

Oligonucleotide Mapping via LC-UV-MS/MS to Enable Comprehensive Primary Structure Characterization of an mRNA Vaccine Against SARS-CoV-2

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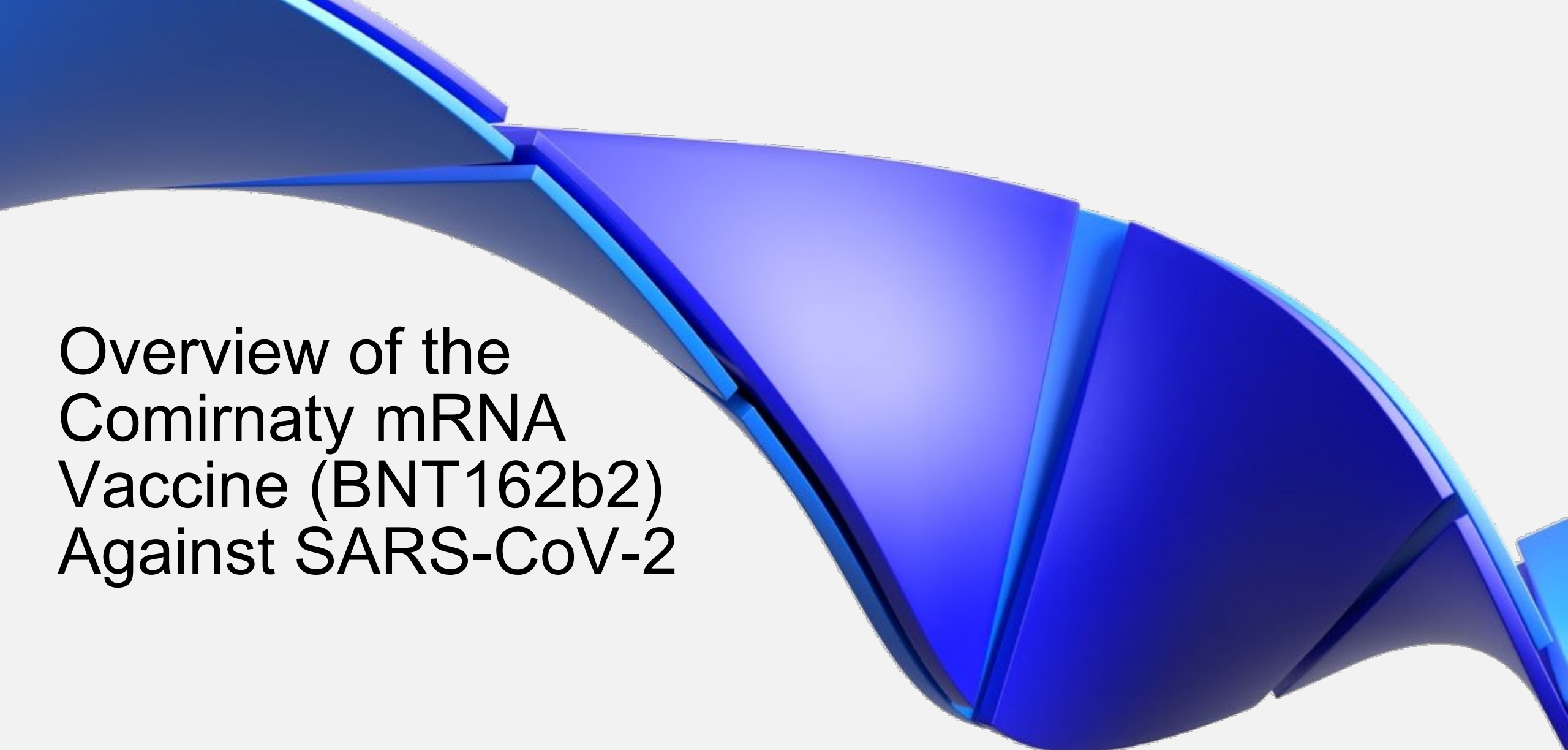


Breakthroughs that change patients' lives



Outline

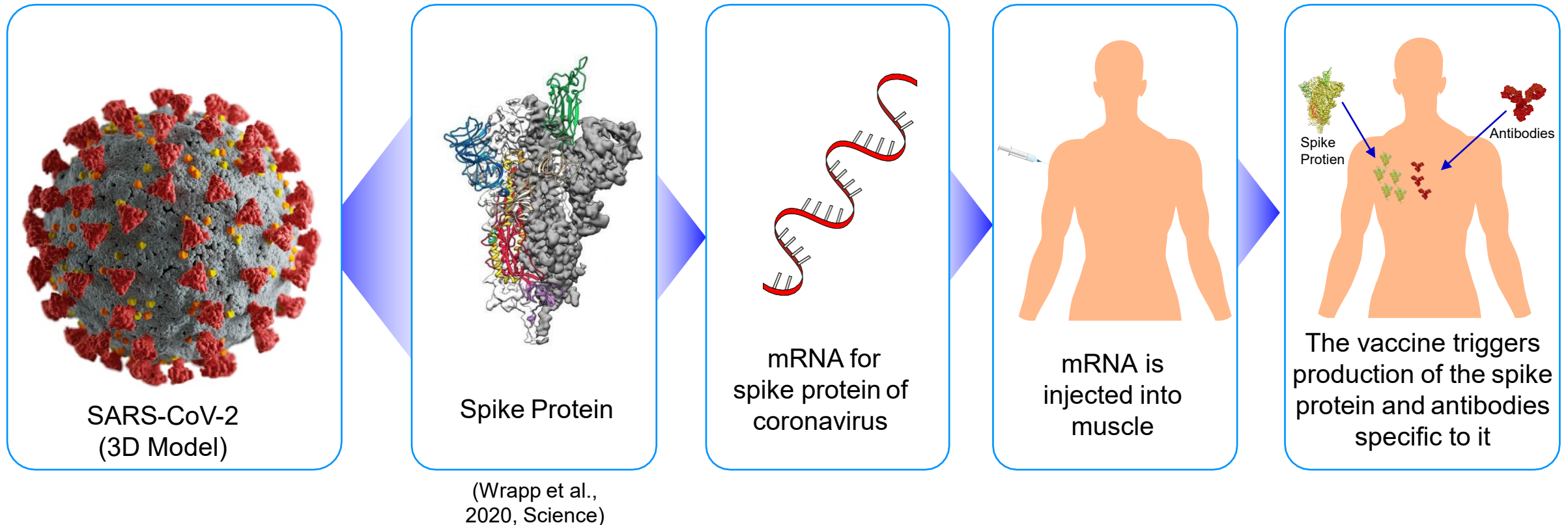
- Overview of the Comirnaty (BNT162b2) mRNA Vaccine Against SARS-CoV-2
- Oligonucleotide Mapping of BNT162b2 mRNA Primary Structure by LC-UV-MS/MS
 - Workflow
 - Core reportables providing direct primary structure understanding
- Oligonucleotide Mapping Method Development Highlights
- Additional Oligonucleotide Mapping Application Highlights
 - Batch Comparability
 - Variant Construct Comparisons



Overview of the Comirnaty mRNA Vaccine (BNT162b2) Against SARS-CoV-2

Basic Design of Pfizer/BioNTech mRNA Vaccine(s) against SARS-CoV-2

- Train patient's immune system to recognize the virus, specifically the spike protein on the surface
- Give the "code" or "recipe" of the spike protein to your cells



Analytical Characterization of the Drug Substance (mRNA) is Critical for Development of a High Quality Manufacturing Process & Product

Drug Substance (mRNA)

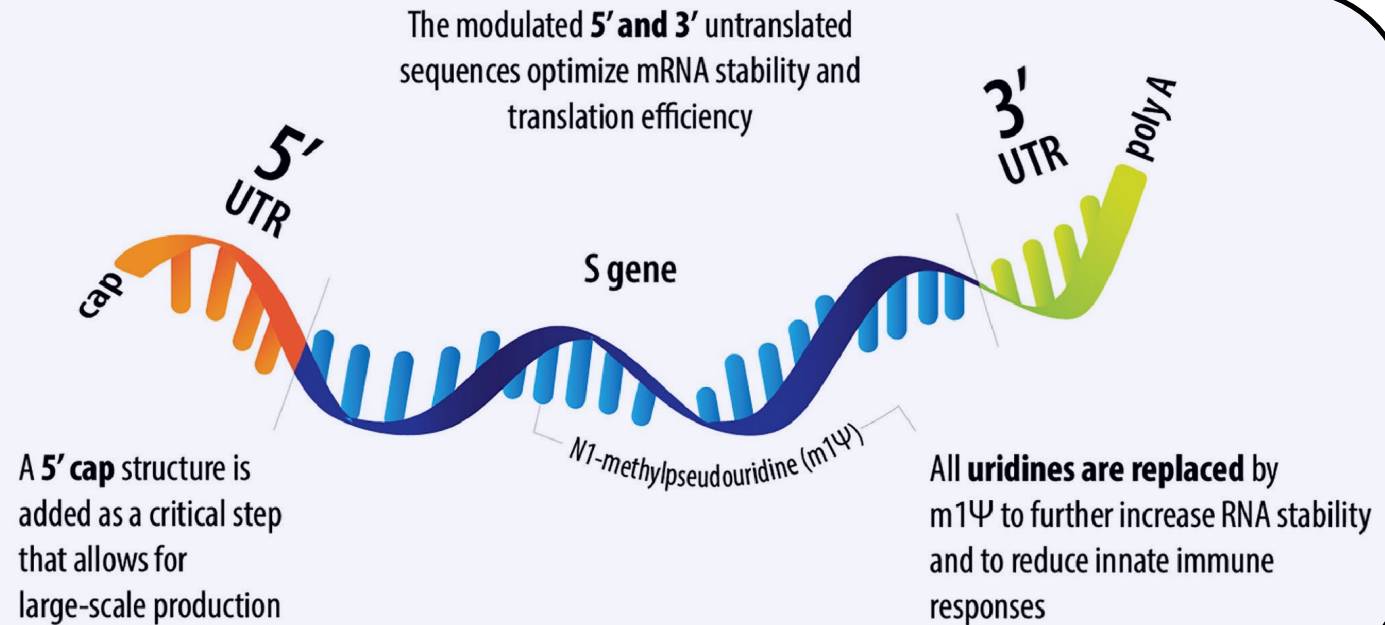


Figure adapted from Lewis LM, Badkar AV, Cirelli D, Combs R, Lerch TF. J Pharm Sci. 2023 Mar;112(3):640-647.

Platform QC Assays

- Compendial methods
- **Purity** by Capillary Gel Electrophoresis
- **Purity** by Immunoblot
- **Concentration** by UV spectroscopy
- **Identity, Impurities** by PCR-based methods

Heightened Characterization

Primary Structure

- **Oligonucleotide mapping (LC-UV-MS/MS)**
- Nucleoside Analysis (LC-UV-MS)
- NextGen Sequencing (NGS)

Higher Order Structure

- Circular Dichroism (CD)

Protein Expression

- FACS
- Western Blot

Oligonucleotide Mapping of mRNA Primary Structure by LC-UV-MS/MS is Applied in Three Ways to Support mRNA Vaccine Development

Direct Primary Structure Understanding

- 5' terminus (capping)
- 3' terminus (poly(A) – tail)
- Full-length mRNA

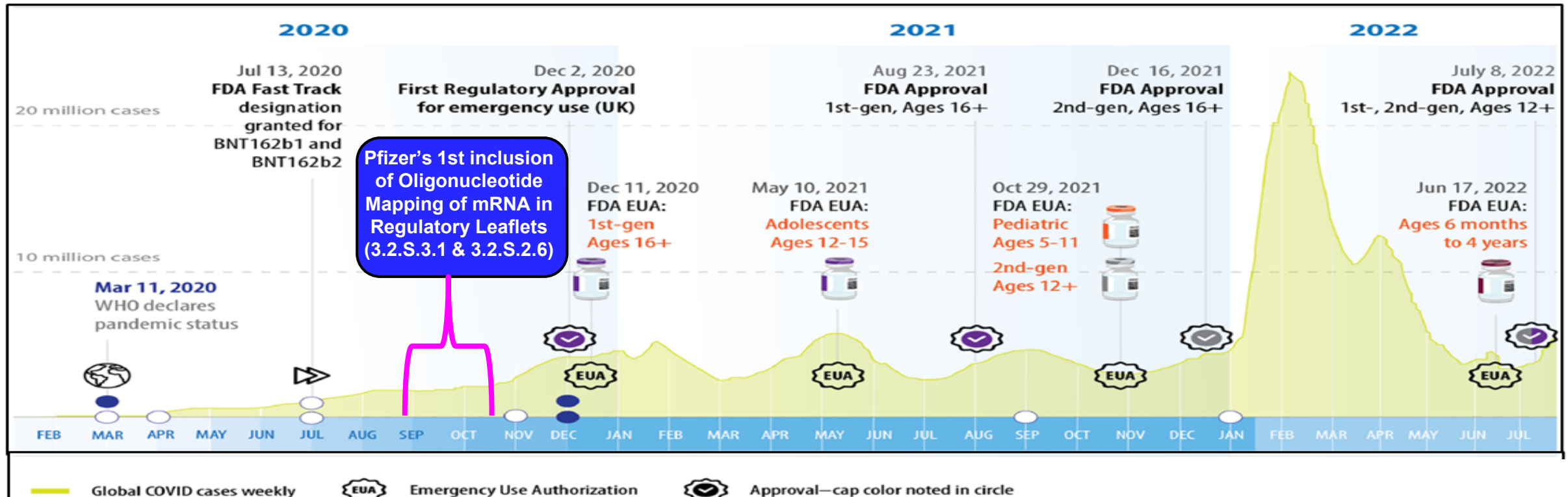
Batch Comparability Assessment

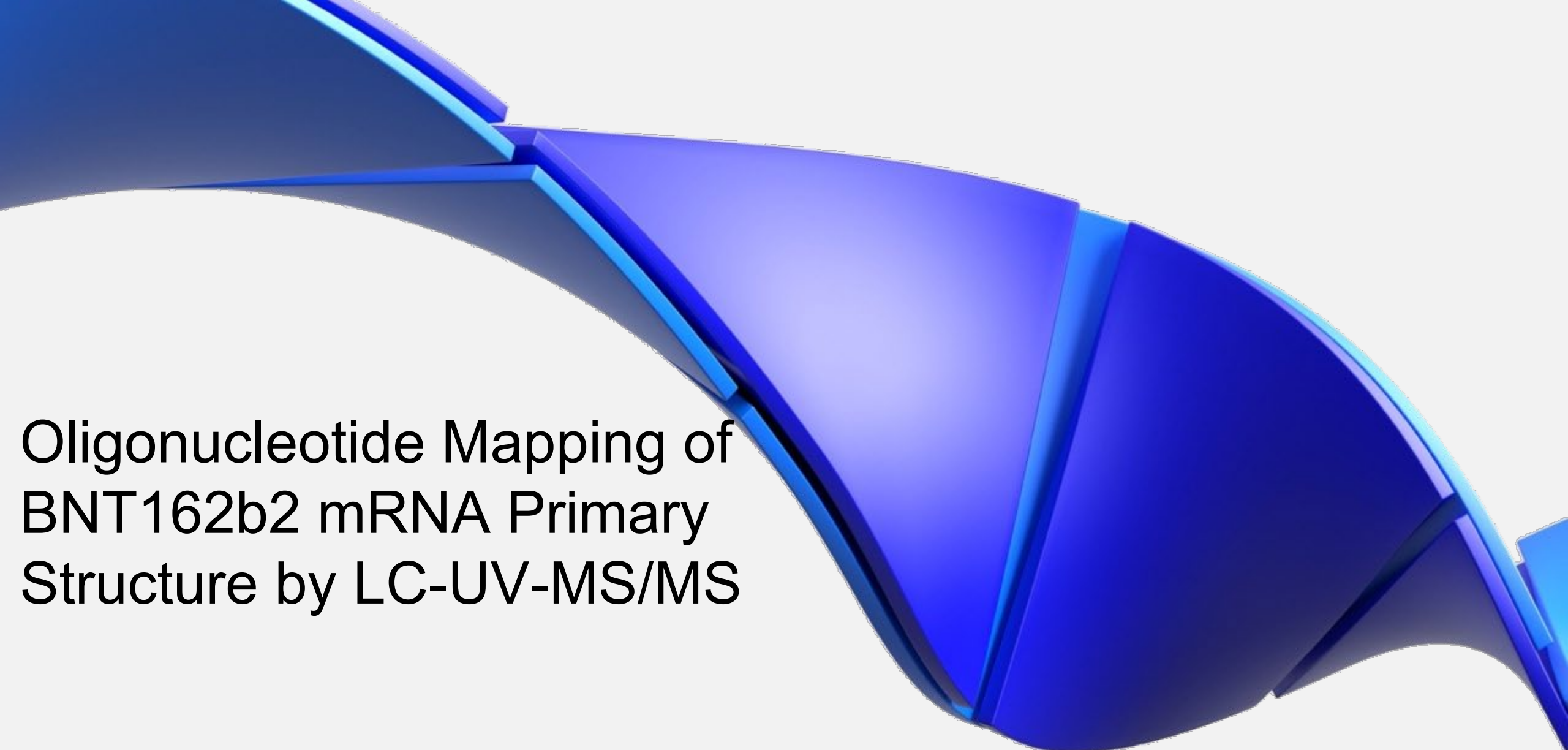
- Process changes
- Scale-up
- Scale-out

Orthogonal Identity

- BNT162b2 (Original)
- Variant constructs (Delta, Omicron)

Oligonucleotide Mapping of mRNA Primary Structure by LC-UV-MS/MS has Supported Regulatory Filings And Launches in 180+ Markets Globally

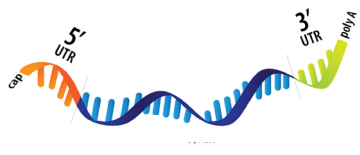




Oligonucleotide Mapping of
BNT162b2 mRNA Primary
Structure by LC-UV-MS/MS

Simple, Robust, Semi-Automated Workflow

Rapid One-Pot, One-Enzyme
Sample Preparation



Ribonuclease T1
Cleave 3' to G



+

IP-RP-HPLC-UV



MS-MS/MS



+

Semi-Automated Data Analysis

Commercial
Software



+

In-House
VBA Tools



Semi-Automated Data Analysis Workflow



1. Automated Search

- Mass table by retention time
- Identifications (~70% Coverage)



2. Automated LC-UV Annotation



- Match Peak IDs to Chromatogram
- Reformatted Mass Table



3. Supplement LC-UV Annotation



- Data mining & MS/MS Analysis Tools



4. Supplement Missing Coverage



- Data mining & MS/MS Analysis Tools



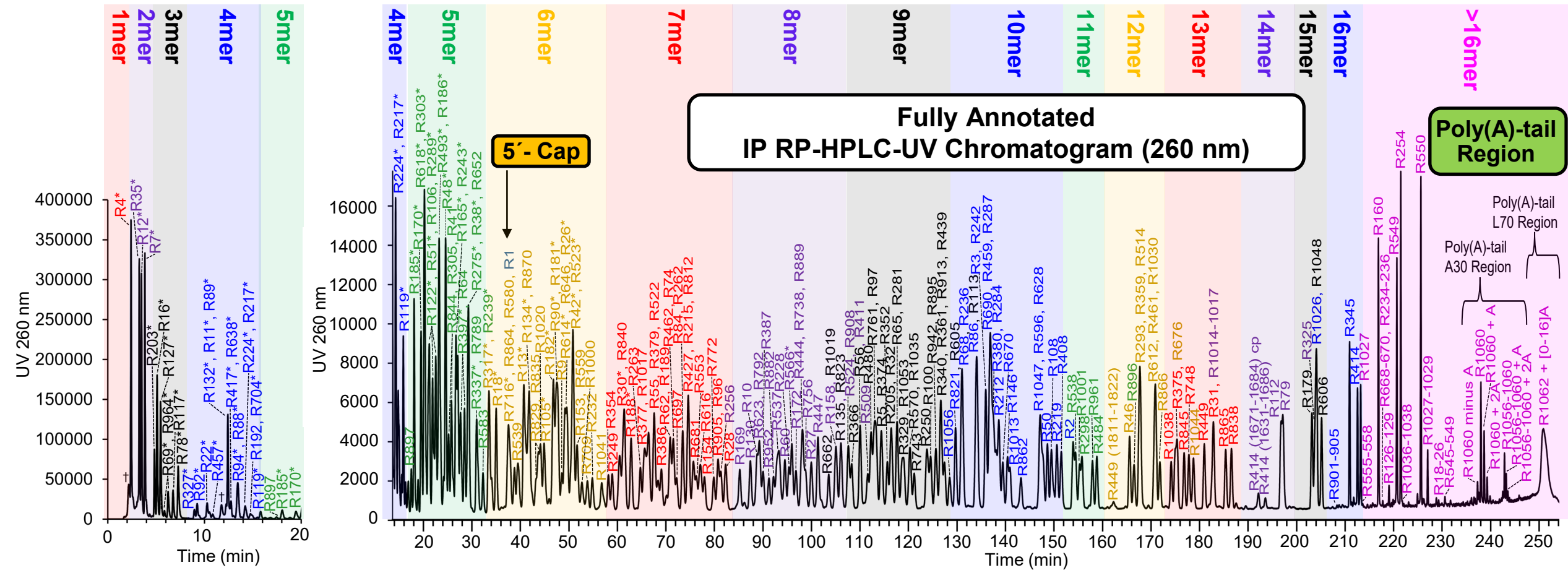
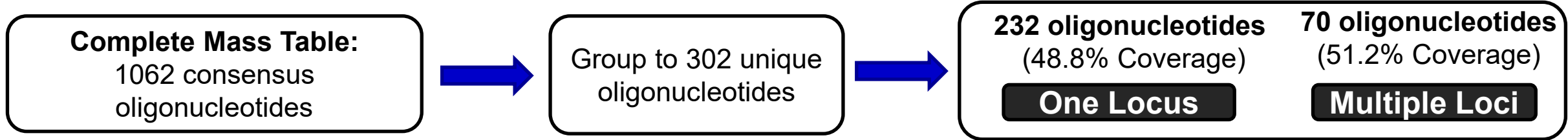
5. Add 5' & 3' Termini Characterization



Final Reportables

- Fully-Annotated Chromatographic Map
- Sequence Coverage Calculation & Map
- Curated Mass Table
- 5' & 3' terminus characterization

Result: Fully Annotated BNT162b2 Oligonucleotide Map



Result: 100% BNT162b2 Sequence Coverage Observed

Approximately half of consensus RNaseT1 cleavages map to one locus and half contain multiple loci

232 oligonucleotides
(48.8% Coverage)

One Locus

70 oligonucleotides
(51.2% Coverage)

Multiple Loci

46 oligonucleotides

14 oligonucleotides

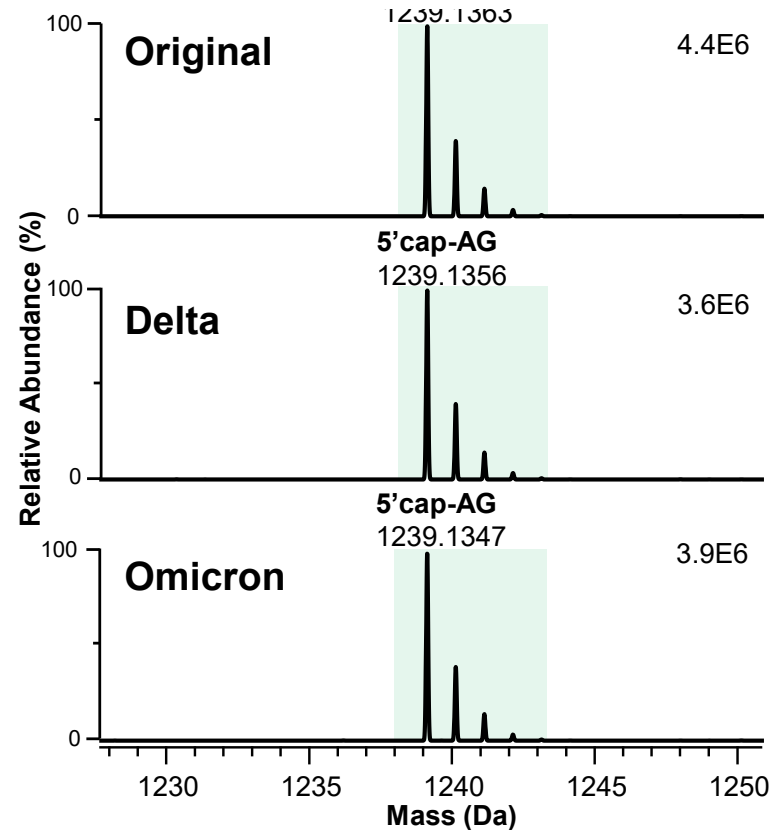
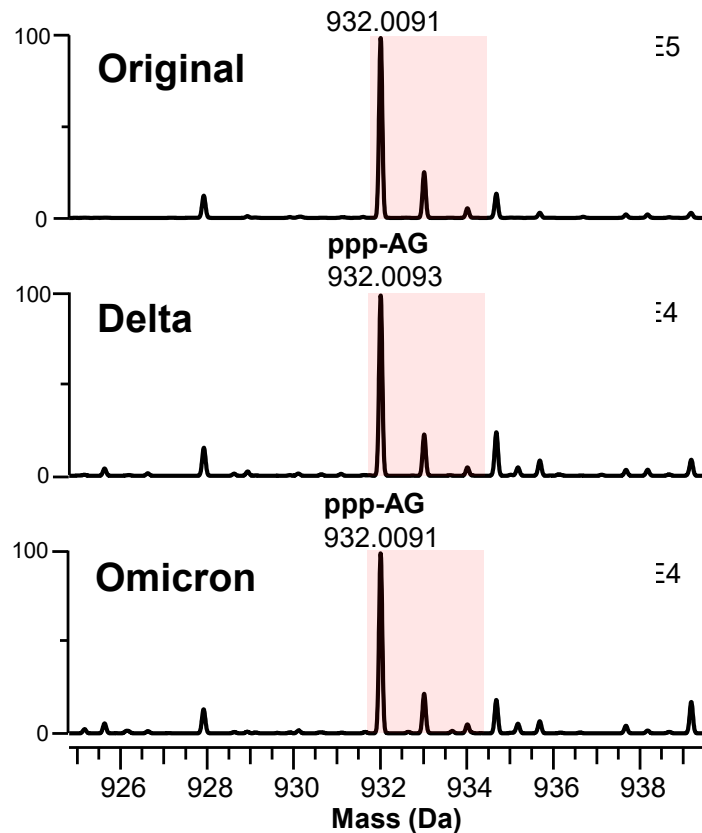
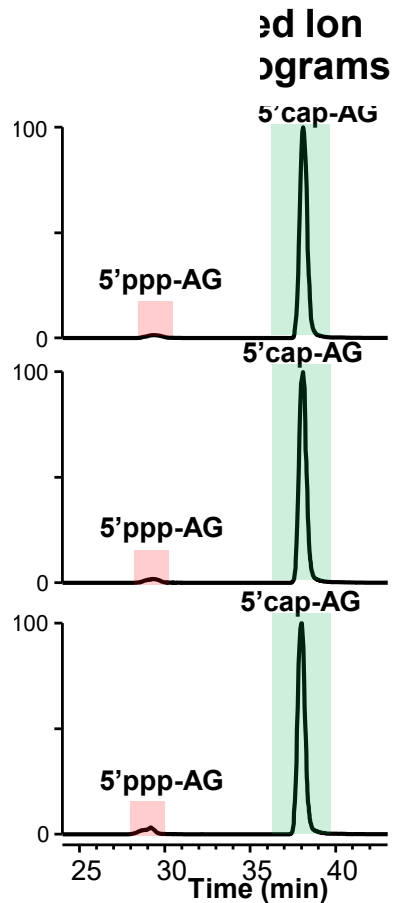
Missed-Cleavages & Non-Consensus Cleavages



Oligonucleotide Mapping Enables Simultaneous Characterization of the 5' Terminus Without Affinity Purification

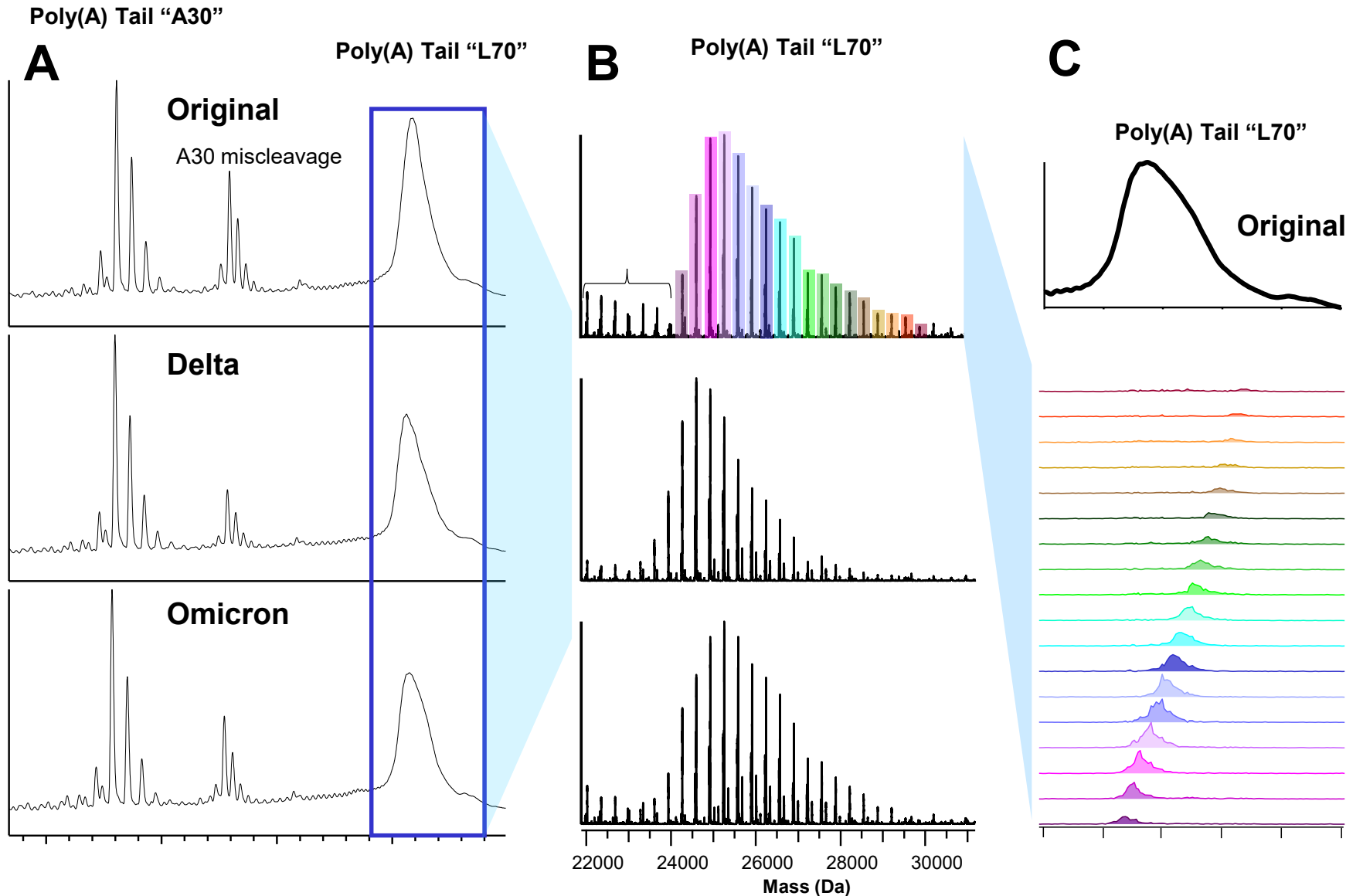


C Capped 5' terminus
Deconvolved Mass Spectra



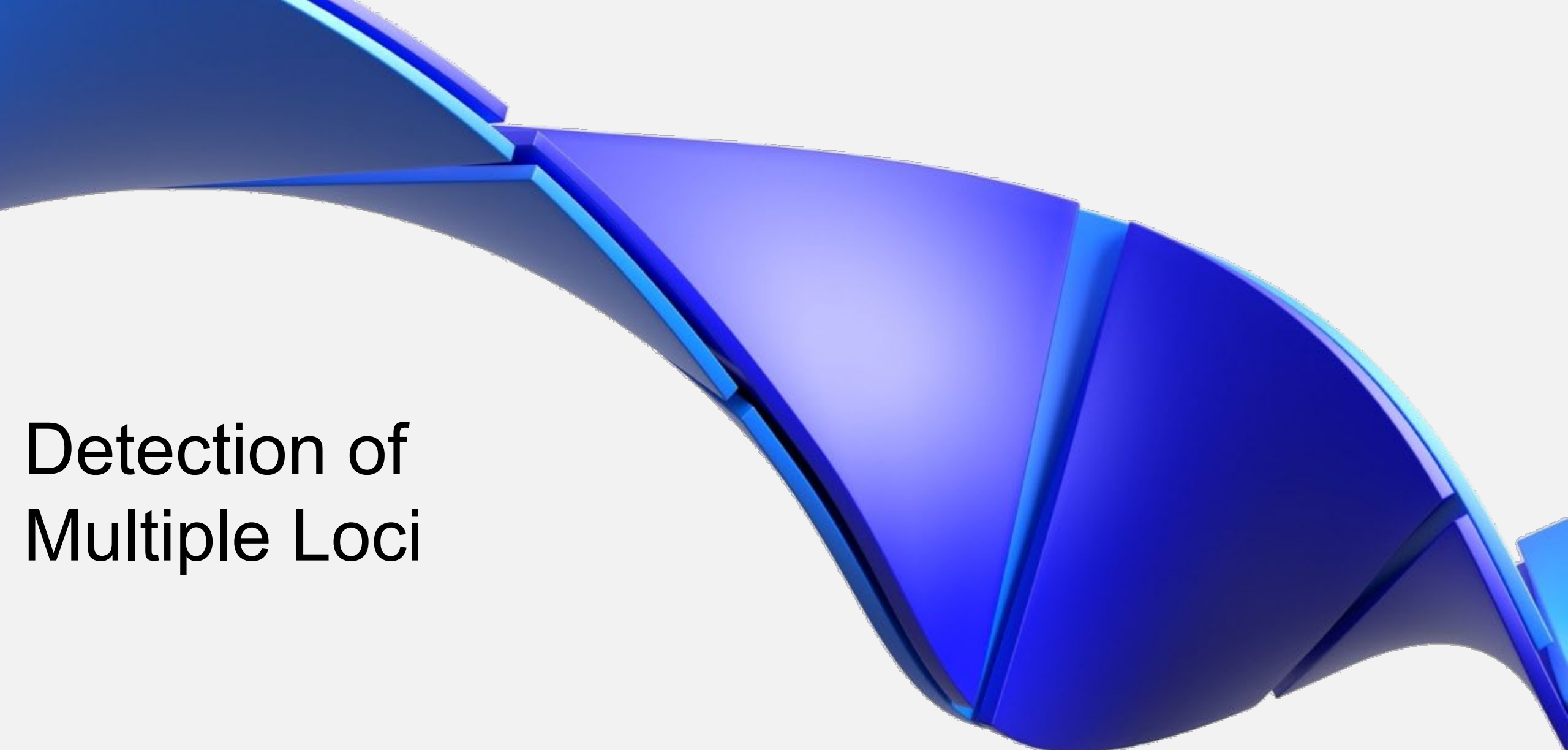
Majority of 5' terminus is capped

Oligonucleotide Mapping of mRNA Enables Simultaneous Characterization of the 3' Terminus Without Affinity Purification



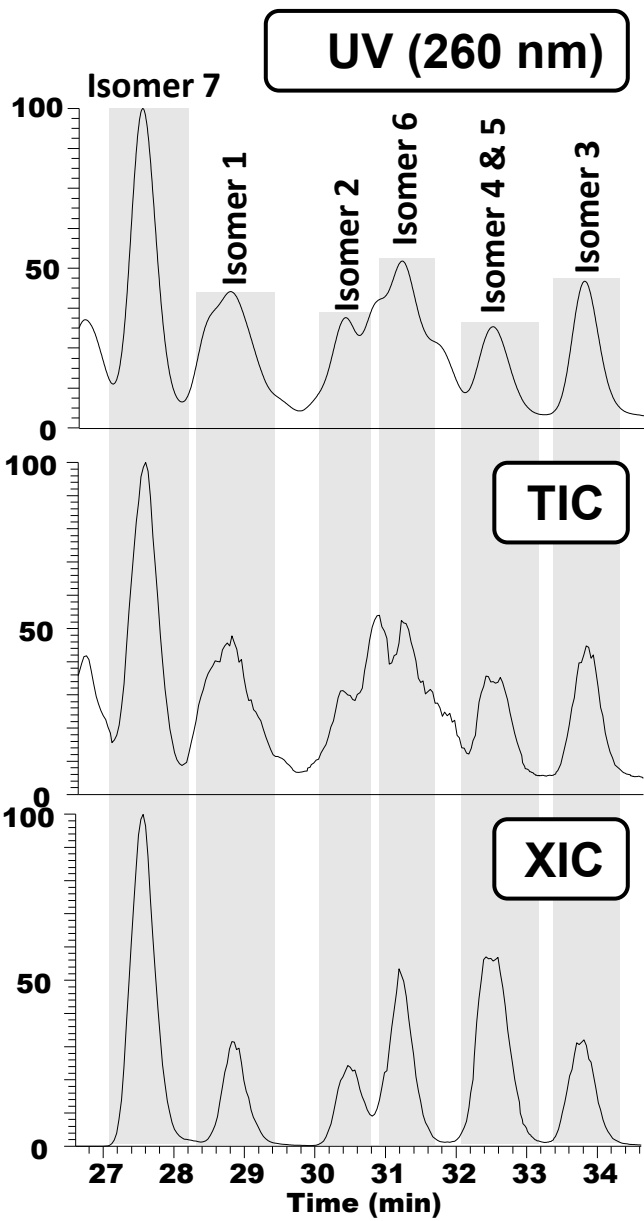
Poly(A) Tail Distribution Characterized

Moderate Shifts in Poly(A) Length Distribution Are Detectable



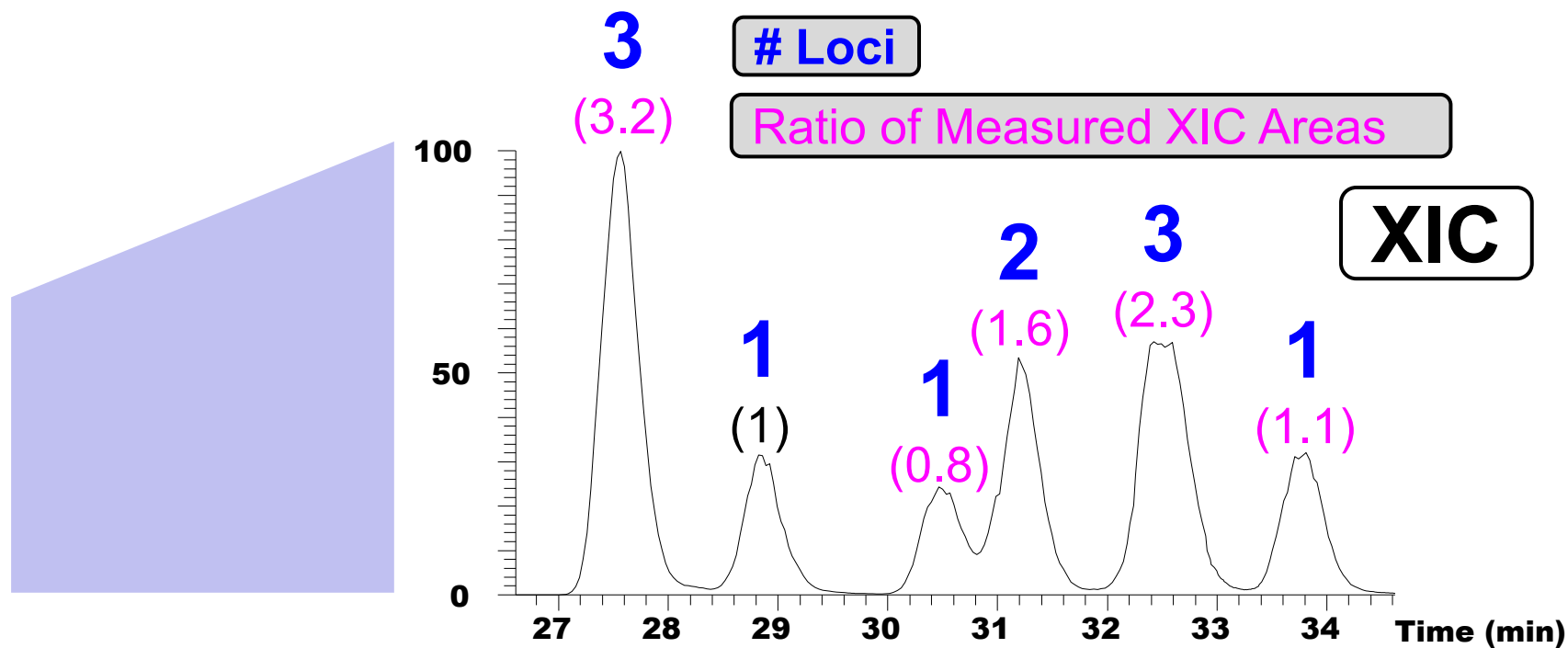
Detection of Multiple Loci

Measured XIC Areas of Non-Unique Sequence Isomers Correlate with their Number of Loci in the Full Length mRNA Sequence



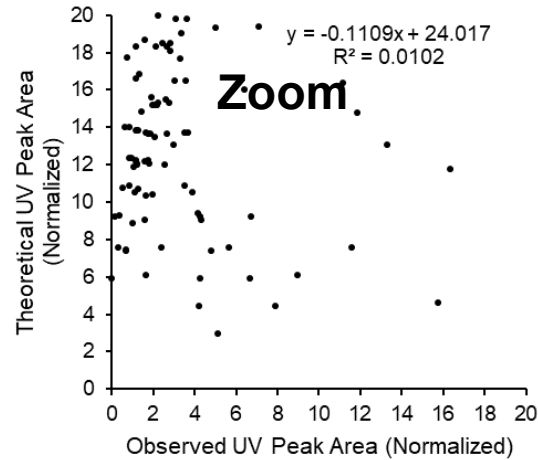
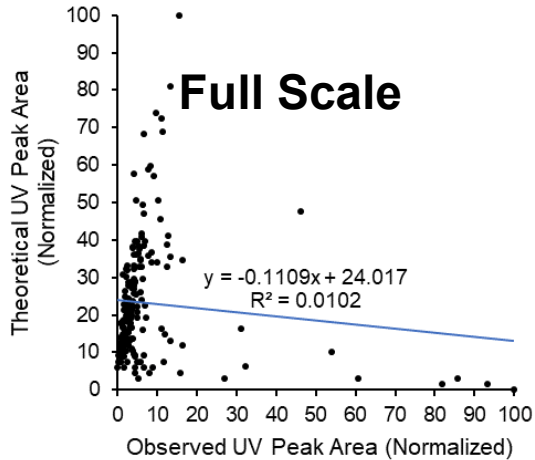
Theoretical				Observed	
Oligonucleotide	Sequence	# of Loci	Monoisotopic Mass (Da)	Retention Time (min)	Ratio of XIC Areas Normalized to Isomer 1
Isomer 1	CVAAG	1	1646.2453	28.9	1
Isomer 2	VCAAG	1		30.5	0.8
Isomer 3	AVCAG	1		33.8	1.1
Isomer 4	AVACG	1		32.5	2.3
Isomer 5	AACVG	2		31.2	1.6
Isomer 6	VACAG	2		31.2	1.6
Isomer 7	CAAVG	3		27.6	3.2

v = N1-methyl pseudouridine



Measured UV Areas Across Oligonucleotide Map Correlate with Theoretical UV Areas but only when Accounting for All Loci of Non-Unique Oligonucleotides

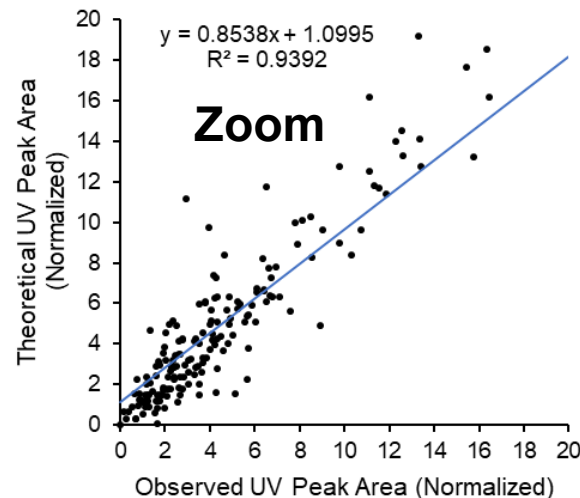
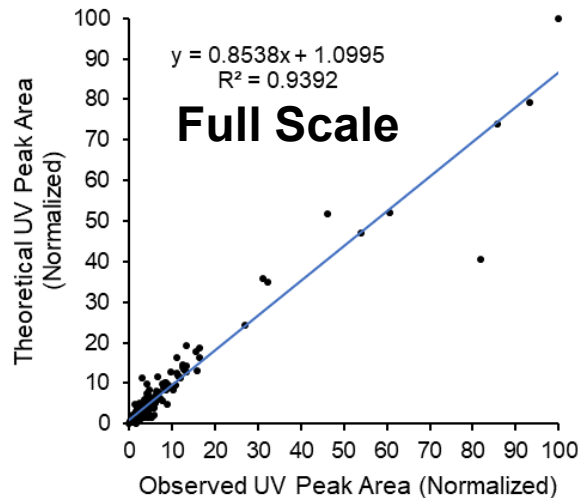
Not Accounting for Multiple Loci of Non-Unique Oligonucleotides



**Poor
Correlation**

Theoretical peak areas were calculated using oligonucleotide compositions and NMR-derived extinction coefficients for pdG*, pdA*, pdC*, and N1-methylpseudouridine monophosphate[‡]

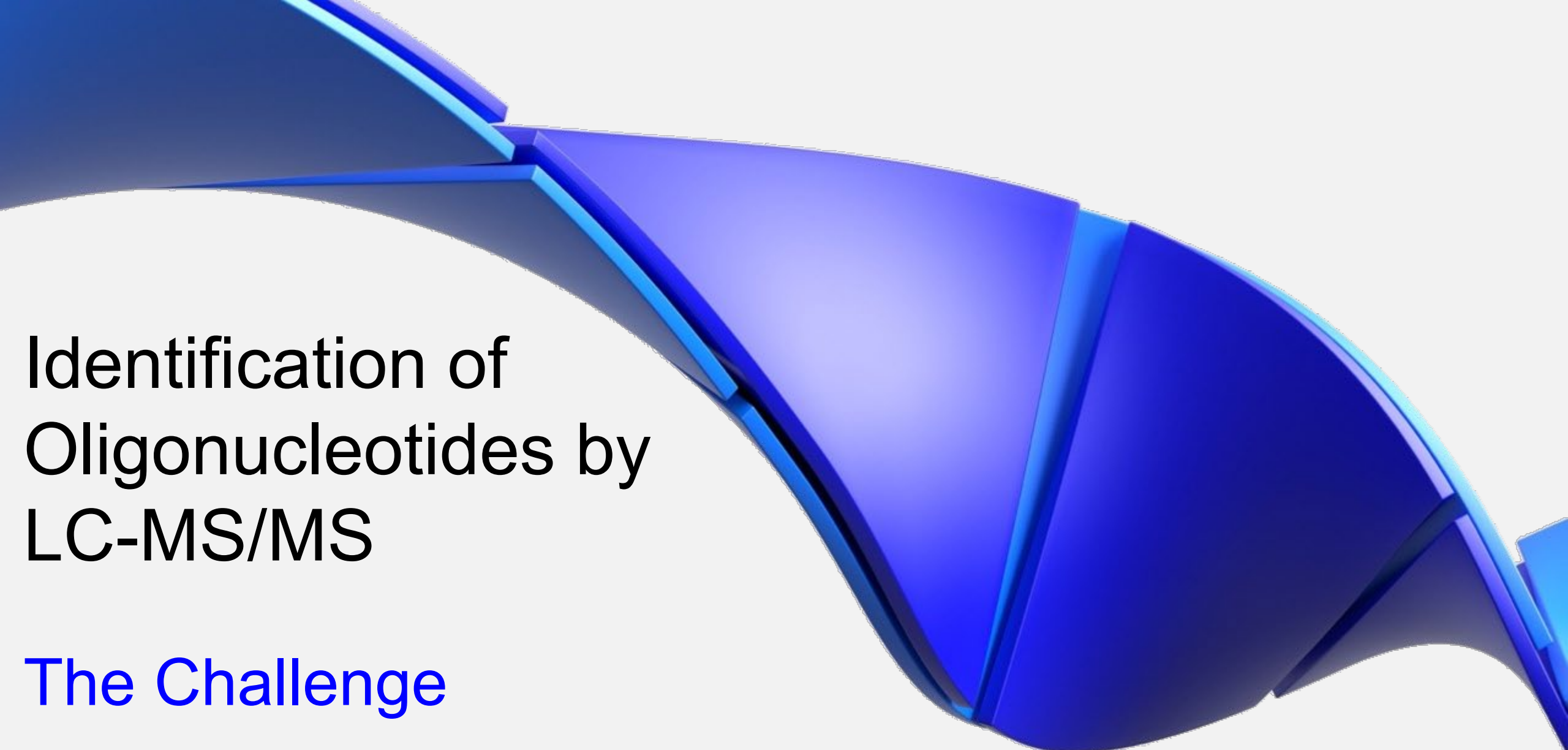
Accounting for Multiple Loci of Non-Unique Oligonucleotides



**Good
Correlation**

*Cavaluzzi MJ, Borer PN. Nucleic Acids Res. 2004 Jan 13;32(1):e13.

[‡]Empirically determined at Pfizer



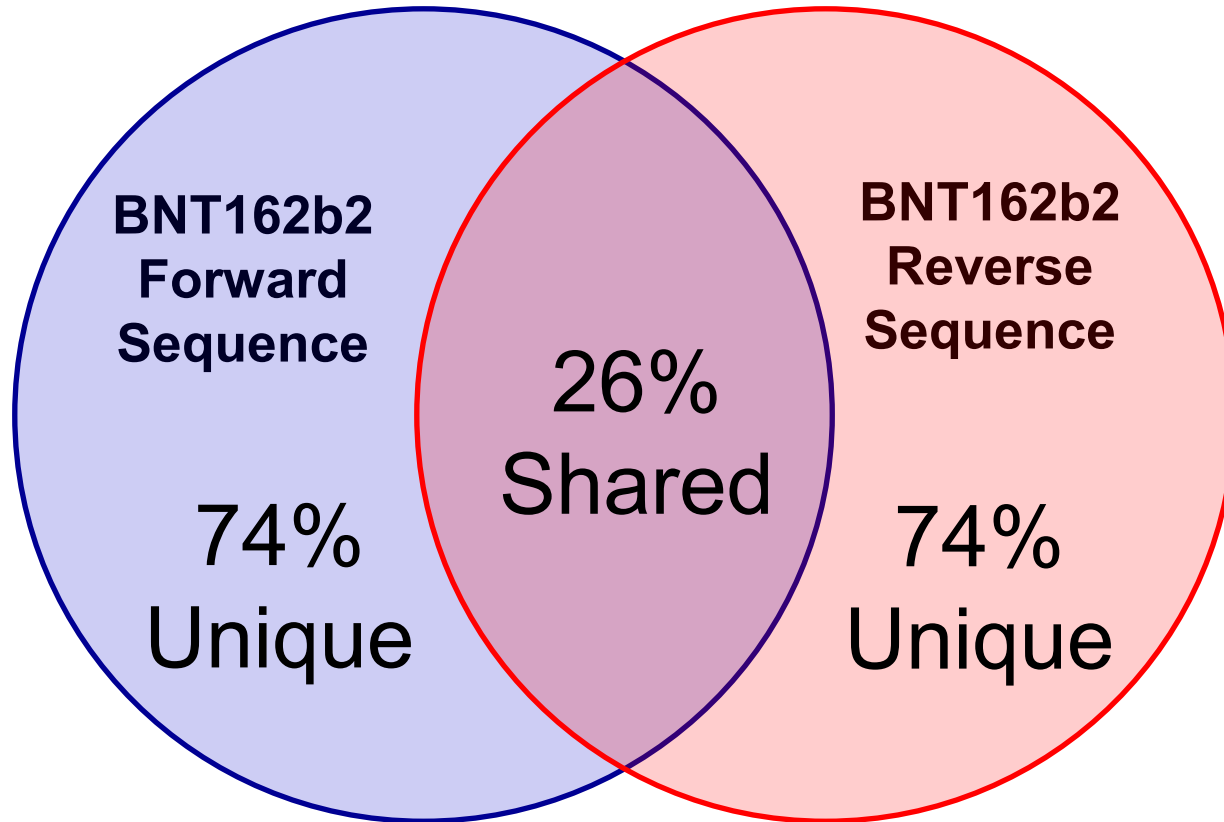
Identification of Oligonucleotides by LC-MS/MS

The Challenge

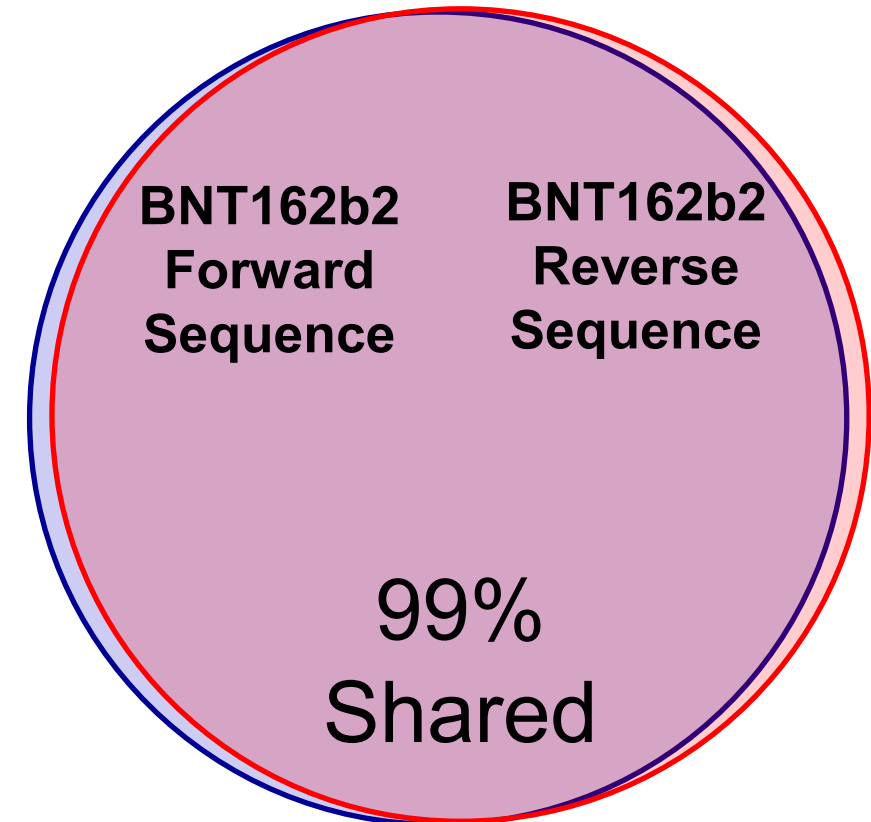
MS/MS Fragmentation is Critical For Differentiating Oligonucleotide Sequence

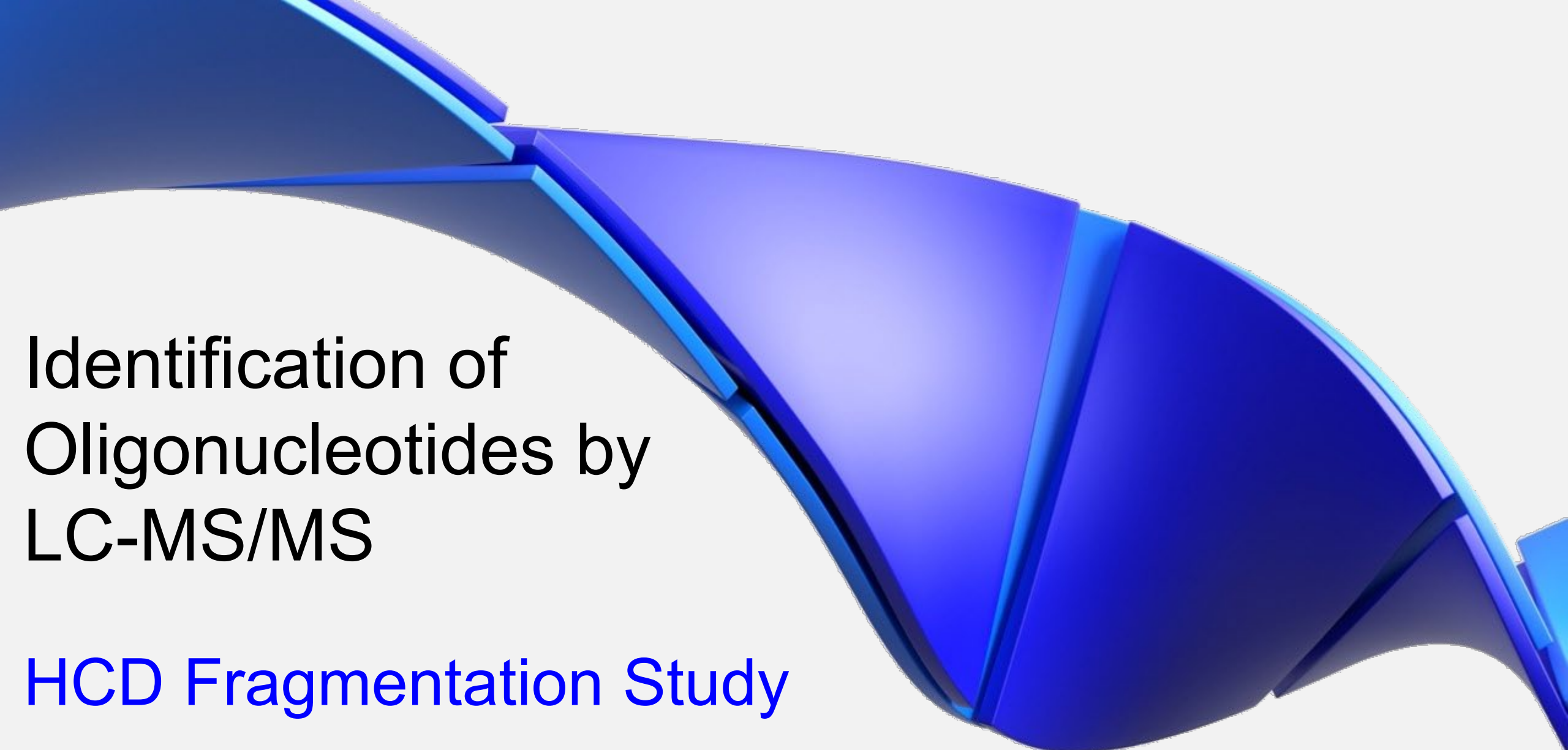
— Isomers Generated Through Enzymatic Digestion by RNaseT1

Most Oligonucleotide
Sequences are Unique



Most Oligonucleotide
Masses are Not Unique





Identification of Oligonucleotides by LC-MS/MS

HCD Fragmentation Study

Higher Energy Collision Dissociation (HCD) Parameters Optimized to Generate Fragmentation Appropriate for Oligonucleotide Mapping

Fragmentation of RNA Oligonucleotides is Complex

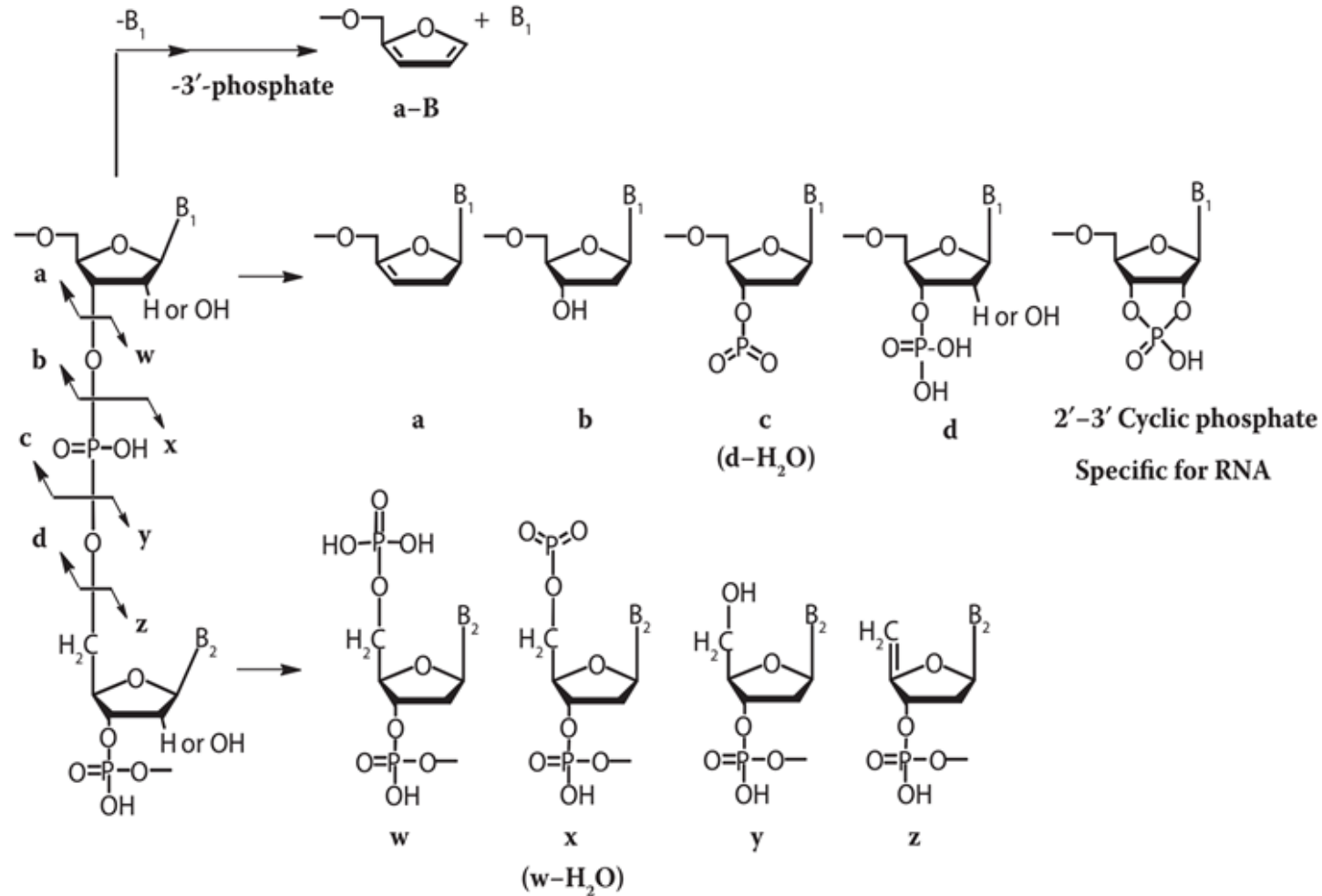
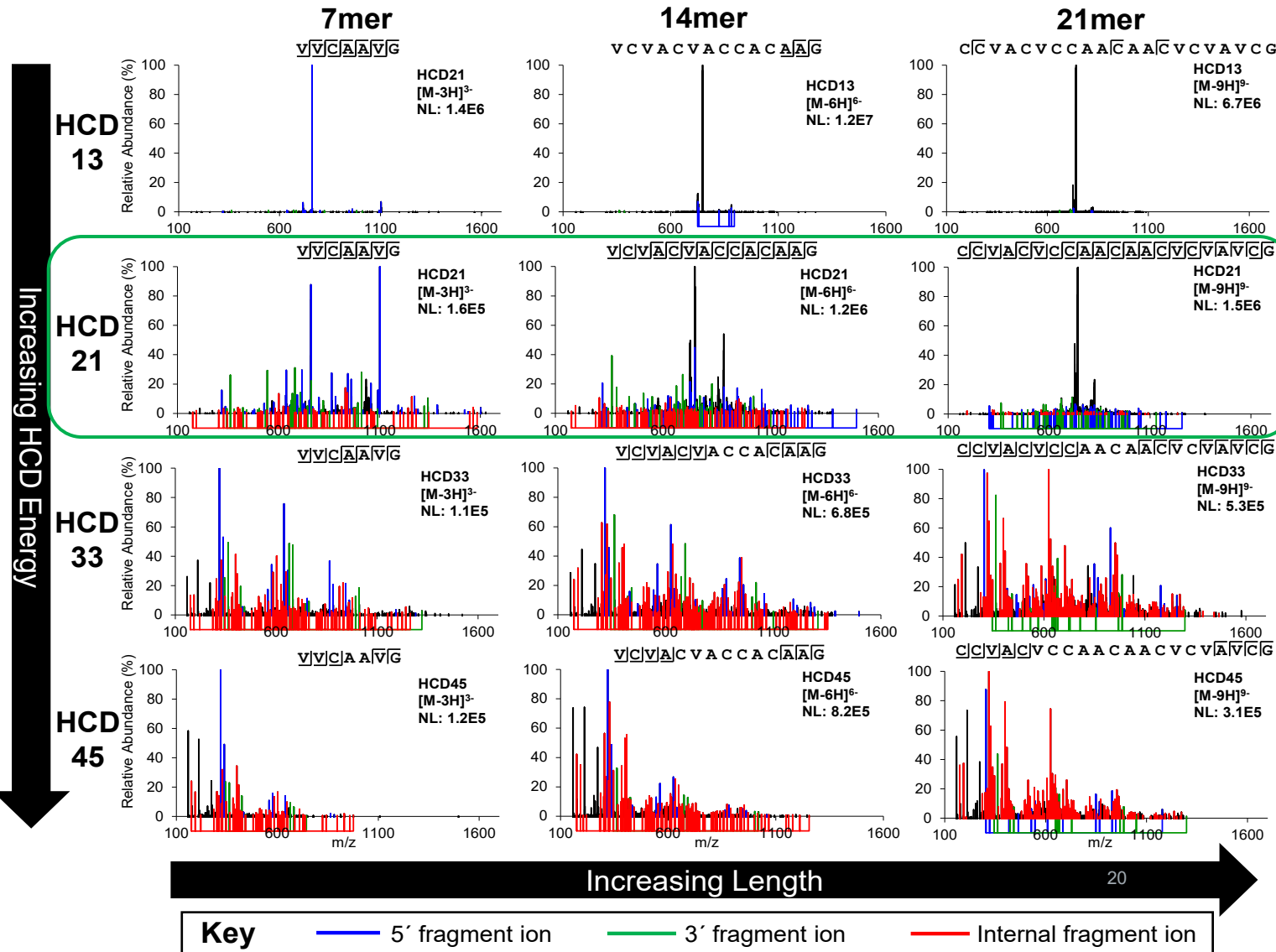


Figure adapted from McLuckey SA, Van Berkel GJ, Glish GL. Tandem mass spectrometry of small, multiply charged oligonucleotides. J Am Soc Mass Spectrom. 1992 Jan;3(1):60-70.

HCD Collision Energy Optimized at Stepped CE 17, 21, 25

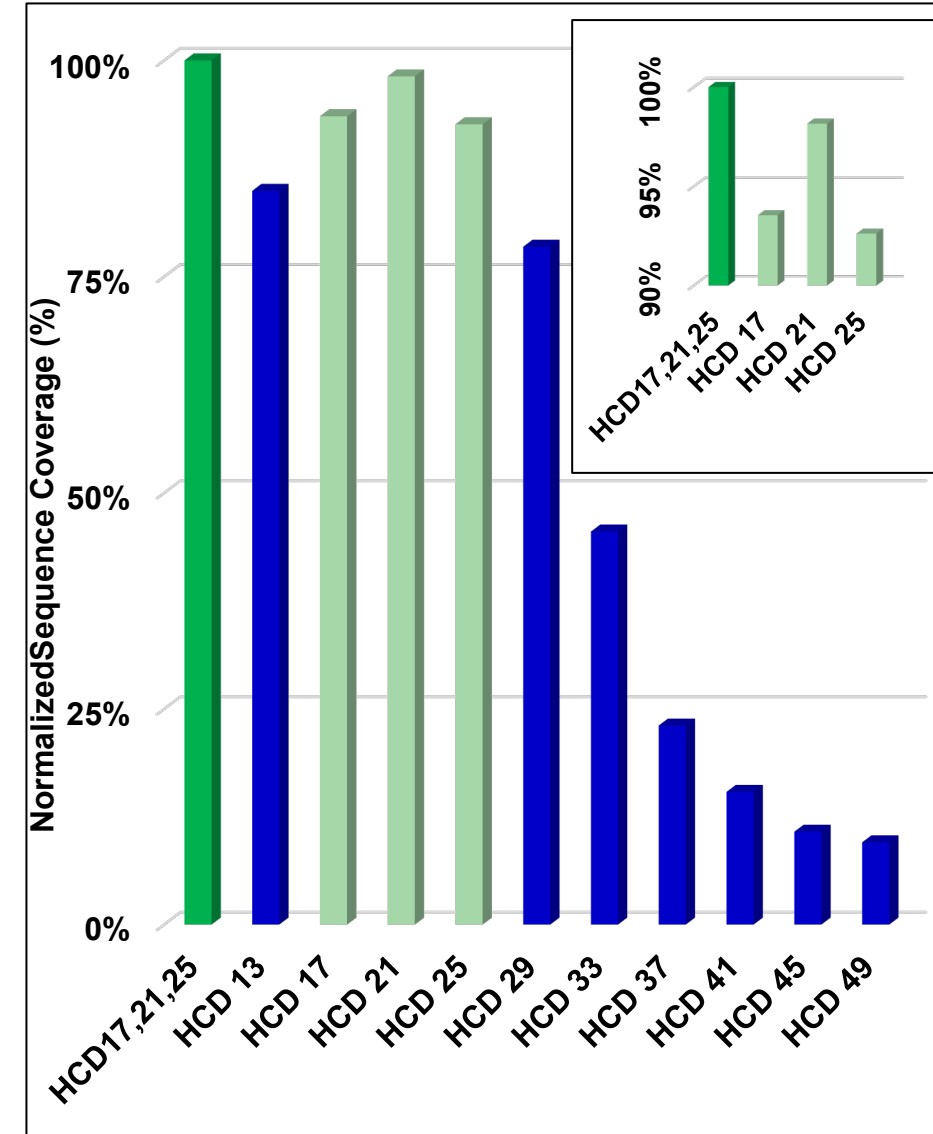
Oligonucleotide Fragment Ion Coverage as a Function of HCD Energy & Length

Normalized BNT162b2 Sequence Coverage Across All Oligonucleotides as a Function of HCD Energy



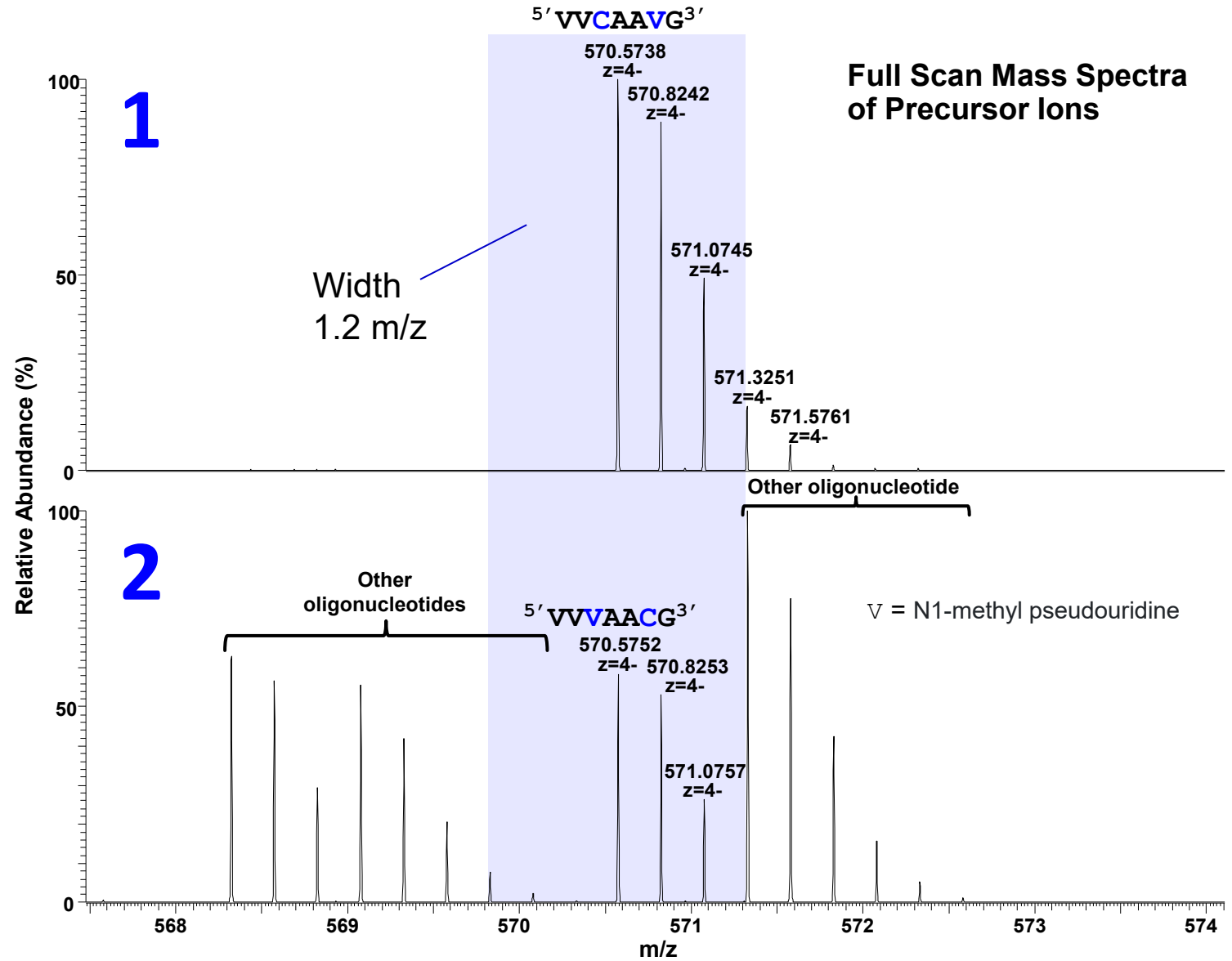
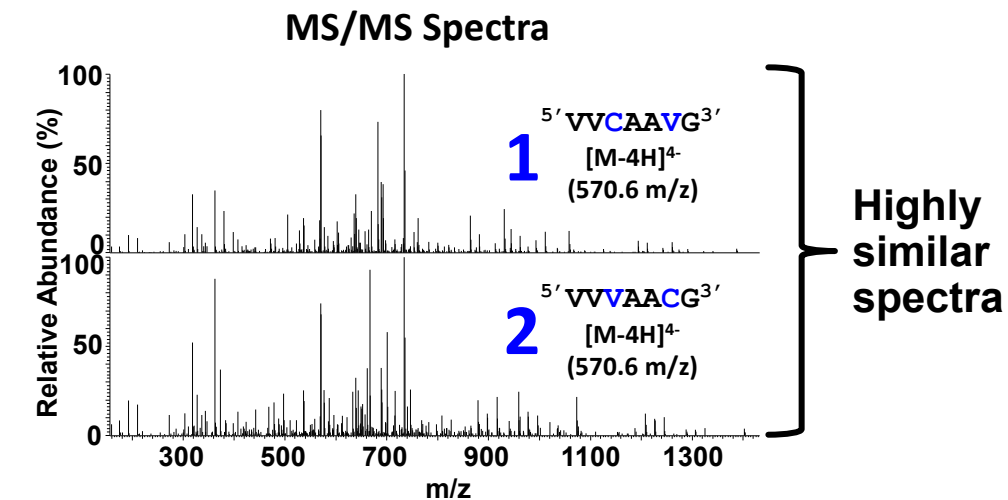
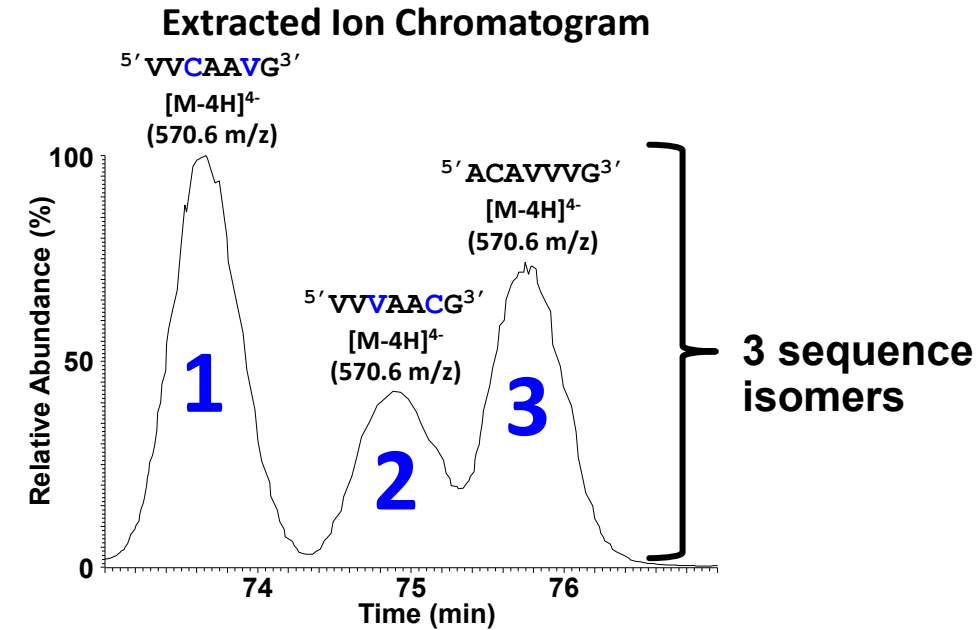
Charge densities are fixed at 0.4 charge / base

∇ = N1-methyl pseudouridine



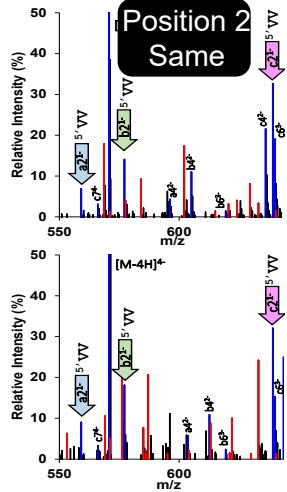
Applying Optimized HCD to Differentiate 2 Sequence Isomers Differing by a Single Exchange in Base Positions

Narrow MS/MS isolation window avoids confounding fragment ions

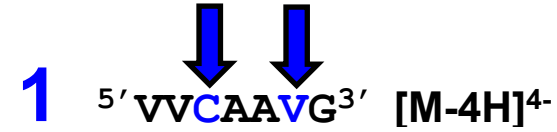


Optimal Fragmentation Enables Differentiation of Highly Similar Sequence Isomers

Observed 5' MS/MS fragments

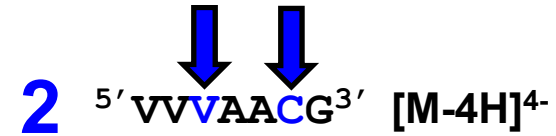


Key — 5' fragment ion (blue), 3' fragment ion (green), Internal fragment ion (red)



Observed 5' fragments				
a	b	c	d	#
		(319.0) ¹⁻	(337.0) ¹⁻	1
(559.1) ¹⁻	(577.1) ¹⁻	(639.1) ¹⁻	(657.1) ¹⁻	2

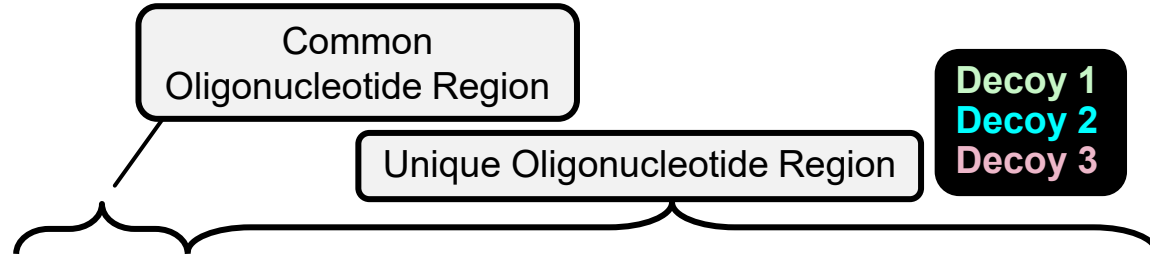
V = N1-methyl pseudouridine



Observed 5' fragments				
a	b	c	d	#
		(319.0) ¹⁻	(337.0) ¹⁻	1
(559.1) ¹⁻	(577.1) ¹⁻	(639.1) ¹⁻		2

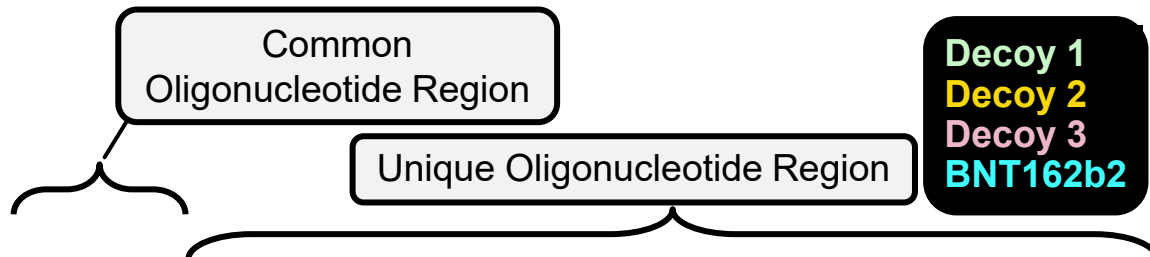
Fidelity of Automated Oligonucleotide Identifications Comprehensively Verified by Decoy Searching

Decoy search **excluding** BNT162b2 mRNA construct

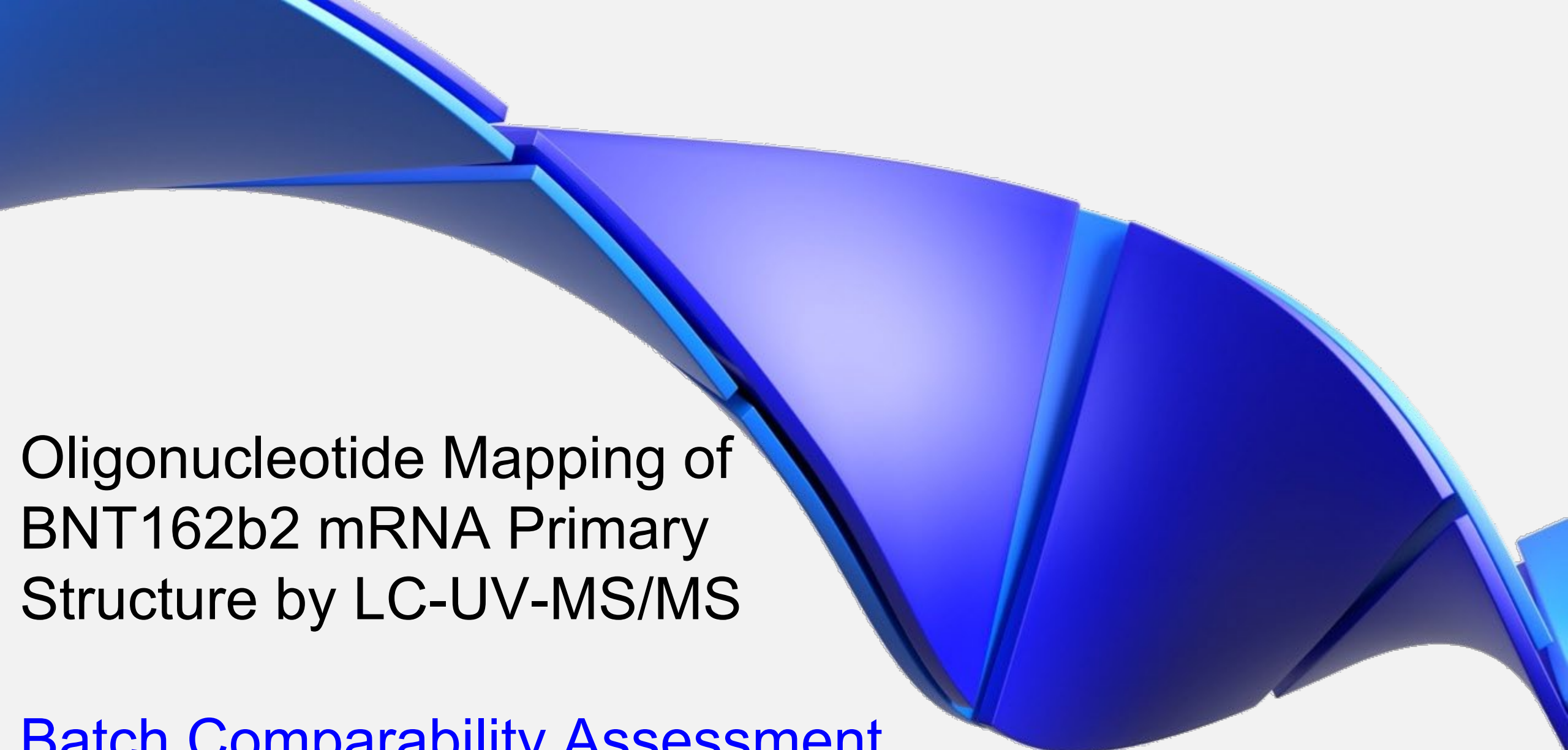


No Preferential Match

Decoy search **including** BNT162b2 mRNA construct



Preferential Match to BNT162b2

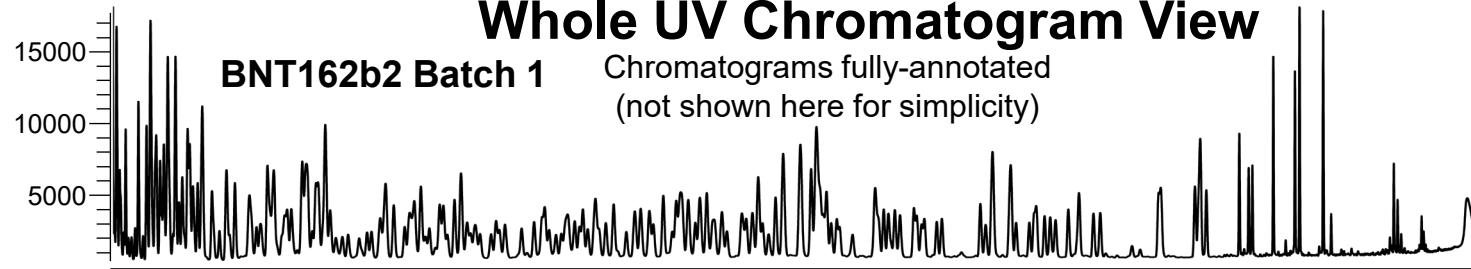


Oligonucleotide Mapping of BNT162b2 mRNA Primary Structure by LC-UV-MS/MS

Batch Comparability Assessment

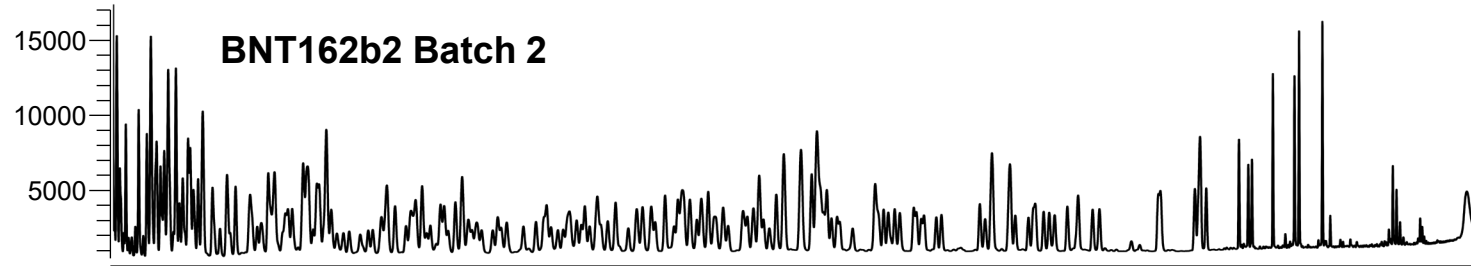
Oligonucleotide Mapping Enables Assessment of mRNA Batch Comparability

Whole UV Chromatogram View

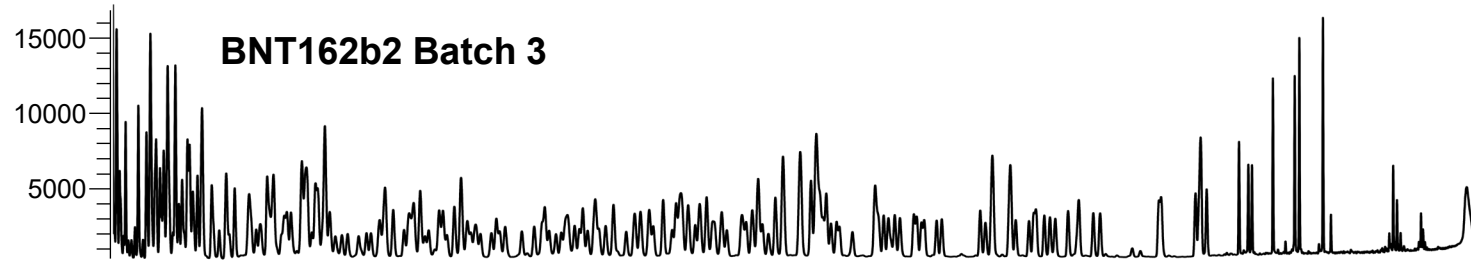


Oligonucleotide Maps Demonstrate Comparability of Multiple BNT162b2 mRNA Drug Substance Batches

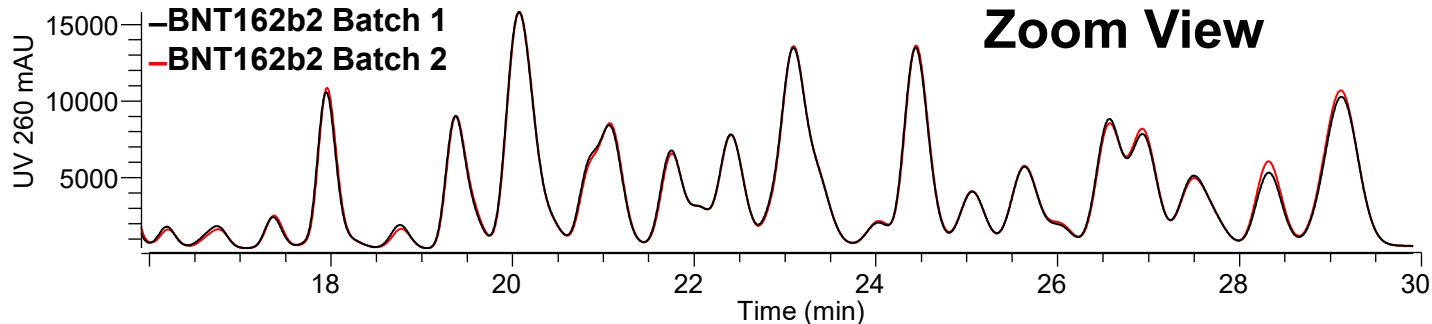
BNT162b2 Batch 2



BNT162b2 Batch 3



Zoom View



- Side-by-side analyses are highly robust
- Chromatographic peaks overlay well

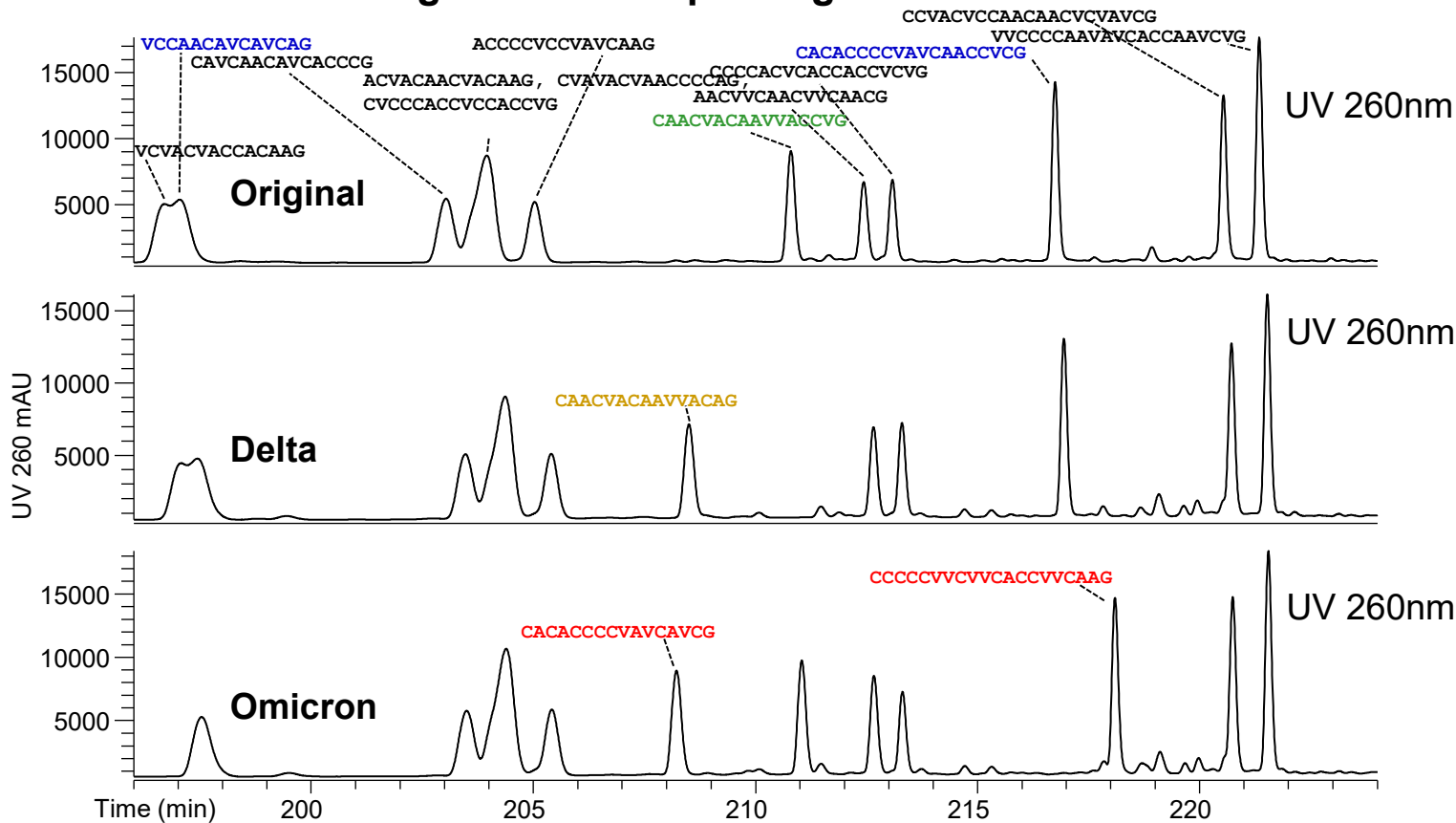


Oligonucleotide Mapping of
BNT162b2 and Variant Construct
Primary Structure by LC-UV-MS/MS

Orthogonal Identity Assessment

Oligonucleotide Mapping Enables Comparison of mRNA for Variant Constructs

Region with Unique Oligonucleotides



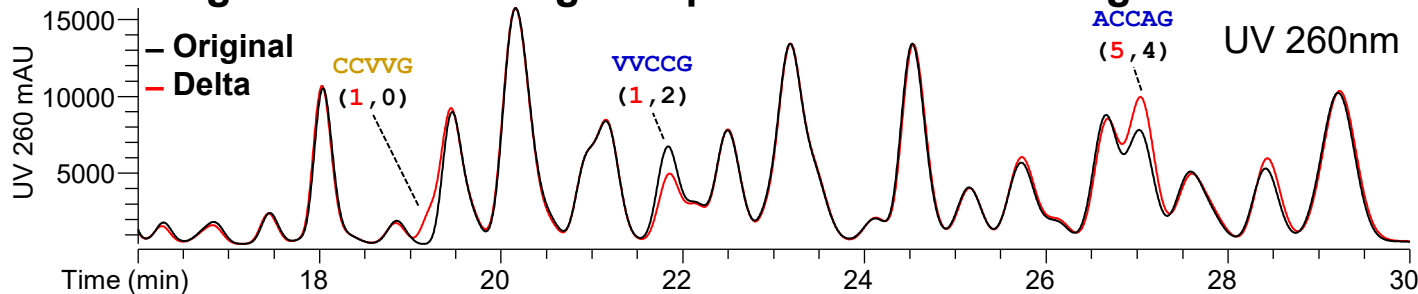
Key

- Present in Original, Delta, & Omicron**
- Present in Original & Delta**
- Present in Original & Omicron**
- Present in Delta**
- Present in Omicron**

Oligonucleotide mapping detects:

- Oligonucleotides unique to one or more constructs
- Differences in number of copies of multi-loci (non-unique) oligonucleotides

Region with Differing # Copies of Multi-Loci Oligonucleotides



Conclusion



- Oligonucleotide mapping via LC-UV-MS/MS directly interrogates the primary structure of RNA, enabling enhanced structural understanding for mRNA vaccines, genetic therapies, and other RNA molecules
- Semi-automated workflow produces a reproducible and fully-annotated oligonucleotide map
 - Fully-annotated LC/UV chromatogram
 - Sequence coverage map (up to 100% sequence coverage - e.g. BNT162b2)
 - Microheterogeneity assessment of 5' terminus capping and 3' terminus poly(A) tail length
- MS/MS fragmentation was optimized and fidelity of identifications verified by decoy sequence searching
- Oligonucleotide mapping assisted the development and commercialization of the Comirnaty® vaccine against SARS-CoV-2
 - Elucidation of Structure (3.2.S.3.1)
 - Comparability (3.2.S.2.6)
 - Data supported regulatory filings to health authorities in 180+ markets
- Gau, B.C., Dawdy, A.W., *et al. Sci Rep* **13**, 9038 (2023)
 - Step-by-step protocol and VBA-enabled data analysis tools are publicly available

Special Thanks

Brian Gau (Pfizer)

Lead oligonucleotide map co-developer & mass spectrometry lead
on Comirnaty mRNA vaccine against SARS-CoV-2

BioNTech

ThermoFisher Scientific

Protein Metrics

Agilent

Waters