

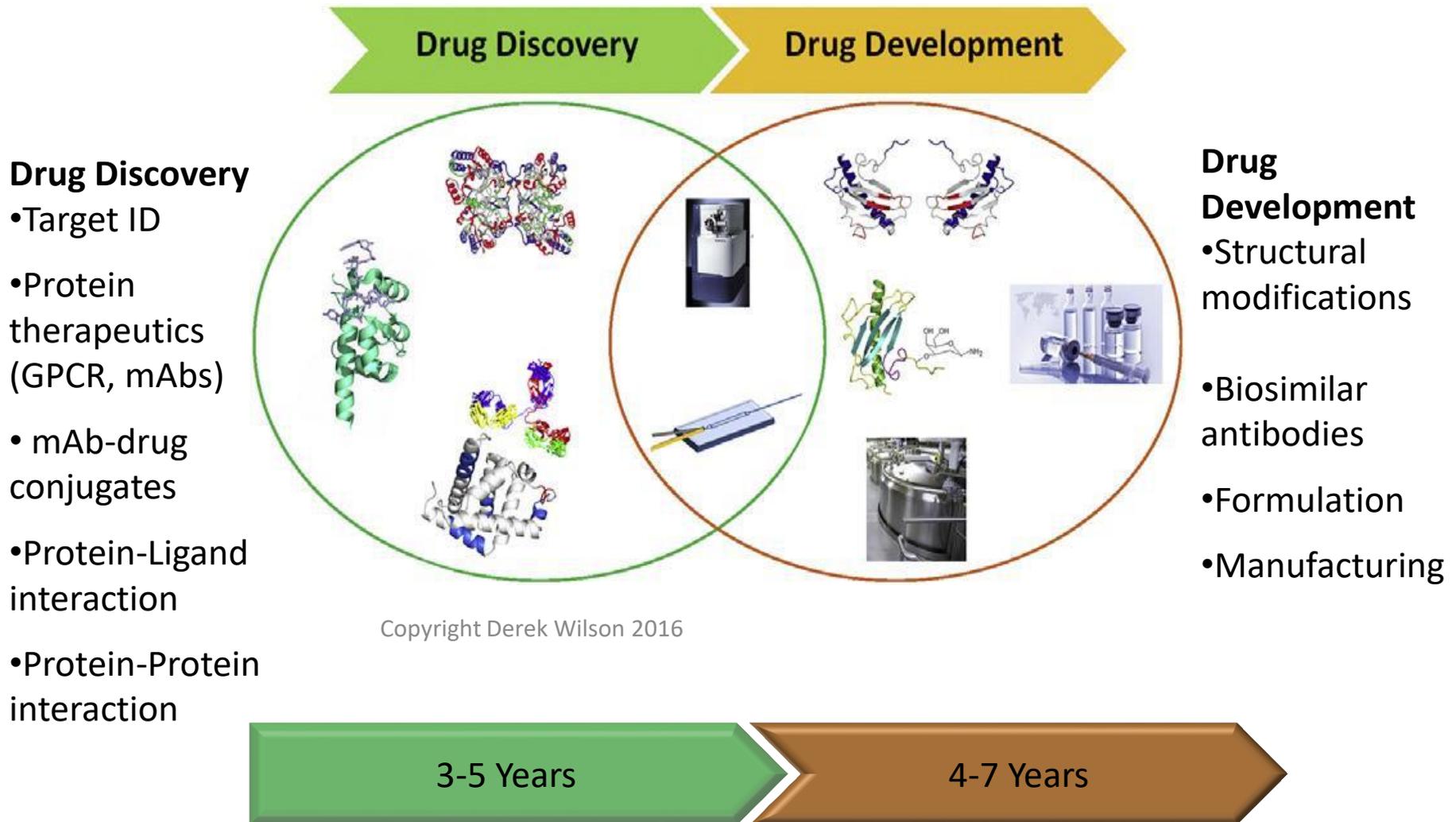
## Mark R. Chance

Center For Proteomics & Bioinformatics  
Case Western Reserve University  
Chief Scientific Officer, Neo Proteomics.

# Hacking Structural Biology for Drug Discovery Using Mass Spectrometry

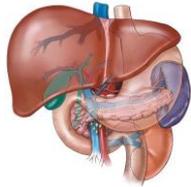
**Declared Conflict: CWRU technologies for footprinting and systems biology are licensed to Neo Proteomics Inc. where MRC is a shareholder and officer. MRC also serves on the Science Advisory Board of GenNext Technologies**

# Drug Discovery & Development

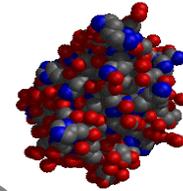


# Proteomics: a key enabling technology for structural & systems biology

mpvlsrprpw...



Where

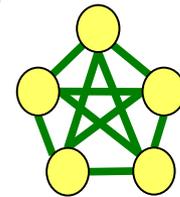
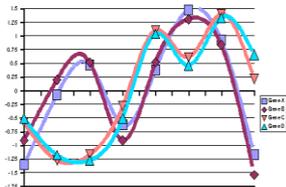


What



When

Relationships



## Proteomics From Molecules to Man



# Turn, Turn, Turn

## Advantages and Limitations of Major Structural Biology Techniques

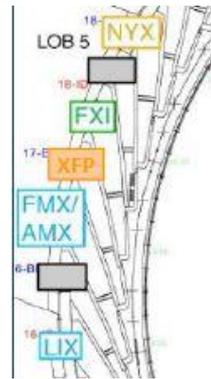
Resolution, size, and limits on amounts of material are approximate for 10-100K protein sizes.

Technique	Size (sample state)	Resolution Limits	Amounts	Notes
NMR	<100 kD (solution)	~3-4 Å	μmoles/milligrams	Requires labeled recombinant protein, disordered regions can be observed but may not be assigned.
Crystallography	Limited by crystal quality	< 1-3 Å	μmoles/milligrams	Mutant constructs necessary for many membrane proteins, disordered regions invisible. Gold standard for structural water
Cryo-EM: Single particle	>100 kD (vitrified ice)	Mostly >3 Å	nanomoles/μgrams	Resolution and size limits improving, best samples have symmetry, disordered regions invisible.
Cryo-EM: Tomography	Cells or tissues)	30-40 Å	thin sections/individual cells	Resolution improving; captures large-scale spatial organization in cells.
SAXS	> 10 kD (solution)	> 20 Å;	nanomoles/μgrams	Native material can usually be used, (similar to FP samples)
Footprinting: HRF-MS [and HDX-MS]	HRF-MS: No limit [<100K for HDX] (solution)	Peptide to single-residue (single base for NA)	picomoles/nanograms	Native material can usually be used (both), absolute surface area can be estimated (HRF), disordered regions visible (HRF). Studies in cells/tissue possible (HRF).

CWRU at  
NSLS-II

MS-Footprinting/HDX  
•Local conformation change  
•Binding interfaces

A  
Structural  
Biology  
Village



High-Resolution  
Structure & Models  
(Cryo-EM, & MX)  
•Medium to High resolution  
•Local and global

Integrative  
Approach to  
Structure  
Determination

Small Angle  
X-ray Scattering/EM/  
Native MS  
•Global Conformation  
•Large Complexes  
•Identity & Stoichiometry

Computational  
Approach  
•Homology Modeling  
•Docking  
•Molecular Dynamics

Other Biophysical Data  
•Crosslinking, mutagenesis

# CSB at National Synchrotron Light Source-II



NSLS-II Floor Area



FMX, AMX Crystallography



XFP Footprinting



SCHOOL OF MEDICINE  
CASE WESTERN RESERVE  
UNIVERSITY

*Center for Synchrotron Biosciences*



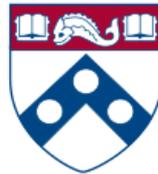
Albert Einstein College of Medicine  
OF YESHIVA UNIVERSITY



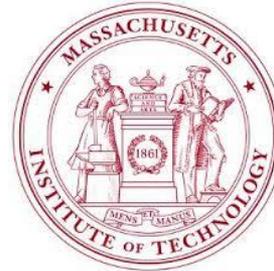
Yale University  
School of Medicine



Stanford  
University



Perelman  
School of Medicine  
UNIVERSITY OF PENNSYLVANIA



JOHNS HOPKINS  
UNIVERSITY



Sharing Resources  
Award 2017



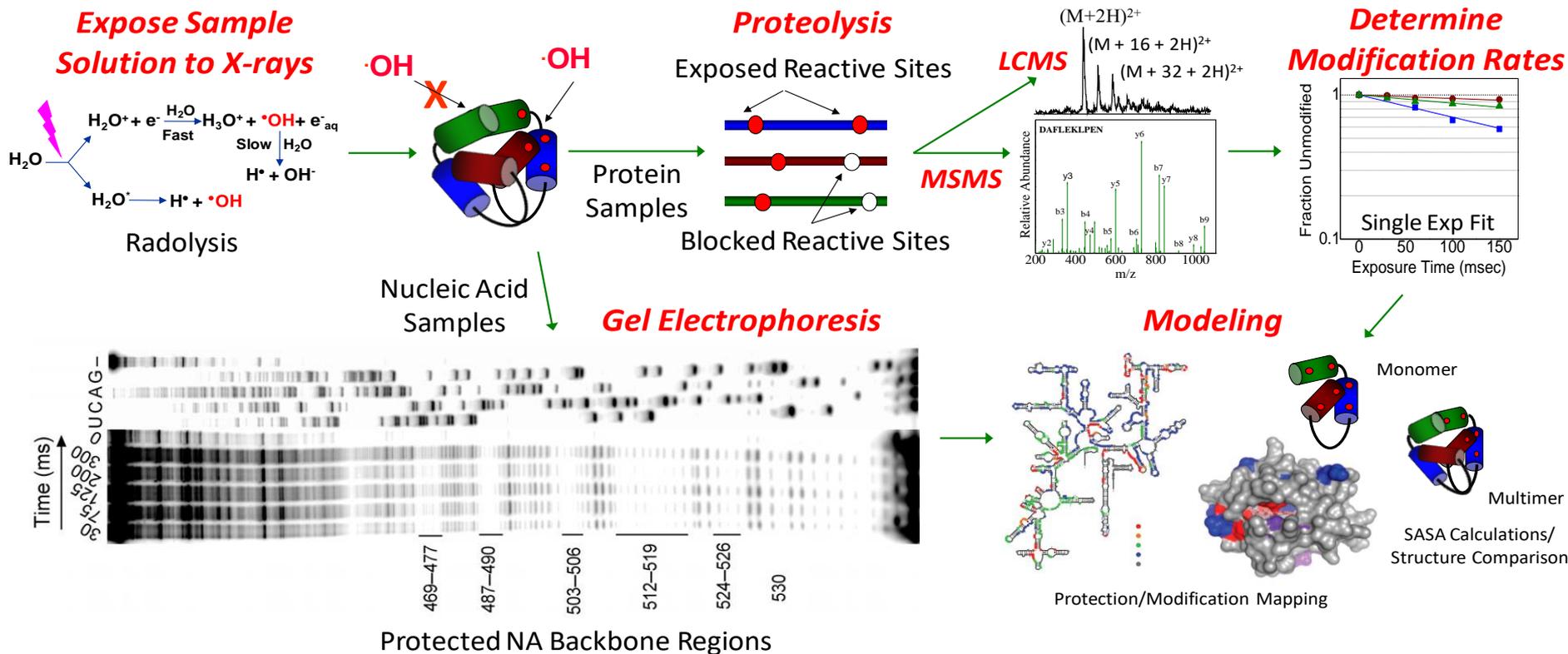
SCHOOL OF MEDICINE  
CASE WESTERN RESERVE  
UNIVERSITY

*Center for Proteomics and Bioinformatics (2005)*

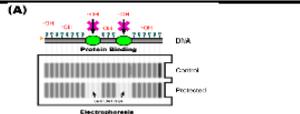
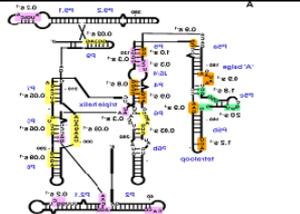
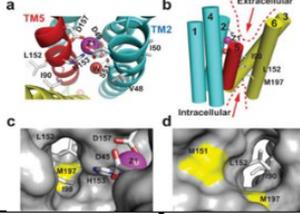
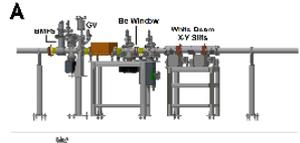
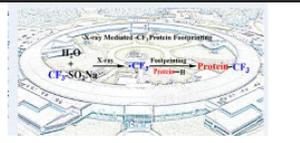
*Center for Synchrotron Biosciences (1994)*

# What is Footprinting?

- Chemical alteration of a macromolecule in solution
  - Sensitive to surface accessibility
    - Identifies binding sites
  - Nucleic Acids Footprinting: 1970
    - Protein Footprinting: 2000s
    - Hydroxyl radical #1 reagent



# Thirty Years of Synchrotron Footprinting Development

Beamline	Years	Key Outcomes	Graphic/Notes	Key Papers
NLSL X19C	1995-1996	Proof of concept as General User		JMB 1997
NLSL X9A	1996-1999	Time-resolved RNA footprinting		Science 1998
NLSL X28C	2000-2014	Protein footprinting (steady-state and time-resolved). Focusing mirror innovation.		PNAS 2009 Nature 2014
ALS (5.3.1 /3.2.1)	2014-2016	Technology and equipment. Mirror and new continuous flow methods.		Comm. Biology 2022
NLSL-II 17-BM	2016-2024	Ultra-high flux density / high-throughput methods		Cell 2019
NLSL-II 17-BM	2024-2029 Proposed	Continued innovation (Multiplex chemistry/direct dosimetry/MS onsite)		

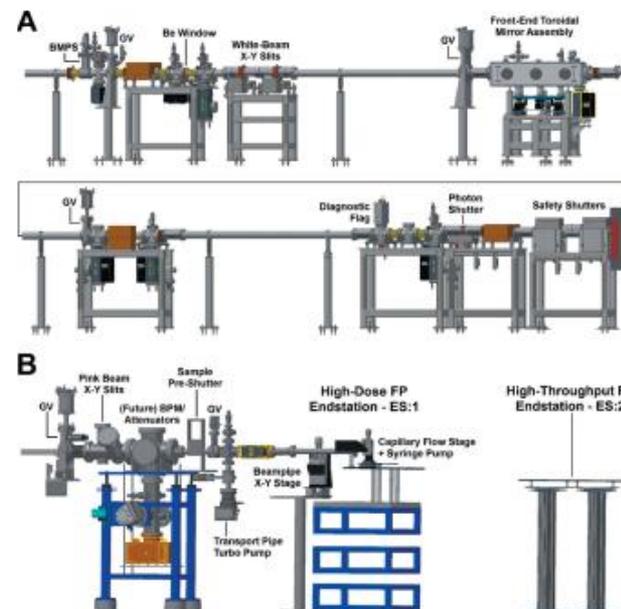


# Generation of OH radicals from radiolysis

## Beamline BM-17- X-ray Footprinting of Biological Materials Beamline



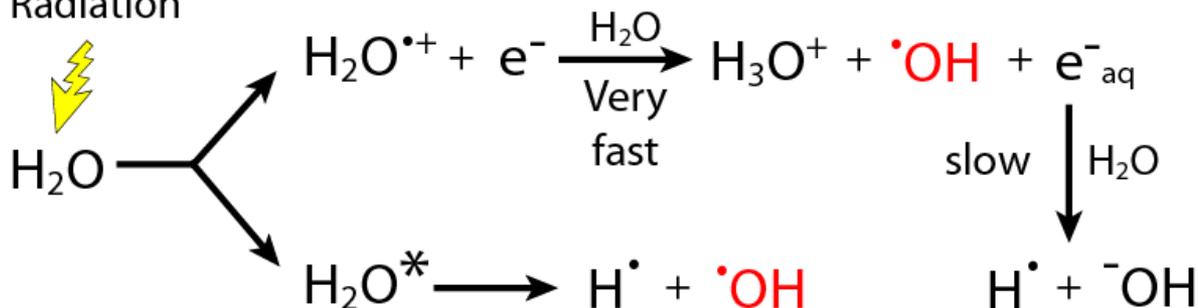
MAJOR  
RESEARCH  
INSTRUMENTATION



## Radiolysis of water

100eV = 2.87 OH radicals

Ionizing  
Radiation



OH radicals also produced by: Photolysis/Fenton/Plasma

## GenNext FOX™ Flash Oxidation Technology

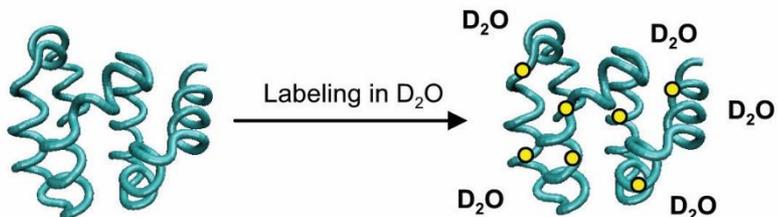
- FOX system has photolysis by high energy plasma lamp.
- Peroxide and sample mixing automatic
- Real-time radical dosimetry determines effective radical yield and enables adjustment for scavenging.
- Product collector provides for automated collection of labeled protein

Benchtop to Beamline  
supported ecosystem



# Molecular Footprinting of Proteins

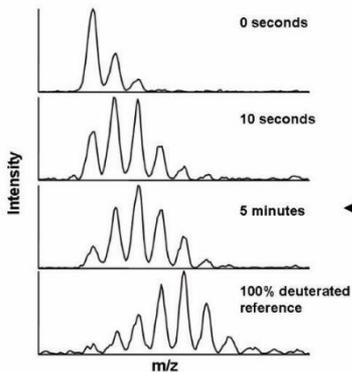
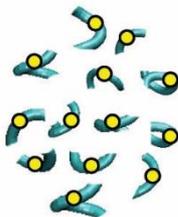
## HDXMS



protein

Quench

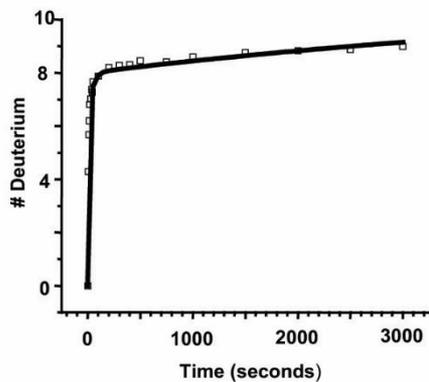
Digestion



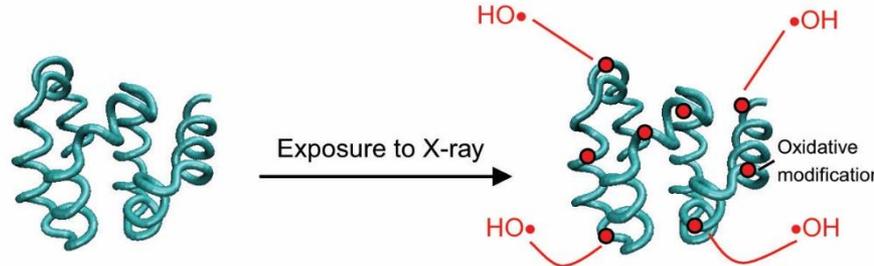
Calculation of deuterium content

Isotope pattern

Determine HX rates

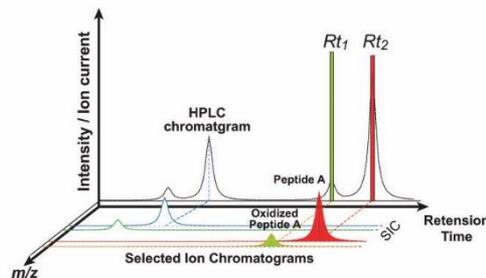
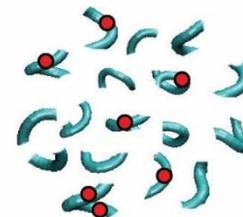


## PFMS



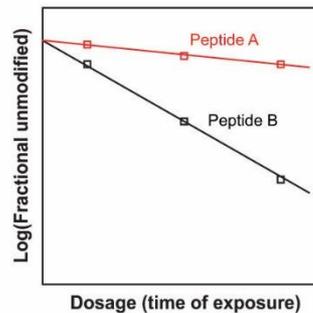
protein

Digestion

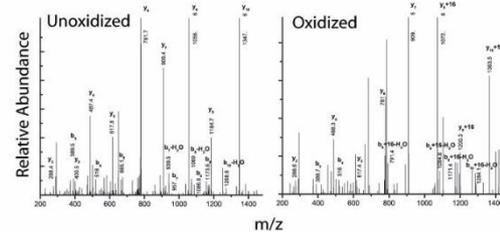


HPLC-MS

Calculation of oxidation rate



MS/MS identify the sites of oxidation



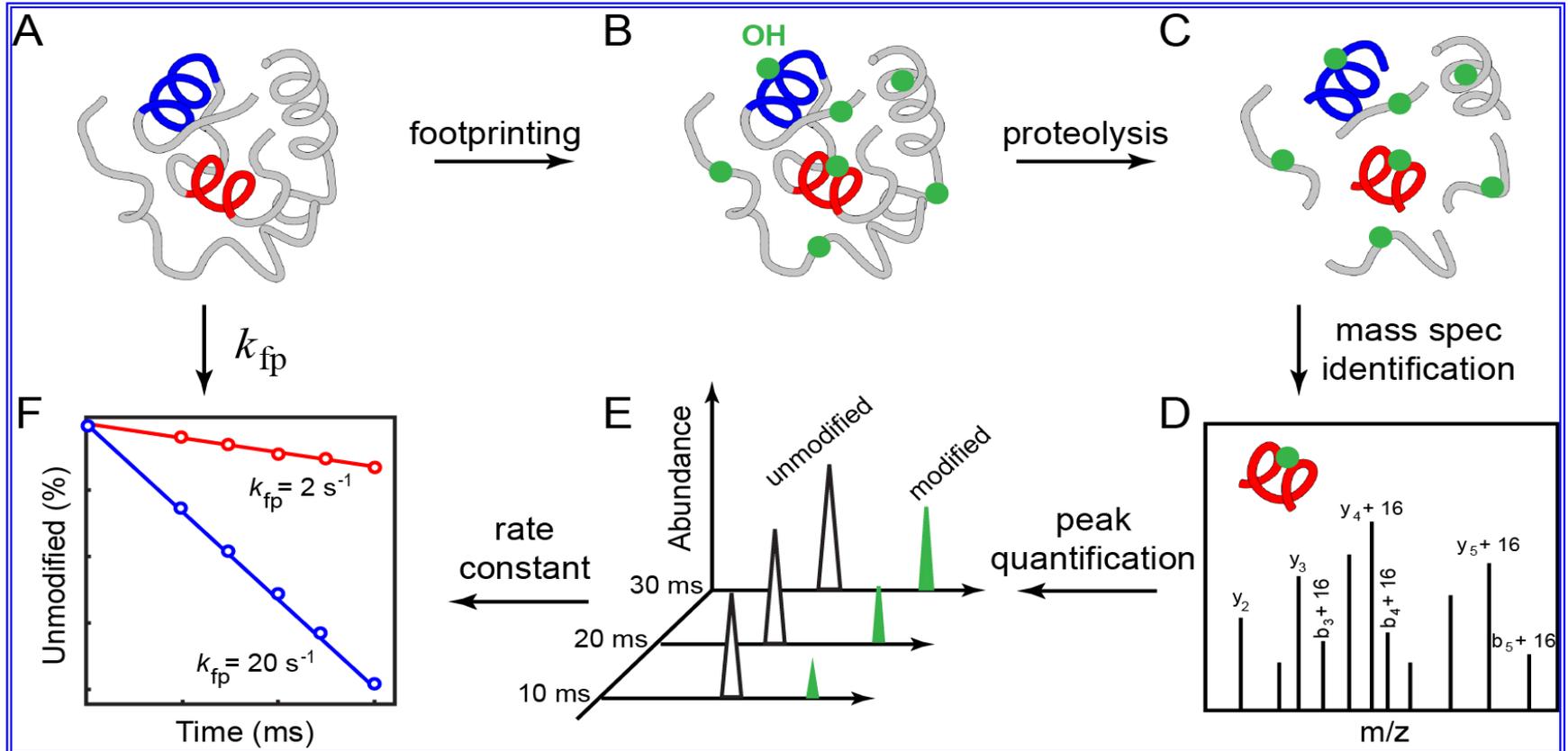
QVYQFEYESLLR  
+16 oxidation

# Complementarity of Footprinting Approaches

- **HDX** unsurpassed for assessing **protein backbone secondary structure** in solution (vs NMR). Drawbacks include size limitations due to need for pepsin cleavage, structural resolution at peptide level. Can use conventional ion-trap instruments.
- **Covalent labeling** reports **solvent accessibility** at **side chain** level resolution, large or complex macromolecules and membrane proteins feasible, many proteases and chromatographic methods. Requires Hi-res instruments.
- **Standard protocols are at peptide level**

# MEDIUM- RESOLUTION FOOTPRINTING STRATEGY

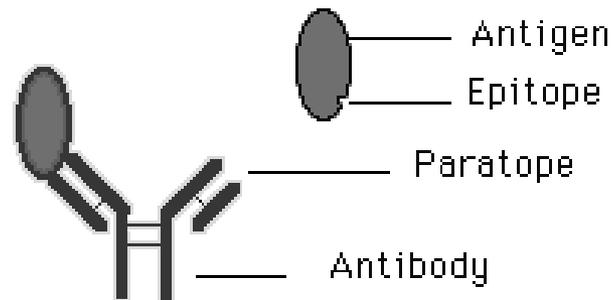
## Peptide level analysis



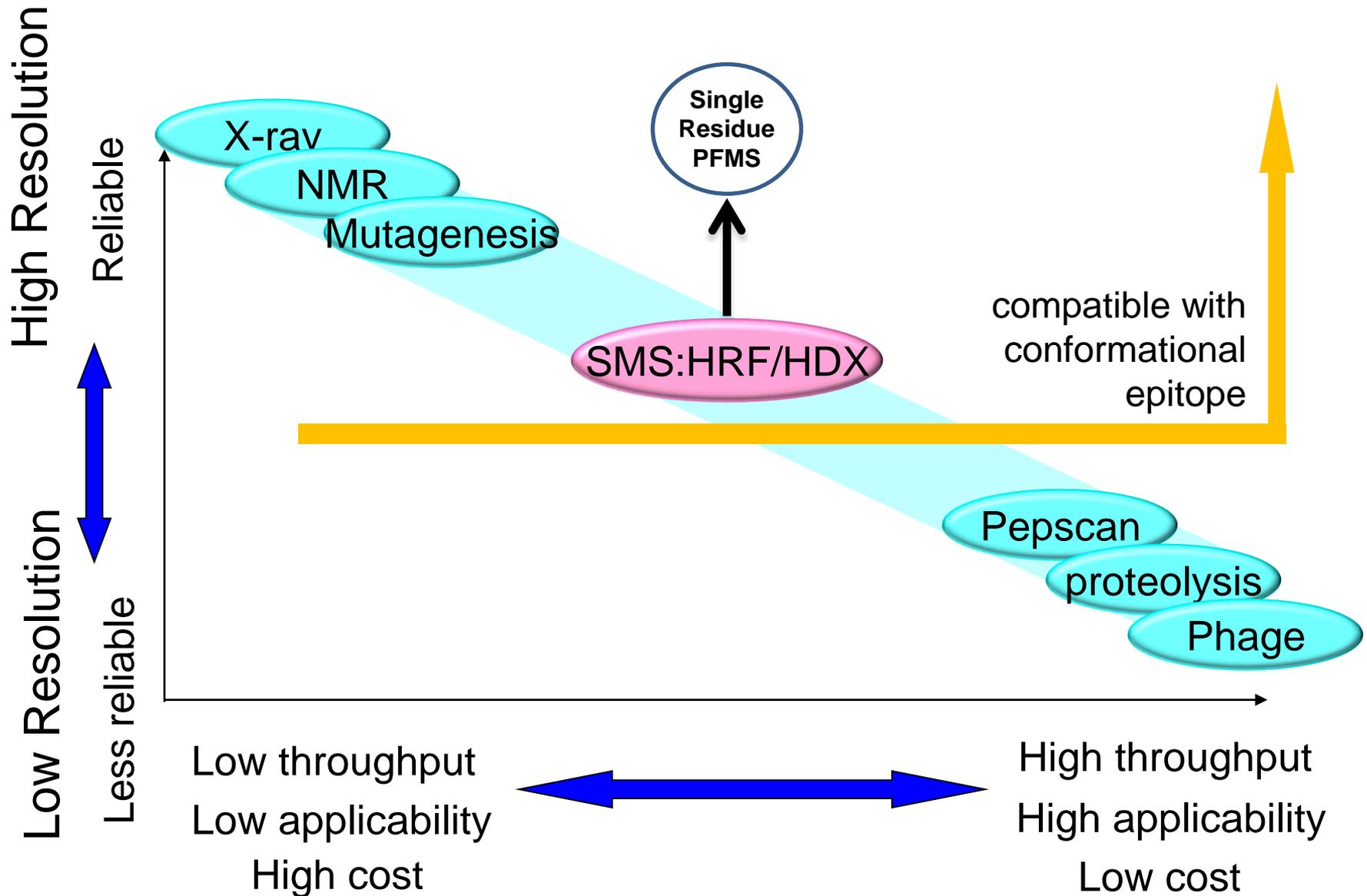
Courtesy of Sichun Yang

# Mass Spectrometry for epitope mapping and structure assessment of mAbs

- **Antigen epitope mapping** by structural mass spectrometry provides novelty and patentability of for mAb substance.
- IP protection can both uniquely establish IP relevant to antigen binding motif and/or block competitors.
- **Paratope mapping** important for re-engineering antibody



# Structure assessment/epitope mapping



Courtesy of Yoshi Hamuro

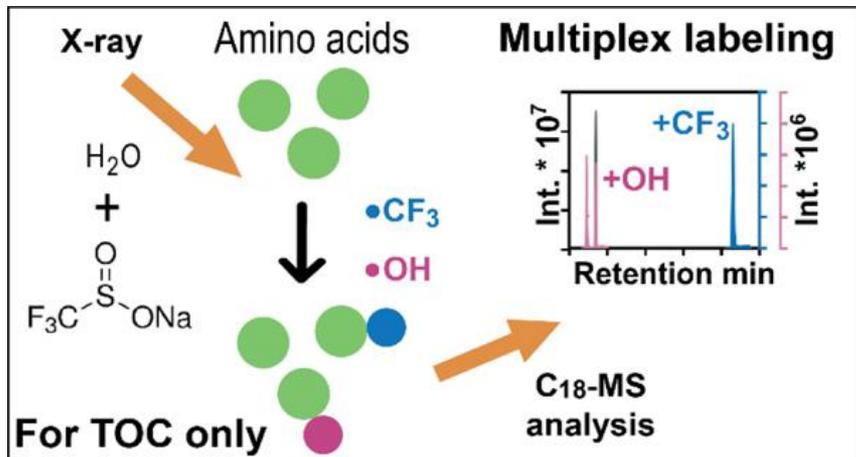
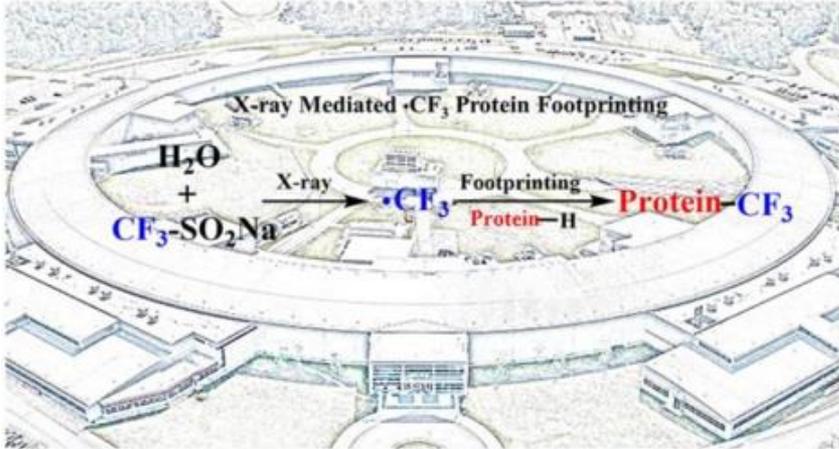
## Enhancing Structural Resolution of PF with chemistry

Reagent chemistry	Principal target residues	Mass shifts
OH radical	Aliphatic, aromatic, Met, Cys	Aliphatic: +16, +14 Aromatic/sulfur containing: +16, +32, +48 Arg: +16, -43; Asp, Glu: +16, -30; His: +16, +5, -22
EDC/GEE	Asp, Glu, C-terminus	+57, +85
Methylene carbene	All residues, minimal specificity	Variable (+14 methylene addition)
Carbethoxy	His, Lys, Tyr, Ser, Thr, Cys	+72

Typically Up to 20% of residues labeled

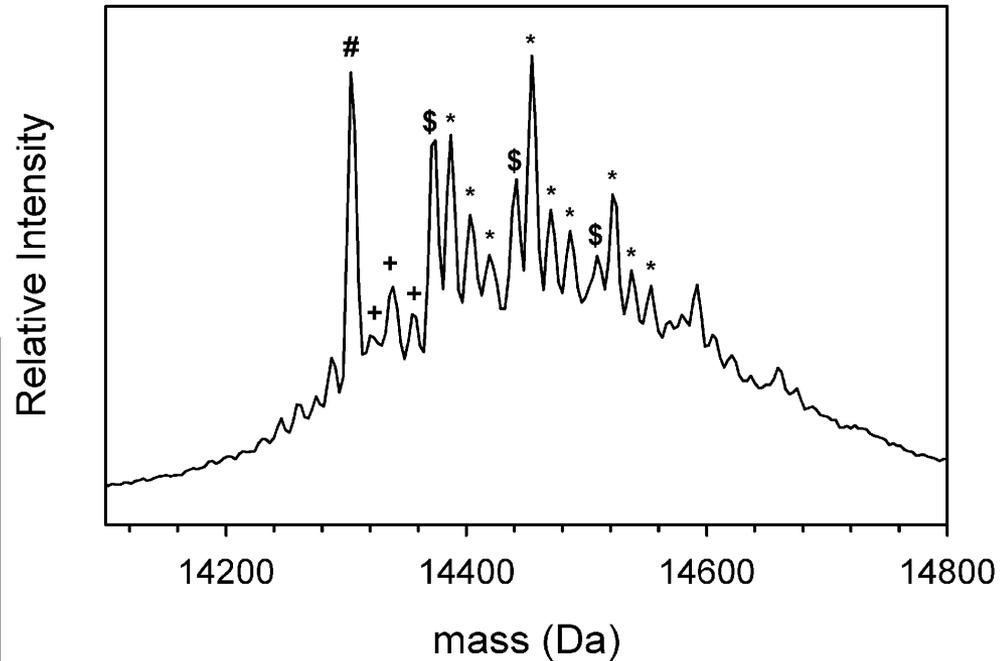
Kiselar & Chance, *Ann. Rev. Biophys.* (2018)

# CF3-OH Multiplex Chemistry



## Intact MS of lysozyme

Charge state +10; 5.7 OH (+) and 0.5 CF3 (\$) per molecule average, \*-mixed CF3-OH labels



Increases density of side chain labeling by 50-150% depending on peptide.

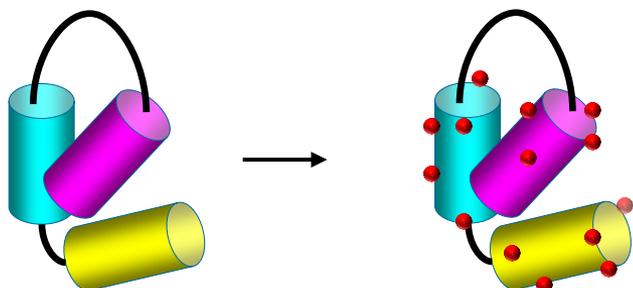
# HIGH-RESOLUTION FOOTPRINTING STRATEGY

## Residue level analysis

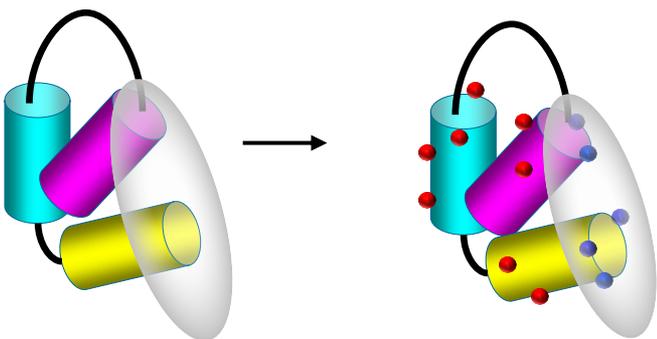
X-ray/Light/H<sub>2</sub>O<sub>2</sub>  
H<sub>2</sub>O-CF<sub>3</sub> → •OH

Flexible  
Proteolysis

UPLC-MS  
analysis

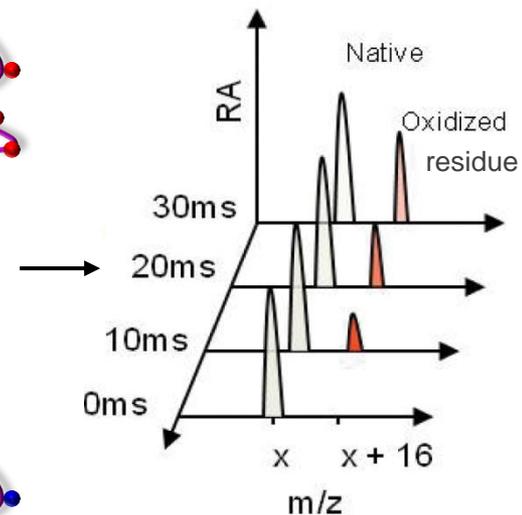
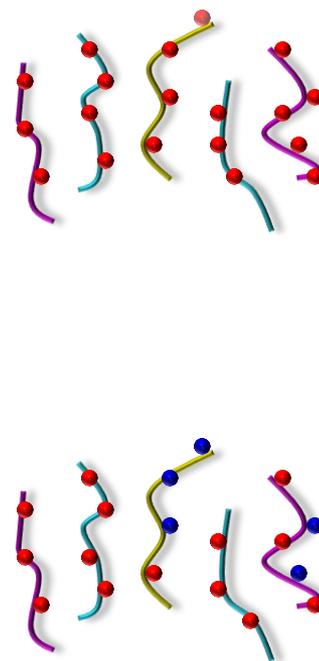


Protein (A state)

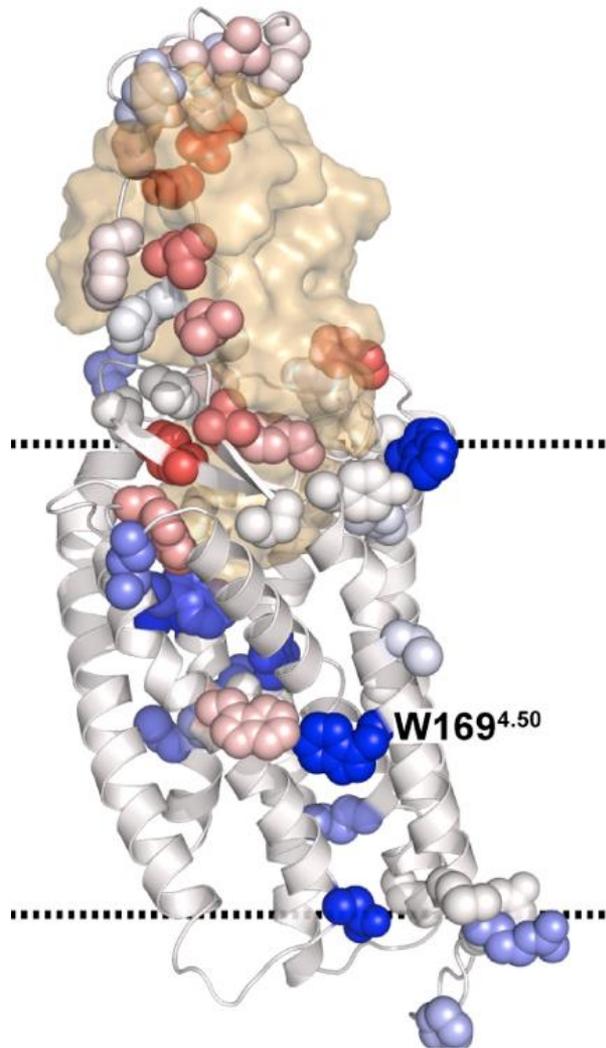


Protein + Ligand (B state)

Trypsin  
Asp-N  
Pepsin

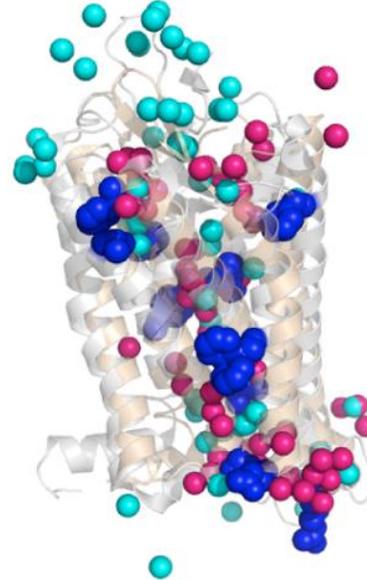


# Chemokine (CXCL12) signaling in 7TMs (ACKR3) occurs through conserved paths mediated by structural waters



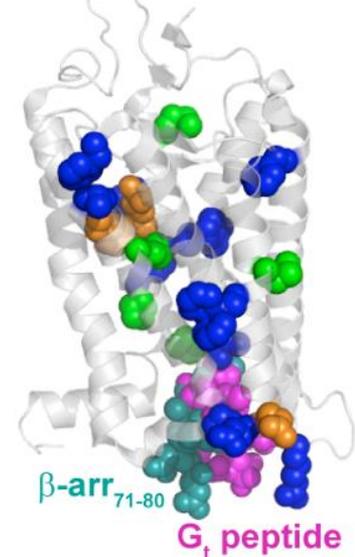
## b Water molecules

Active Rhodopsin  
Active  $\mu$ -OR



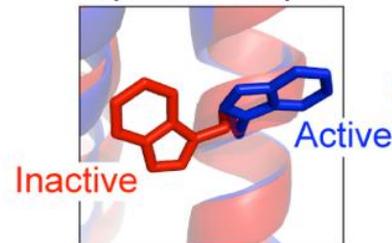
## c Increased oxidation rates in active state

Rhodopsin  
5HT4  
ACKR3

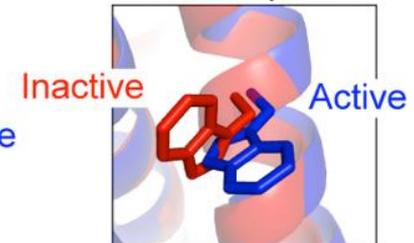


## d

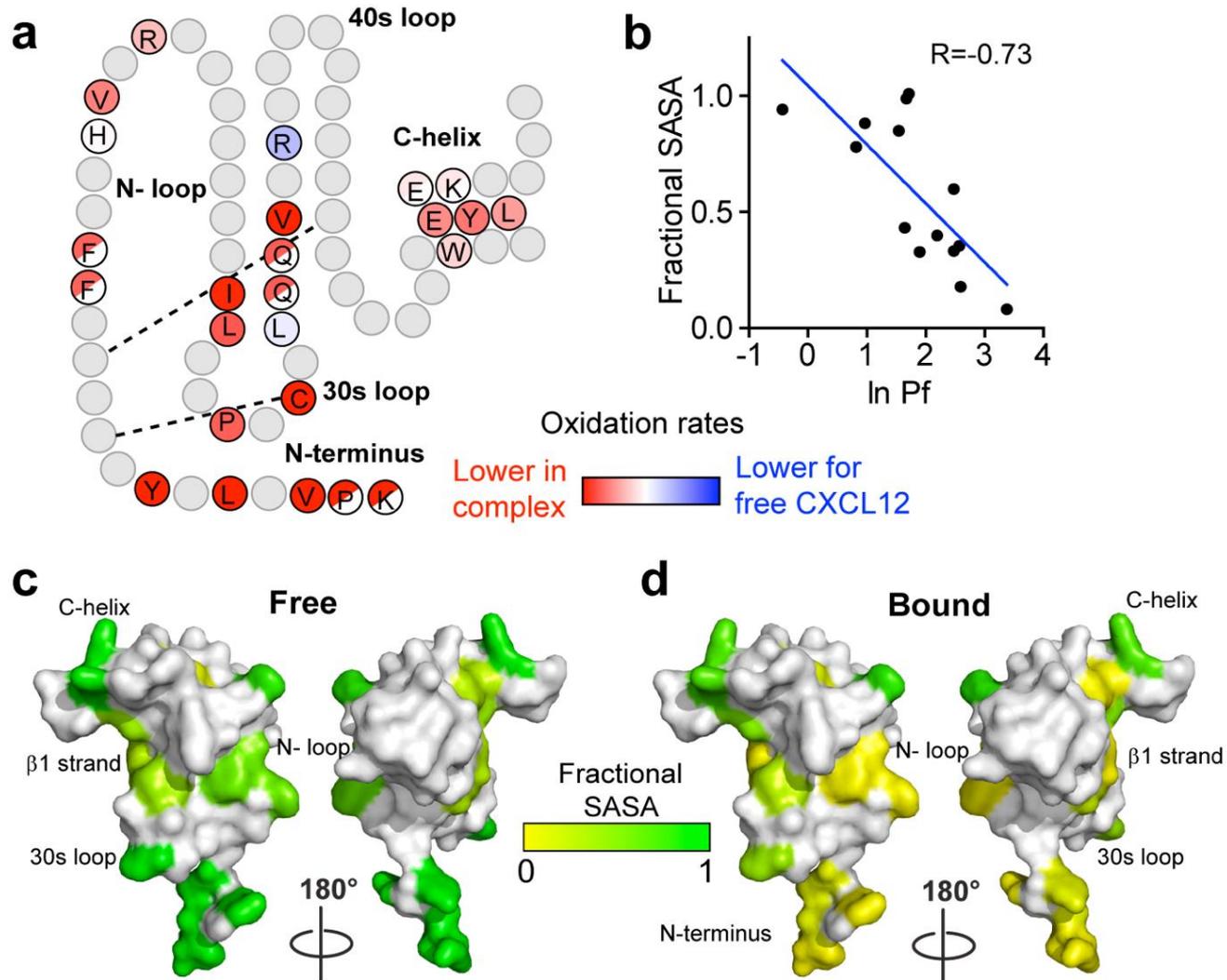
Opioid receptor



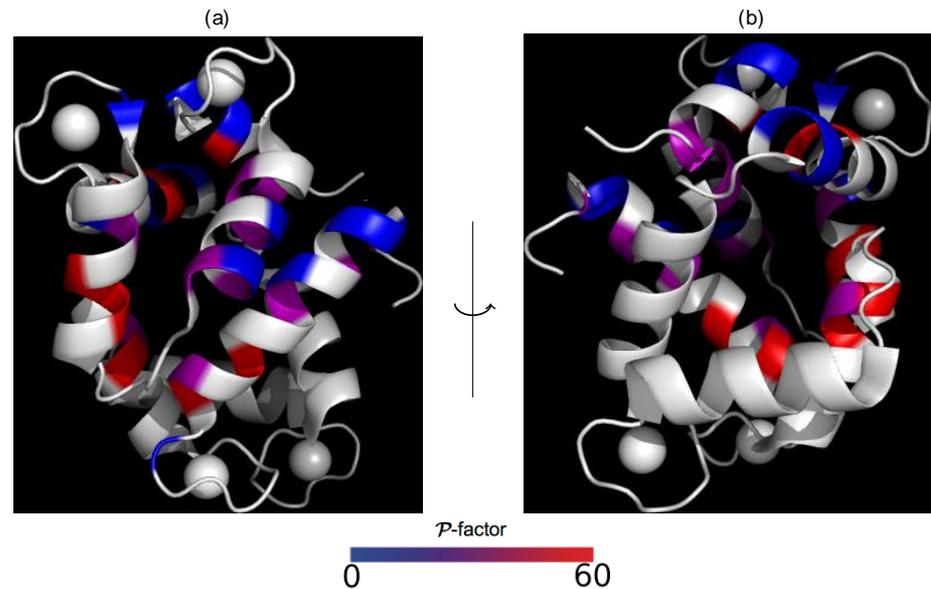
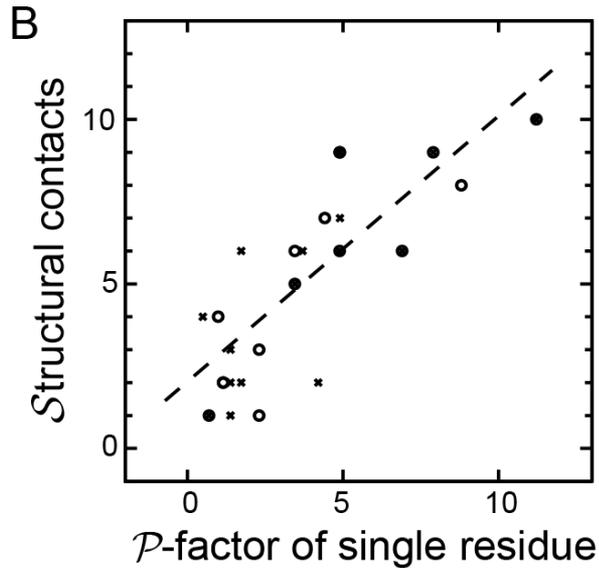
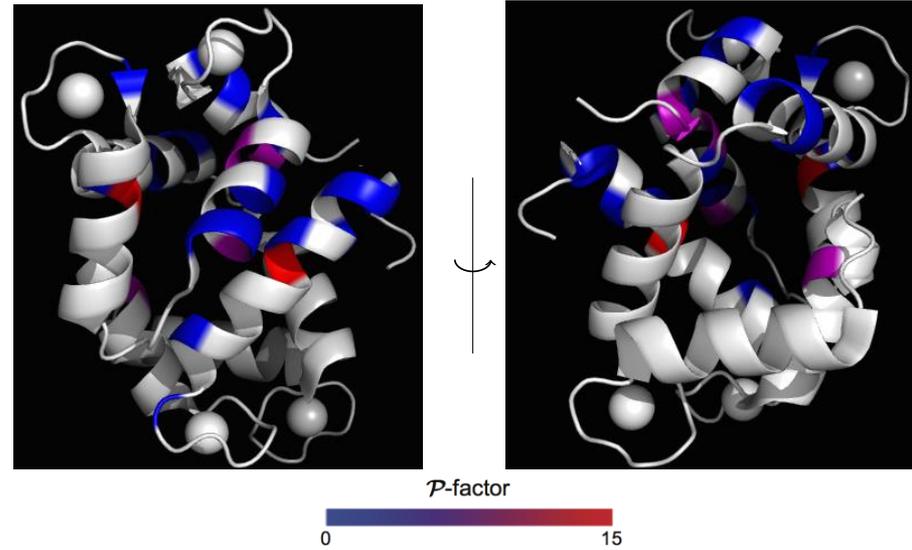
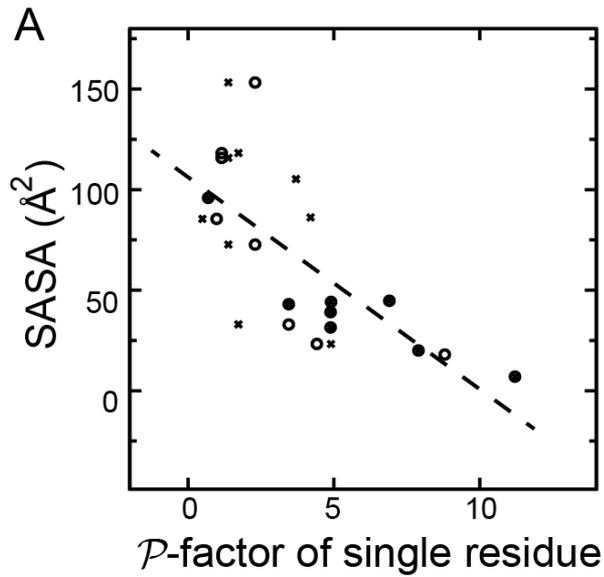
Rhodopsin



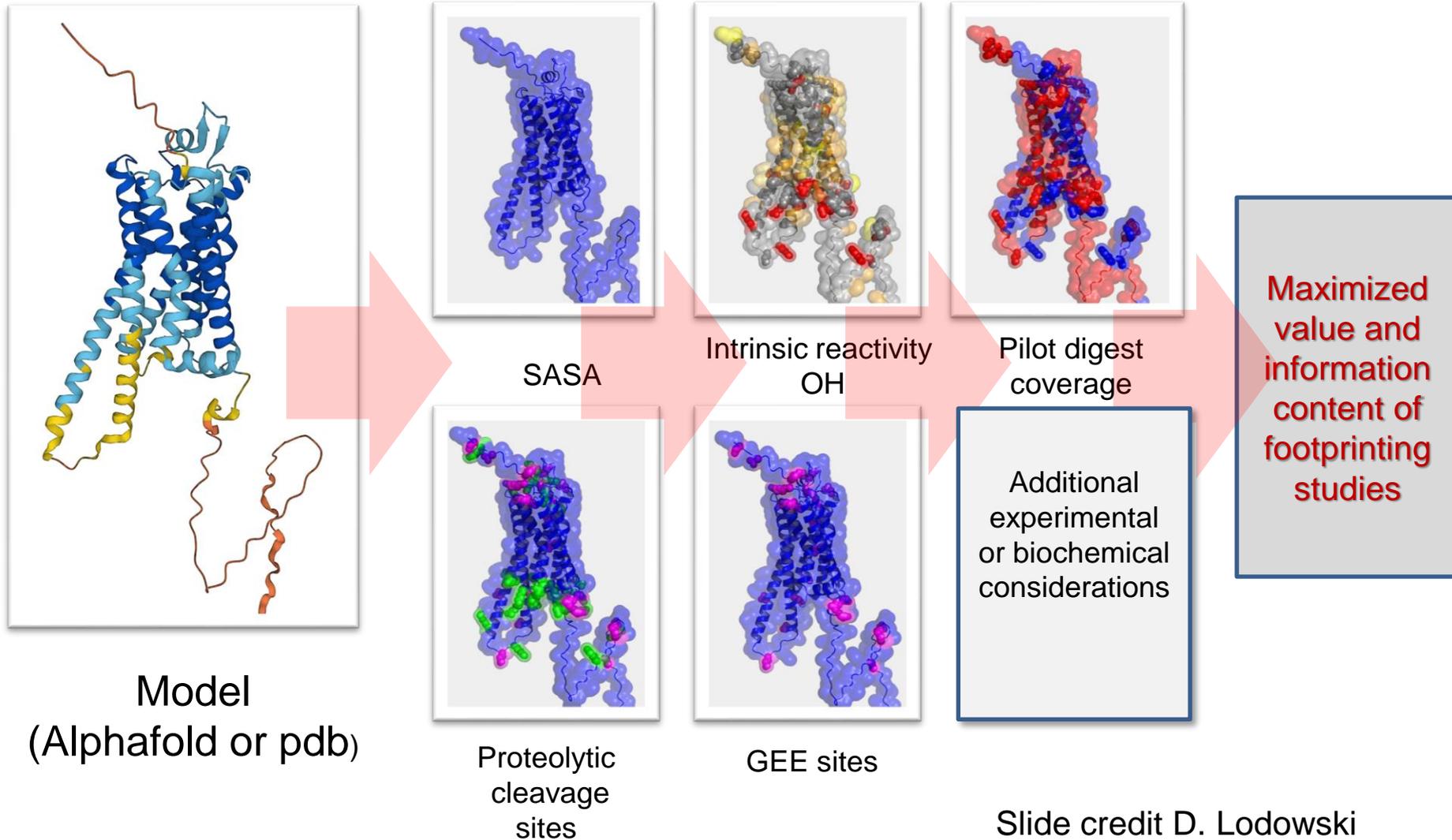
# Binding Epitopes of Chemokine Signaling: SDF-1 and CXCR7



# Calmodulin Structure Assessment



# Structurally aware experimental design



# Single residue resolution epitope protection map using HRPF

Antigen protein surface  
representation  
showing detected  
residues color coded by  
PR value.

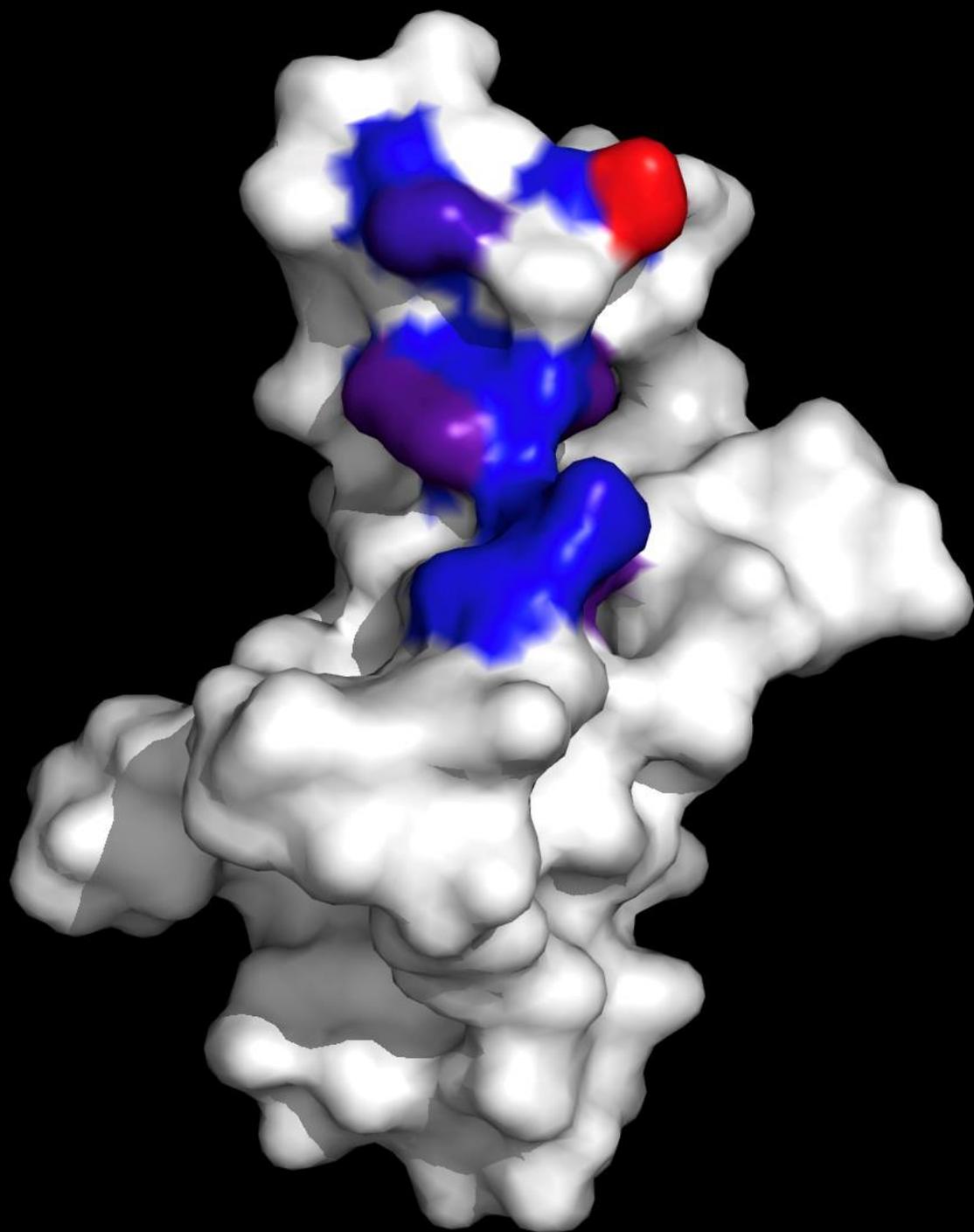
PR of 1.2-1.4 (purple),  
PR 1.4-1.9 (blue) and  
PR  $\geq 2.0$  (red).

Residues in white have  
PR < 1.2.

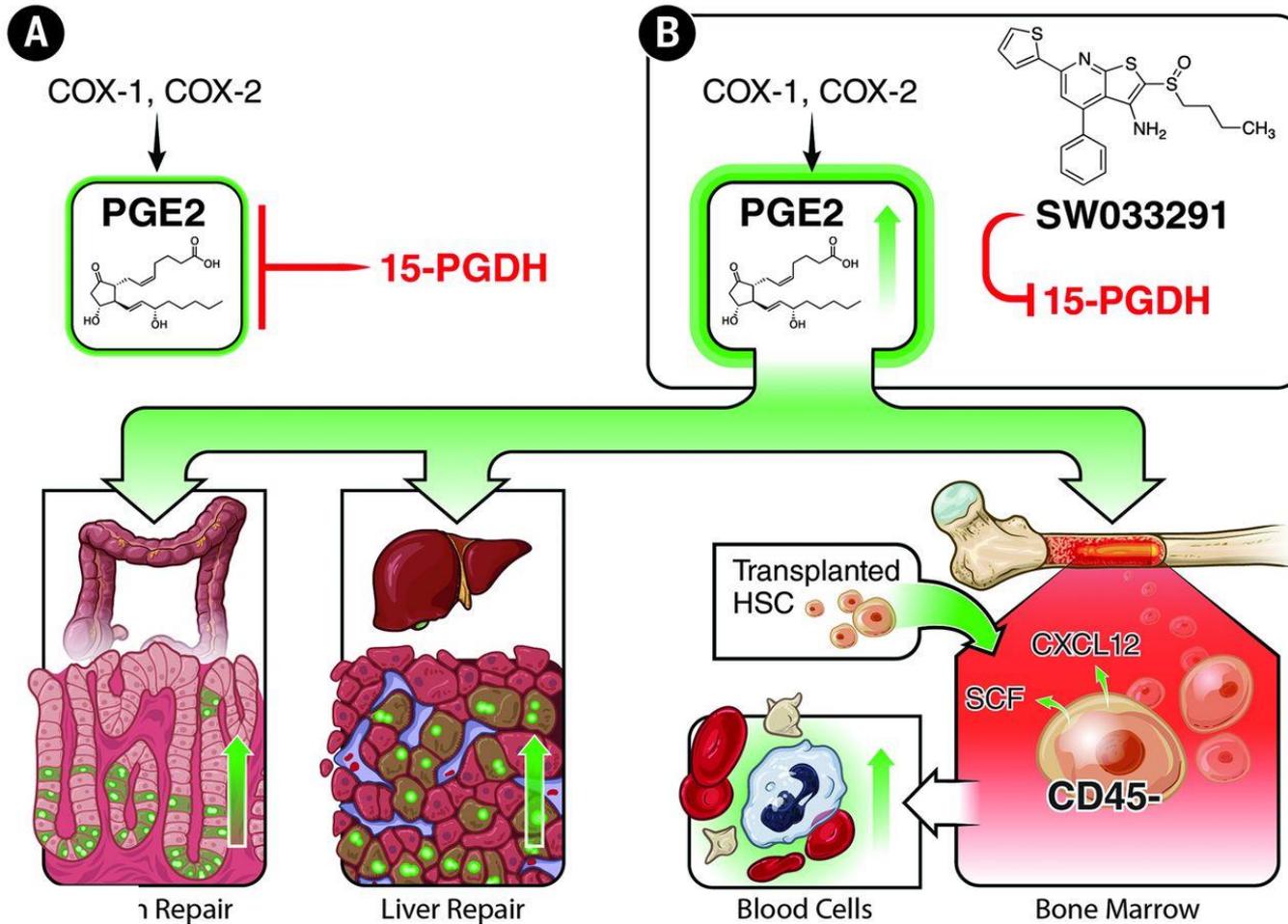
The identified “patch”  
of protections reveals  
a conformational  
epitope



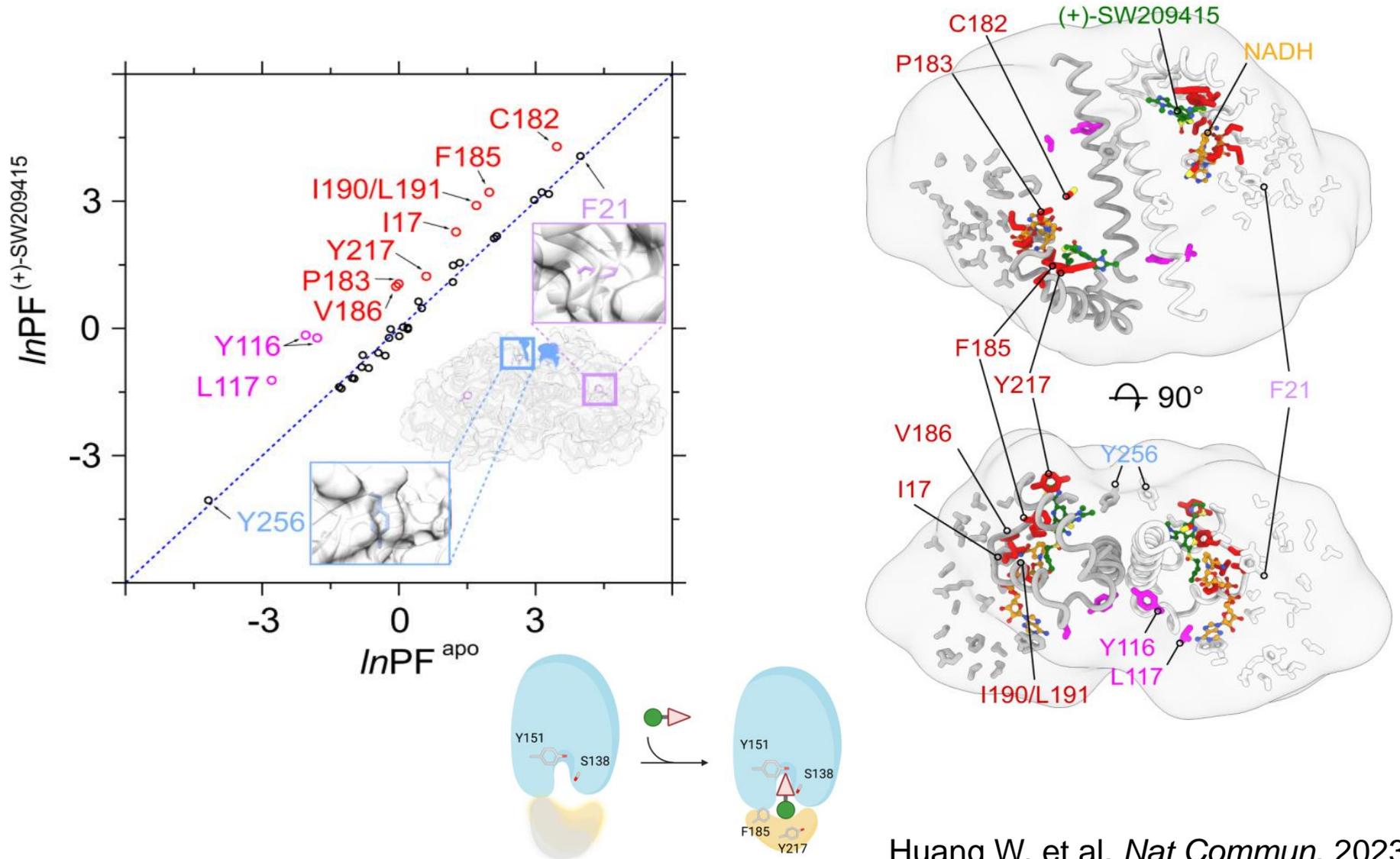
NeoProteomics



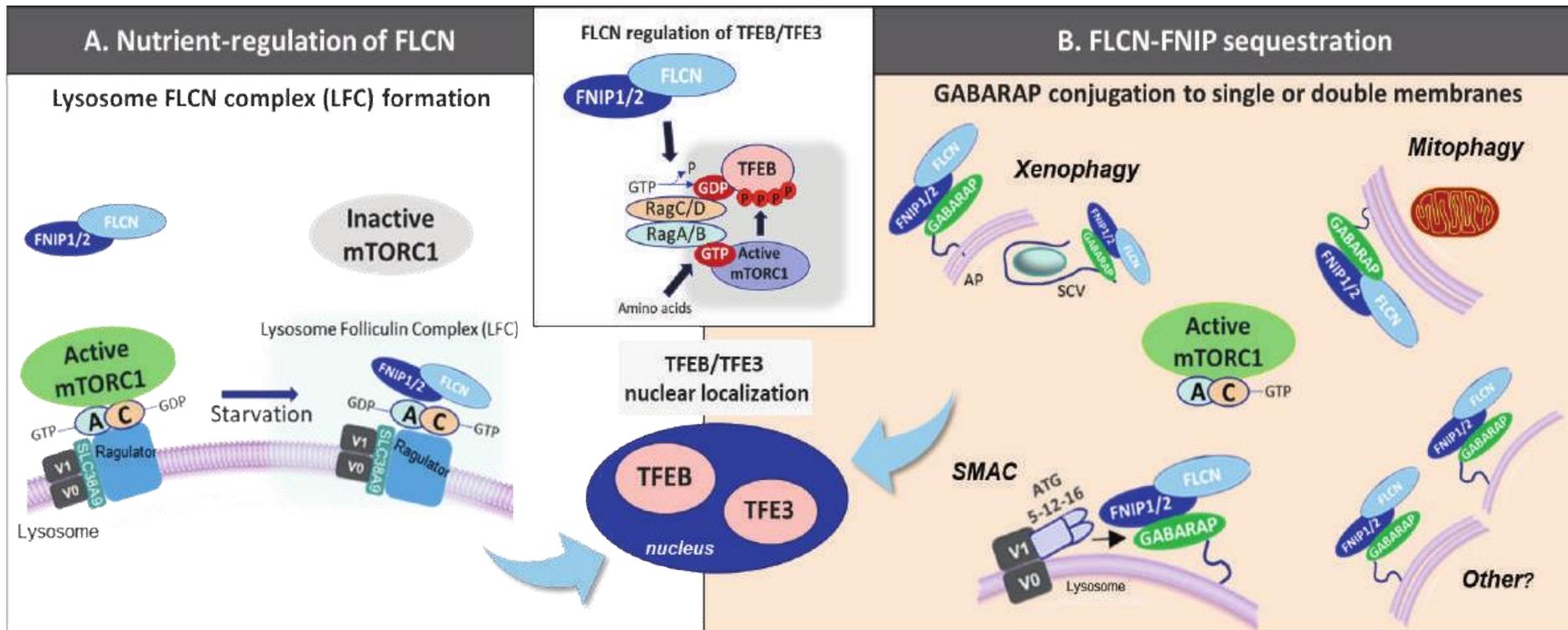
# SOTA Applications of Footprinting: Inhibition of the prostaglandin-degrading enzyme 15-PGDH potentiates tissue regeneration



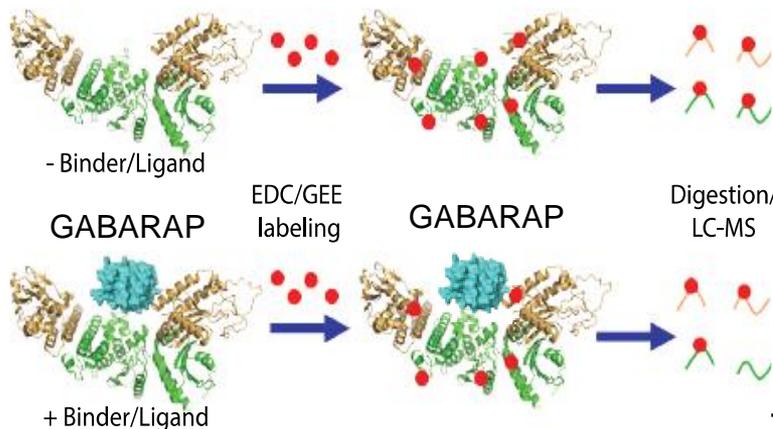
# Understanding the basis for nanomolar binding of SW2109415 using footprinting driven structural analysis



# SOTA Applications of Footprinting in Industry: Ternary complex formation of GABARAP binding to the FLCN/FNIP binary complex activates mTOR: Drug development in autophagy



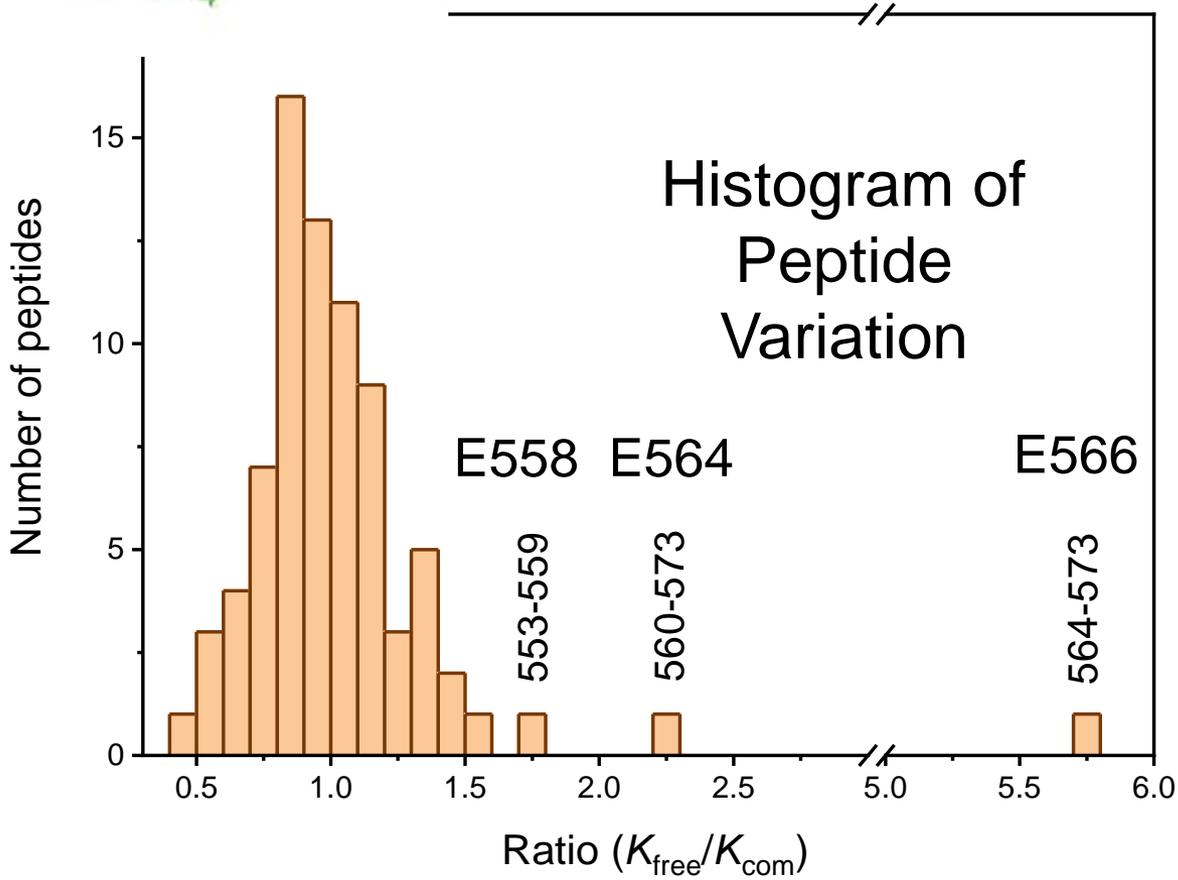
# Mapping the GABARAP ternary complex interaction sites



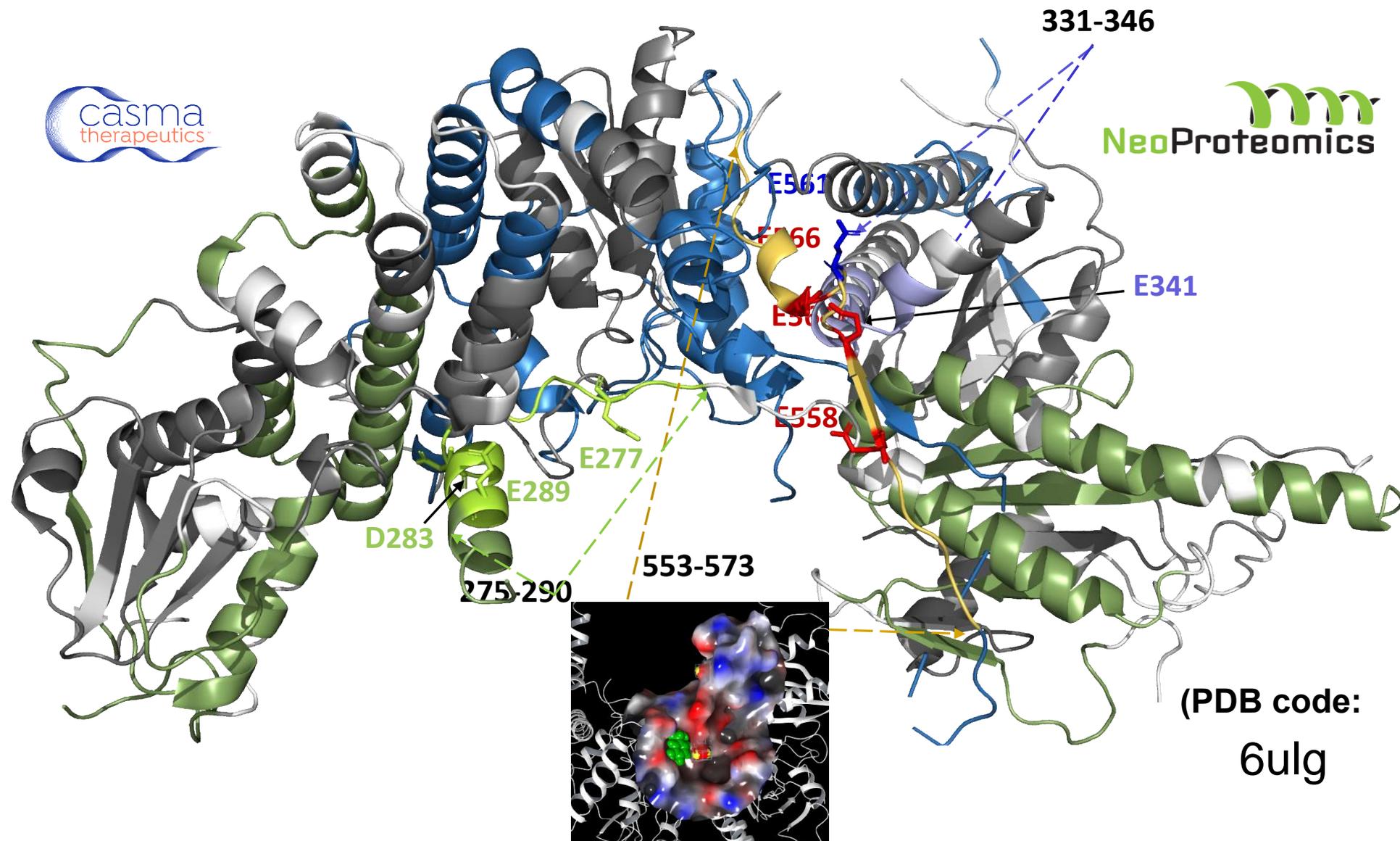
~200 peptides detected (87% coverage)

~90 with reliable labeling

ProtMap-MS software automation

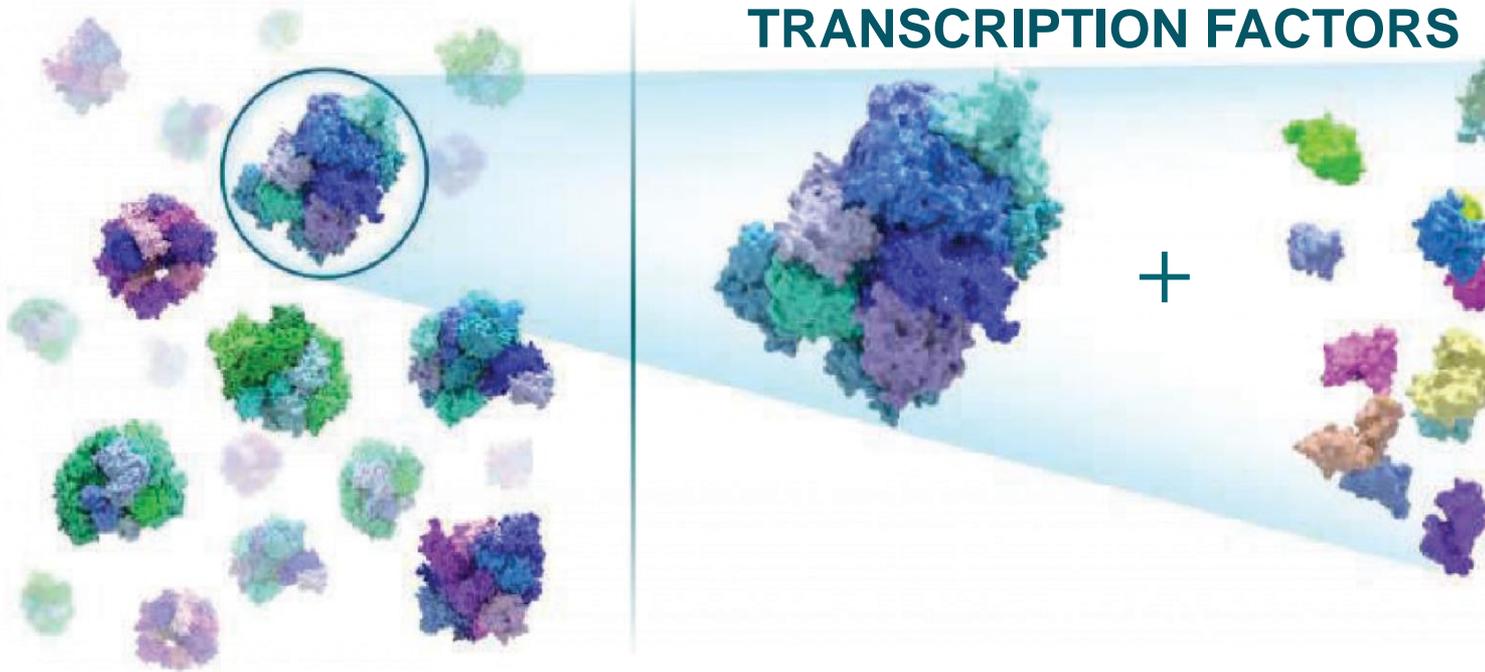


# Modeling GABARAP protections on cryo-EM structure of FLCN-FNIP binary complex.



**Drugging Undruggable  
Mega-Complexes**

**BAF COMPLEX AND ASSOCIATED  
TRANSCRIPTION FACTORS**



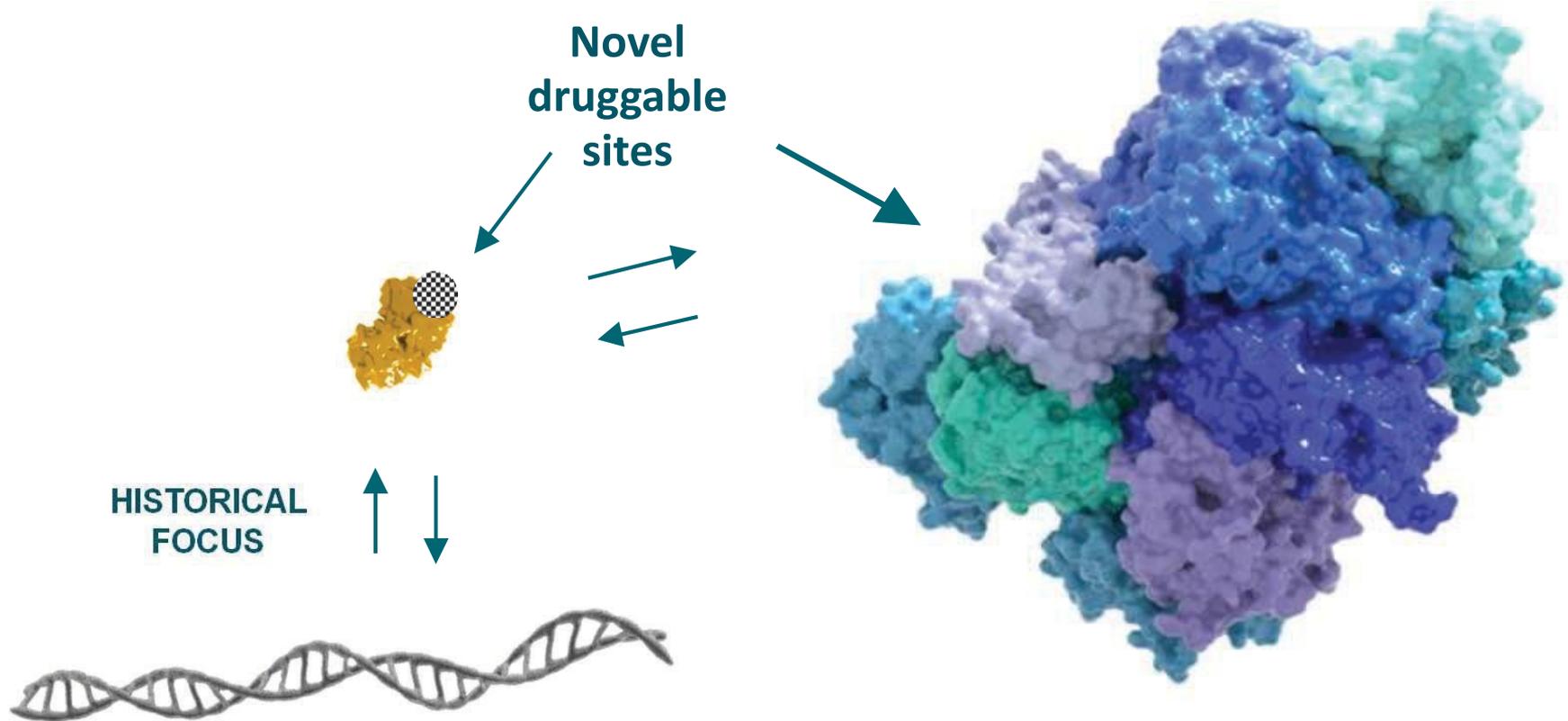
28 Chromatin  
Remodeling  
complexes and  
>1,000 TFs

BAF Complex  
Subunits Mutated  
and Dysregulated in  
Cancer

Estimate >100  
Transcription  
Factors Associated  
with BAF Complex

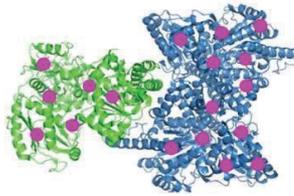
# TF interactions with chromatin remodelers

TFs are implicated in a range of cancers and other diseases.  
Historically difficult to target-featureless and tightly bound to DNA

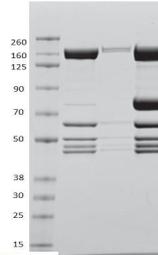


Novel approaches to drug transcription factors:  
including small molecules or degraders

# Mass Spec Footprinting: Survey BAF and TF surfaces



+

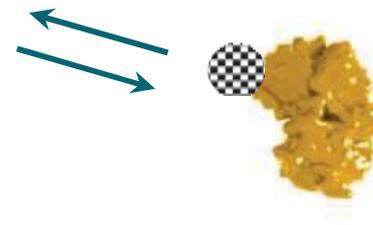
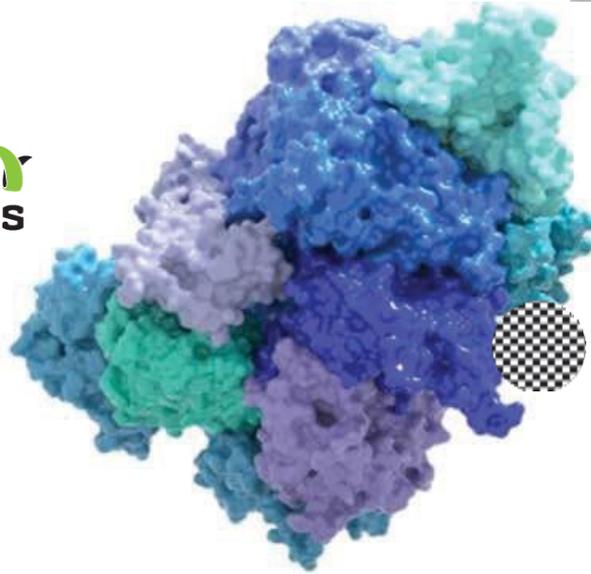


Pull down with BAF-subcomplexes

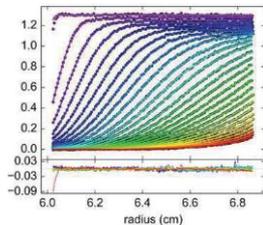
  
NeoProteomics

**FOGHORN**<sup>®</sup>  
THERAPEUTICS

- >100 TFs linked to BAF
- Foghorn-Neo Proteomics pursuing multiple TFs



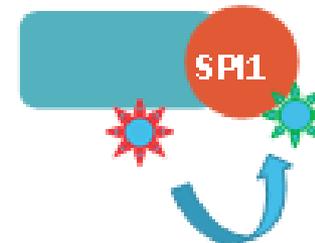
Biophysical, Structural and Biochemical confirmation of target interactions



AUC/SPR/ITC

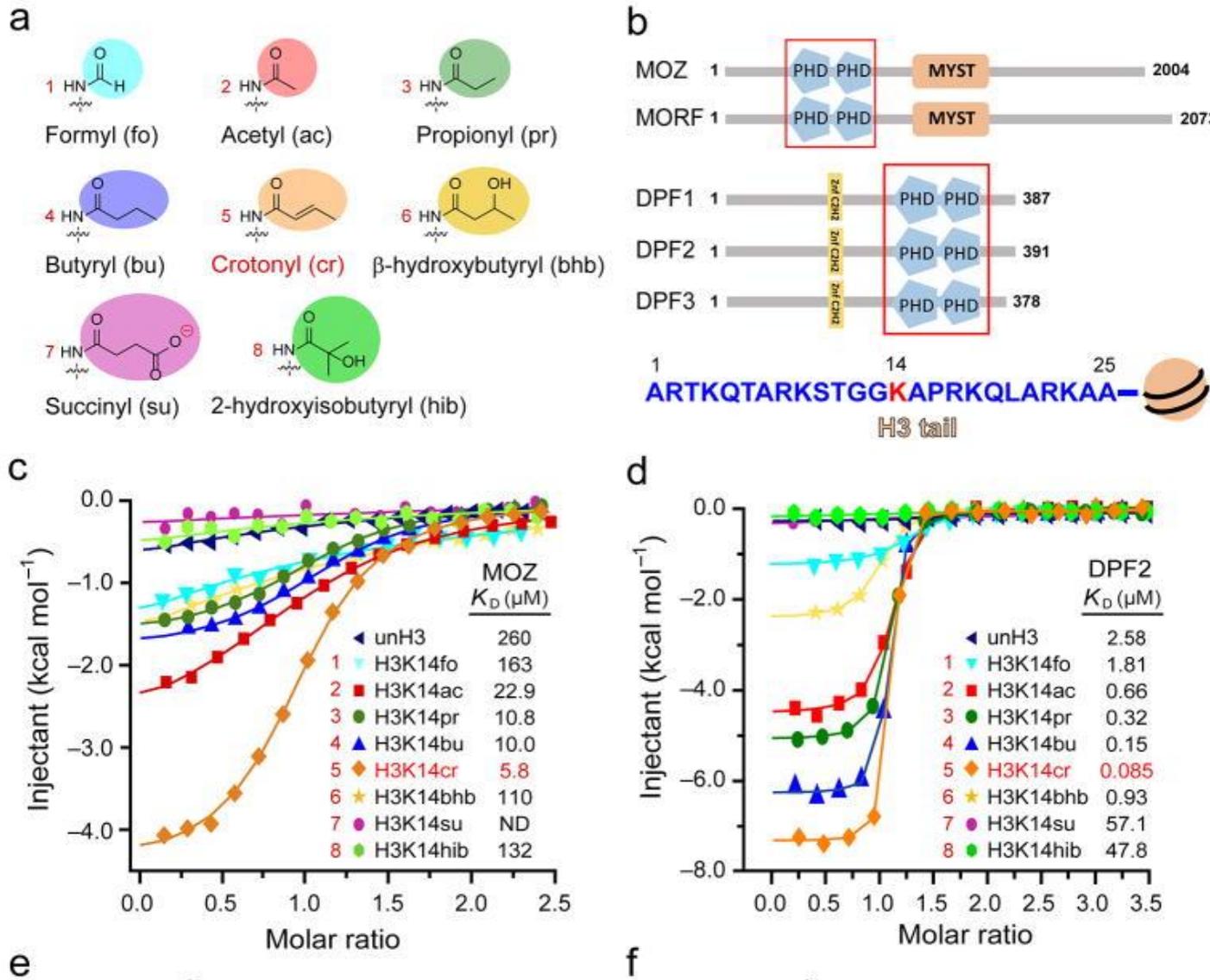


XTAL/NMR

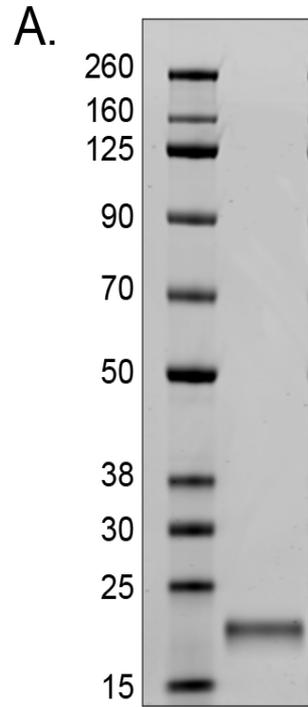


TF-FRET

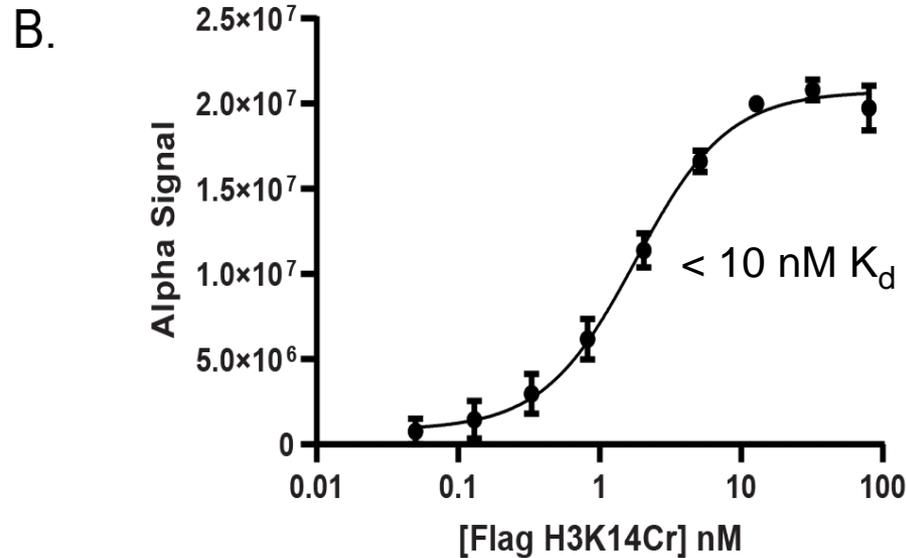
# Crotonylation of histones is read specifically by Double PHD (DPF) fingers of MOZ and DPF2



# DPF domain of BAF45D binds to Histone H3(1-25) K14Cr



Purified  
BAF45D  
vs MW  
markers



Binding  
isotherm of  
peptide +  
BAF45D

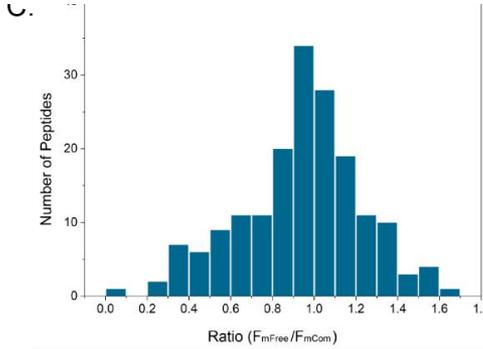
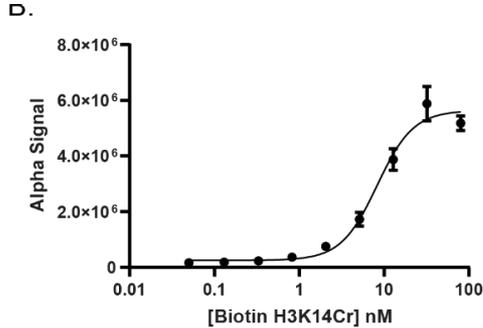
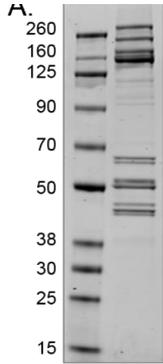
C.

Location	Sequence BAF45D	Protection Ratio
242-253	EDSQPPTPVSQR	1.01
261-281	KGPDGLALPNNYCDFCLGDSK	1.19
<b>286-300</b>	<b>TGQPEELVSCSDCGR</b>	<b>1.40</b>
301-318	SGHPSC <u>L</u> QFTPVMMAAVK	NL
<b>322-328</b>	<b>WQCIECK</b>	<b>3.41</b>
<b>329-350</b>	<b>CCNICGTSEND<u>D</u>QLLFCDD<u>C</u>DR</b>	<b>4.34</b>
351-379	<u>GYHMYCLTPSMSEP</u> PEGSW SCHLCLDLLK	1.05

PHD1

PHD2

# Full BAF Binds H3K14Cr at BAF45D PHD 1 and PHD 2 sequences



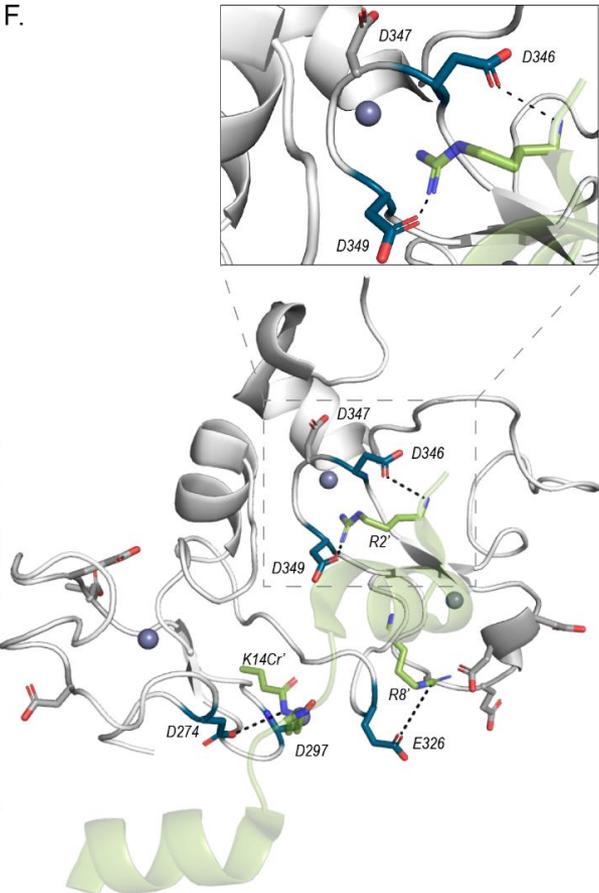
Histogram shows labeling of 177 peptides on BAF and complex out of 630 total detected (63% coverage)

Residues shown in contract with crotonly peptide modeled from MOZ domain homolog

Reveals Novel contacts with BRG/BRM

D.

Location	Sequence BAF45D within BAF	Protection Ratio
93-107	LSFPSIKPDTDQTLTK	0.44
109-124	EGLISQDGSSLEALLR	0.82
138-152	VDDDSLGEFPVTNSR	0.92
156-178	RILEPDDFLDDLDDEYEEDTPK	0.98
197-205	LDASILEDR	1.13
206-216	DKPYACDICGK	1.05
220-241	NRPGLSYHYAHSHLAEIEGEDK	0.64
242-253	EDSQPTPVSR	0.99
261-281	KGPDGLALPNNYCDFLGDSK	1.23
<b>286-300</b>	<b>TGQPEELVSCSDCGR</b>	<b>1.33</b>
301-318	SGHPSCLQFTPVMMAAVK	NL
<b>322-328</b>	<b>WQCIECK</b>	<b>1.24</b>
<b>329-350</b>	<b>CCNICGTSENDQLLFCDDCDR</b>	<b>1.50</b>
351-379	GYHMYCLTPSMSEPPGWSCH LCLDLLK	0.89
410-423	DHDGDYKDHIDYK	0.57



PHDs

E.

Location	Residue level Modification	Protection Ratio	
		BAF45D within BAF	BAF45D
261-281	D264	1.02	0.86
	D274	ND	2.37
	D279	1.07	0.93
286-300	D290	0.87	ND
	D291	1.15	1.12
	D297	1.29	1.40
322-328	D326	1.24	3.41
329-350	D337	ND	1.28
	D339	ND	1.34
	D340	ND	1.28
	D346	1.76	5.22
	D347	0.90	1.19
	D349	1.41	6.51

# Conclusions: Hack Your Structural Biology Problems with SMS

Footprinting and Structural Mass Spectrometry Provides Actionable High-Resolution Information optimized for:

- Antibody-Antigen interactions
- Protein Degradation and Molecular Glues (modest binding affinity)
- Protein-small molecule interactions
- Mega Complexes in Gene Trafficking and Metabolism

Dr. Hope Barkoukis,  
Chair of Nutrition

## Center for Proteomics and Bioinformatics

Mark Chance, PhD, Director  
Janna Kiselar PhD., Associate Director

Proteomics and Small Molecule  
Mass Spectrometry Core  
Danie Schlatzer, Director

Systems Biology Graduate  
Program  
Cheryl Cameron, PhD, Director

Center for Synchrotron  
Biosciences  
Mark Chance, Director,  
Erik Farquhar, Asst. Director

