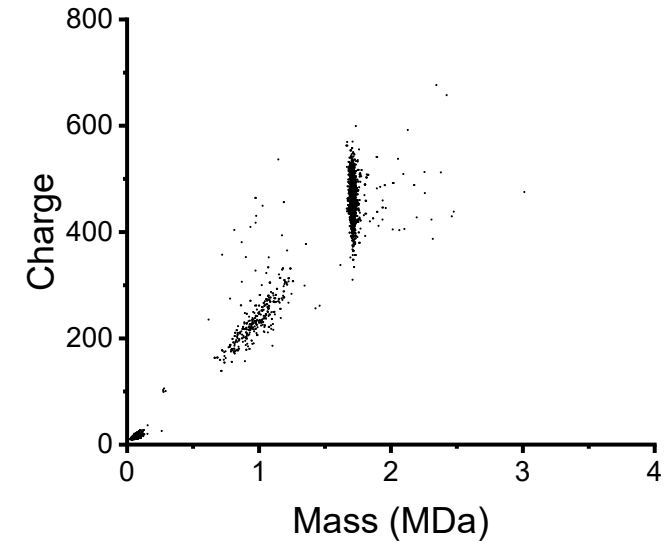
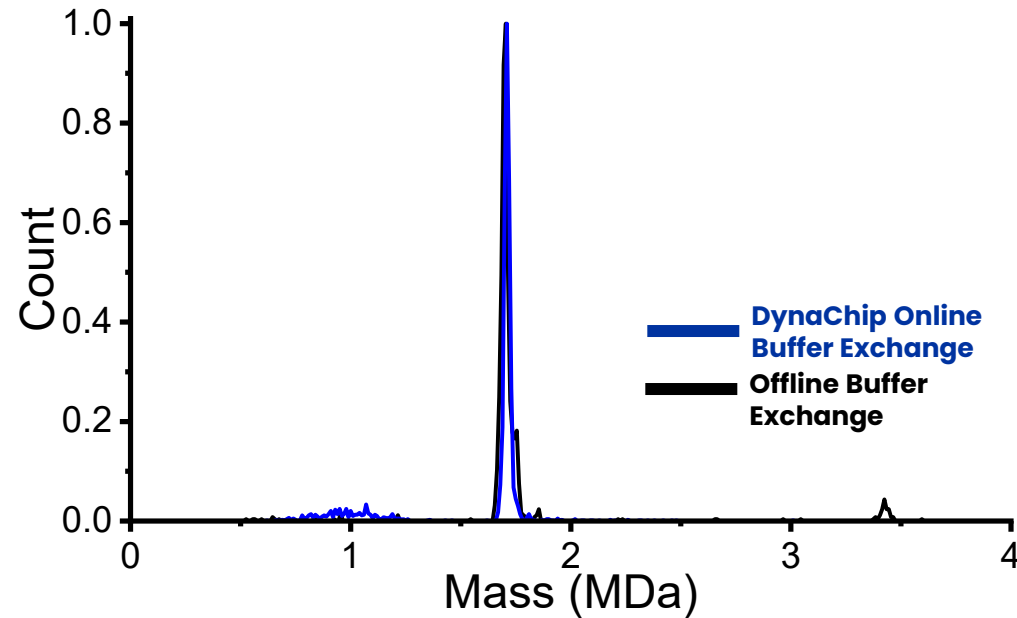
Data Acquired on: TrueMass CDMS

# Customization of Buffer for "Gentle" Ionization

*DNA Plasmid (pUC19) Exchanged into **20 mM Ammonium Acetate***

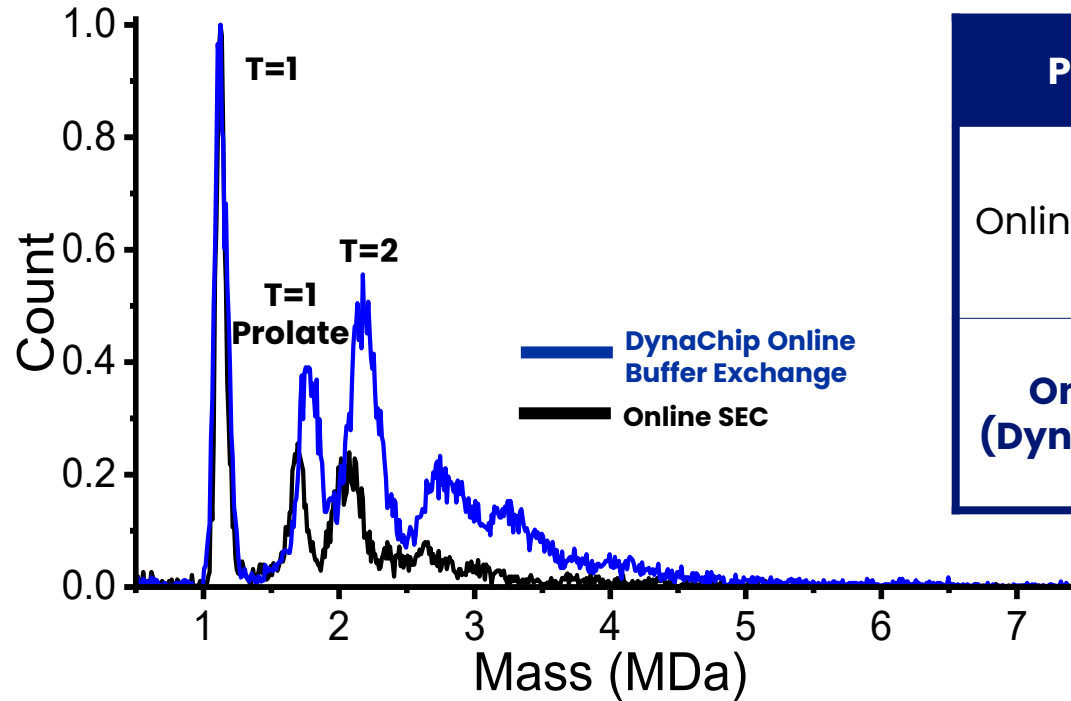
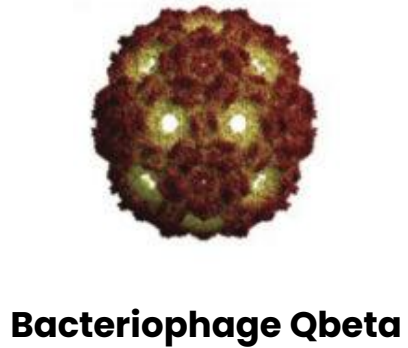


**DNA Plasmid (pUC19)**  
20 mM ammonium acetate



| Prep                     | Mass (MDa)   | FWHM (MDa)   |
|--------------------------|--------------|--------------|
| Offline (2x spin column) | 1.700        | 0.036        |
| <b>DynaChip</b>          | <b>1.711</b> | <b>0.028</b> |

# Comparing SEC-CDMS & DynaChip-CDMS



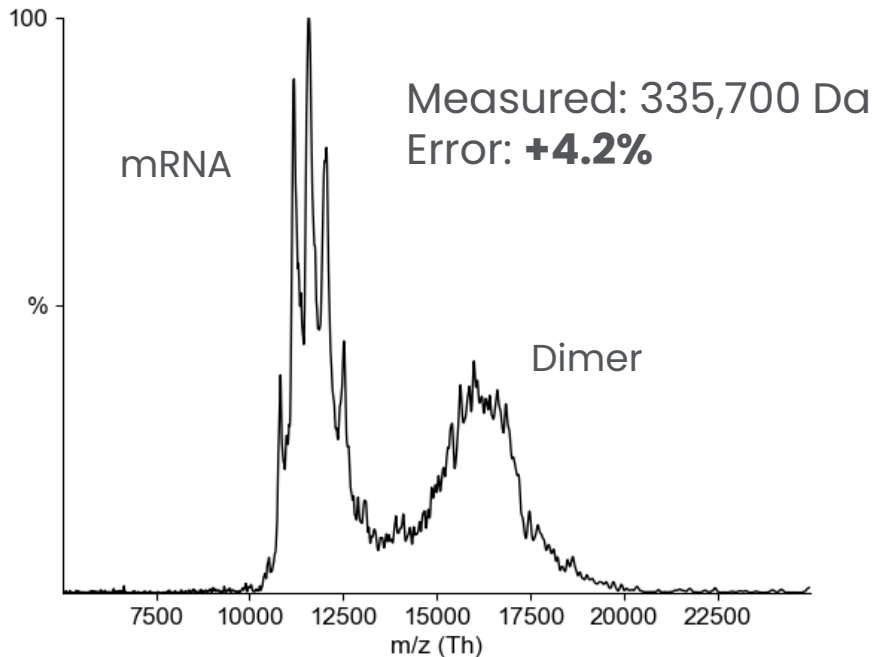
| Prep              | Structure   | Mass (MDa)   | FWHM (MDa)   |
|-------------------|-------------|--------------|--------------|
| Online (SEC)      | T=1         | 1.127        | 0.085        |
|                   | T=1 Prolate | 1.693        | 0.033        |
|                   | T=2         | 2.068        | 0.057        |
| Online (DynaChip) | T=1         | <b>1.127</b> | <b>0.081</b> |
|                   | T=1 Prolate | <b>1.781</b> | <b>0.050</b> |
|                   | T=2         | <b>2.181</b> | <b>0.102</b> |

**DynaChip XI** comparable results to SEC at higher throughput *while preserving sensitive biomolecular structures.*

# Efficient Desalting and Native mRNA Analysis

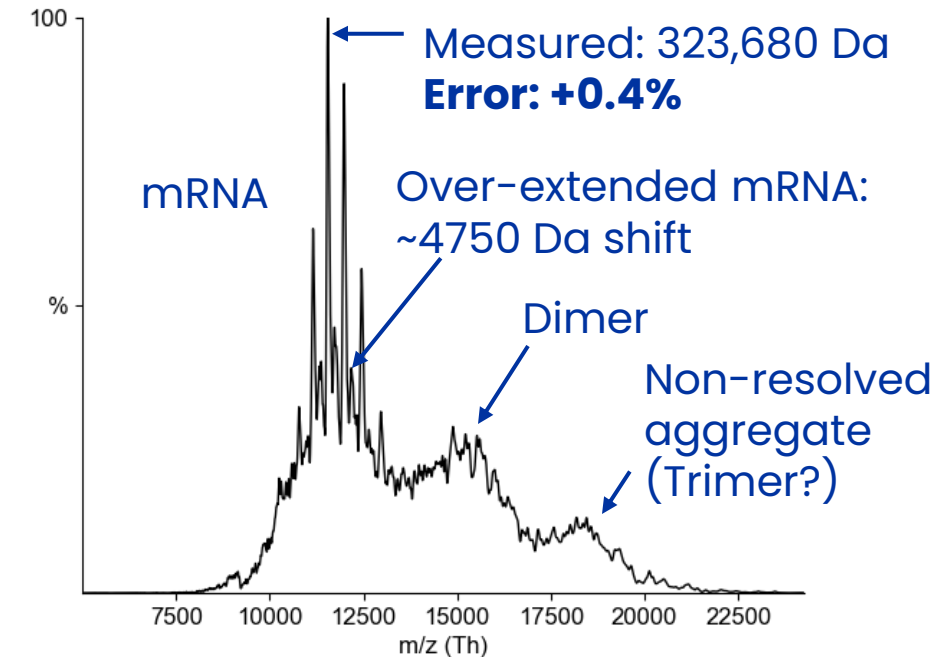
*Higher Spectral Quality and Richer Information*

**Dilute & Shoot:**  
**30 x dilution, long acquisition**



1 mg/ml EGFP mRNA in 1mM Sodium Citrate  
Sequence Mass: 322,316 Da  
Length: 997 nt

**DynaChip X1: 5 uL injection**



In collaboration with:

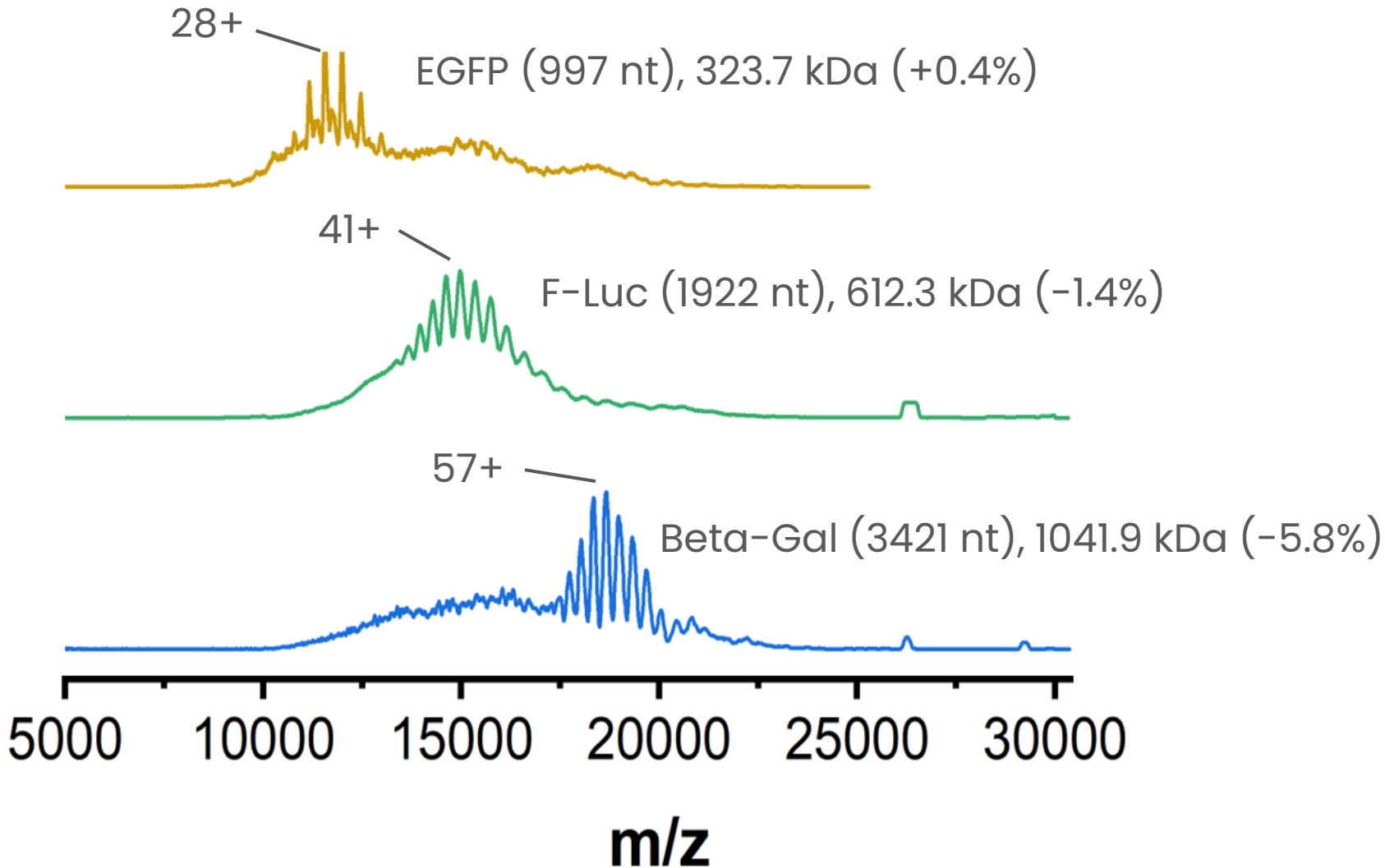


- ✓ Significant improvement in spectral quality and ESI intensity
- ✓ Significant removal of adducts as demonstrated +0.4% error in mass
- ✓ **Higher resolution revealing overextended population**

Data Acquired on: Thermo Q Exactive UHMR MS

# Rapid and Efficient Online Cleanup of mRNA

*Enabling Native Charge State Resolution of Megadalton Species*



- ✓ DynaChip enables resolution on RNAs in the 1000nt + range
- ✓ F-Luc and Beta-Gal measured mass were lower than predicted sequence mass.
- ✓ Lowered mass could be due to degradation—only possible tell that with sufficient clean-up and removal of adducts

In collaboration with:



Data Acquired on: Thermo Q Exactive UHMR MS

# Thanks to CASSS MS, Collaborators, Teammates

All technical data available online at  
[www.AndsonBiotech.com/Resources/](http://www.AndsonBiotech.com/Resources/)



None of this possible without  
Andson Biotech's Technical Team:

**Suzanne Bock** Microfab Engineer   **Aryan Dhingra** Mechatronics Engineer   **Carter Asef, PhD** FAS   **Casey Vantucci, PhD** Senior Scientist   **Austin Culberson, PhD** Principal Engineer

Thanks to our Collaborators!



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