

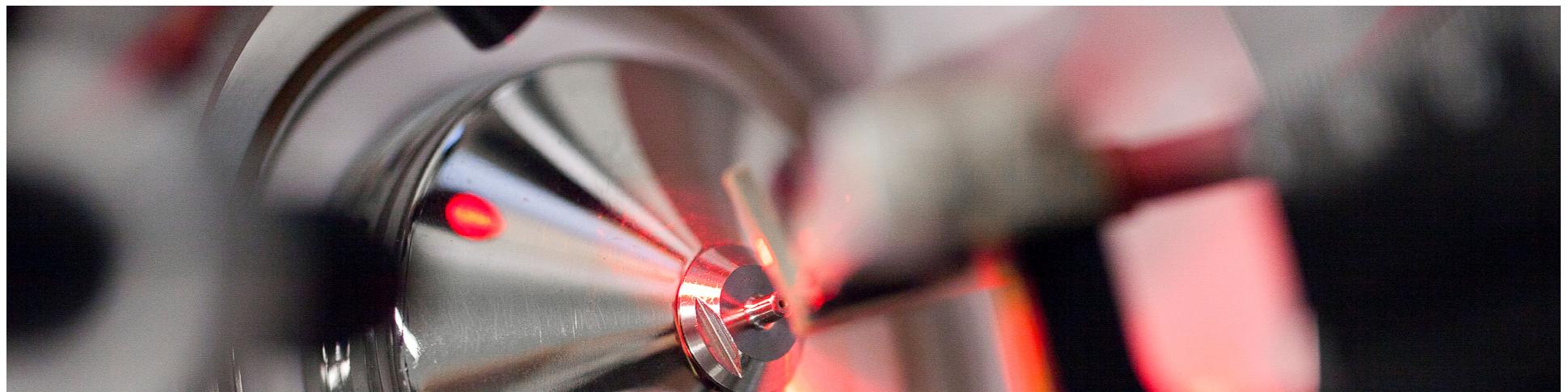
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# The Impact of Immunoglobulin G1 Fc Sialylation on Backbone Amide H/D Exchange

Felix Kuhne

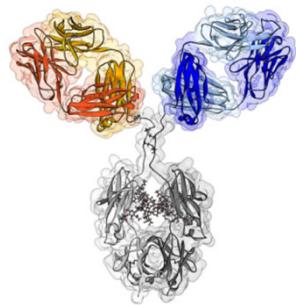
Institute of Hygiene, University of Muenster, Robert-Koch-Strasse 41, 48149, Muenster, Germany

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## Roche Penzberg is a center of excellence for therapeutic proteins

Research and early clinical development of therapeutic proteins



Preclinical research for oncological immunotherapy, molecular information, active ingredient development and tissue-based diagnostics



Providing preclinical and clinical trials with protein therapeutics



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## **Immunoglobulin G1 Fc glycosylation**

- *critical quality attributes (CQAs) affect the mAb structure and function*

## **In vitro glycoengineering (IVGE)**

- *how to generate specifically glycosylated mAbs in vitro*
- *mAb quality attributes and glycan quantification*

## **Structure-function analysis of trastuzumab IVGE**

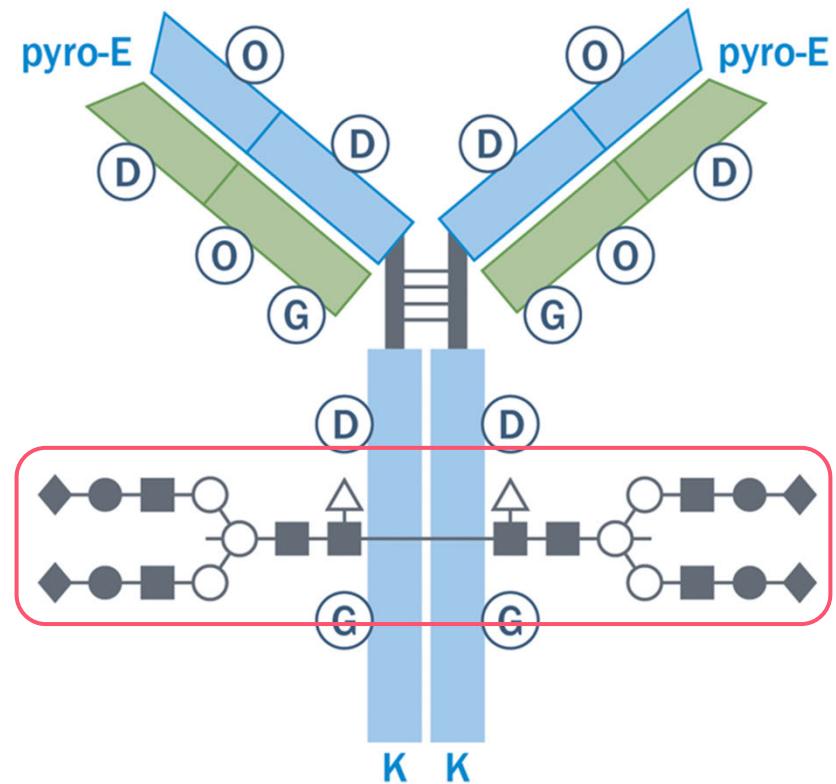
- *higher-order structure effects elicited by mAb Fc glycosylation*
  - *the impact of sialic acid linkages on the mAb conformation*
  - *functional testing and structure to function correlations*
-

## Current status (April 2017) of innovative antibody, Fc fusion protein, and chimeric antigen receptor (CAR) drug candidates

Antibody format	Stage of development			Totals
	Phase I/II	Phase III	Approved for marketing at some point	
Naked IgG	30	51	<b>52</b>	493
Naked antibody fragments	7	2	4	13
Immunocytokines	9	2	0	11
Fc fusion proteins	23	3	11	37
Bispecific antibodies	58	1	2	61
Antibody-drug conjugates	75	9	3	87
Radioimmunoglobulins	13	2	2	17
Antibodies only	575	70	74	719
T or NK cells expressing CAR antibodies	145	0	0	145
Totals	720	70	74	864

**Naked mAbs are glycoproteins (like many other therapeutic proteins, e.g. EPO, IFN-β and Factor VIII)**

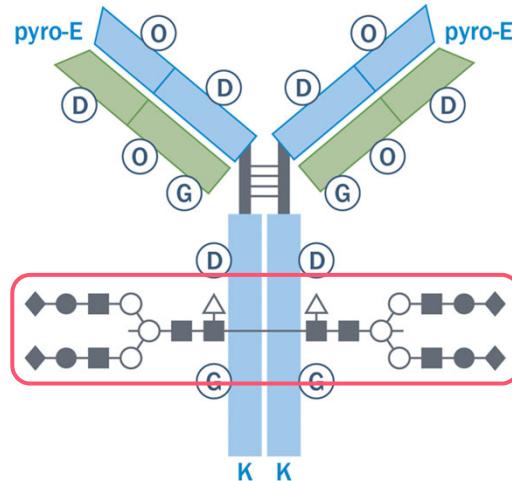
## mAb modification sites



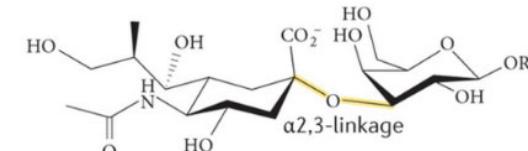
<b>pyro-E</b>	Pyro-Glu (2)
<b>D</b>	Deamidation (3 x 2)
<b>O</b>	Methionine oxidation (2 x 2)
<b>G</b>	Glycation (2 x 2)
	High mannose, G0, G1, G1, G2, (5)
	Sialylation (5)
<b>K</b>	C-term Lys (2)

## mAb Fc glycosylation

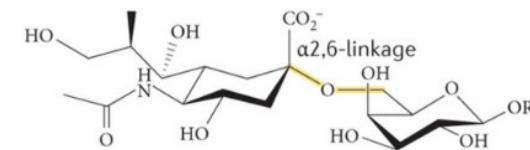
 conformational differences



► CHO cells



► Human



Glycans/glycosylation	Impacts
$\alpha 1\text{-}3\text{-galactose}$ ; $N\text{-glycolylneuraminic acid}$	Immunogenicity
Terminal sialylation	$\downarrow$ binding to $\text{Fc}\gamma\text{RIIIa}$ , $\downarrow$ ADCC; $\uparrow$ PK/PD
Afucosylation	$\uparrow$ binding to $\text{Fc}\gamma\text{RIIIa}$ , $\uparrow$ ADCC, $\uparrow$ ADCP
Galactosylation	$\uparrow$ binding to $\text{C}1\text{q}$ , $\uparrow$ CDC, moderate effect on ADCC
High-mannose	$\downarrow$ PK/PD; $\uparrow$ binding to $\text{Fc}\gamma\text{RIIIa}$ , $\uparrow$ ADCC; $\downarrow$ binding to $\text{C}1\text{q}$ , $\downarrow$ CDC

$\uparrow$  positive impact;  $\downarrow$  negative impact.

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## Immunoglobulin G1 Fc glycosylation

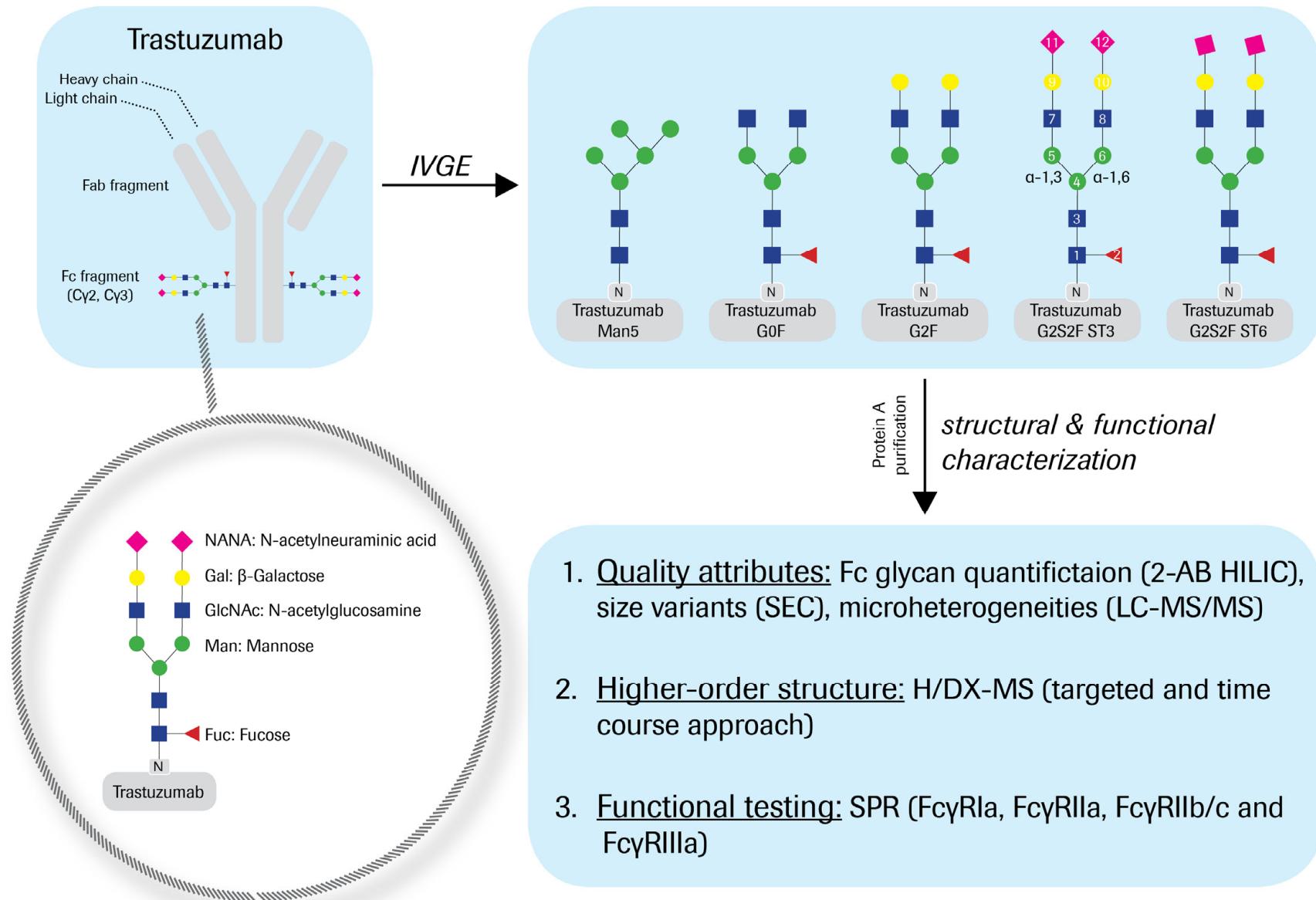
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-



1. Quality attributes: Fc glycan quantification (2-AB HILIC), size variants (SEC), microheterogeneities (LC-MS/MS)

Light chain

Table 1. Relative quantification of 2-AB labeled (trastuzumab) N-glycans (2-AB HILIC).

Fc Glycosylation (%)	Trastuzumab <sup>1</sup>	RM	Man5	G0F	G2F	G2S2F ST3	G2S2F ST6
G0	3.5	3.6	0.5	5.2	0.1	n.q.	0.4
G0F	34.2	39.3	3.4	80.6	n.q.	0.0	0.2
G1	2.8	2.9	1.0	0.1	0.1	0.2	0.7
G1F	41.9	39.0	3.3	1.4	0.4	0.1	0.1
G2	0.4	0.3	0.1	n.q.	5.4	0.5	0.4
G2F	9.3	7.2	0.8	0.7	83.1	1.6	1.8
G2S1	0.1	0.1	n.q.	0.2	n.q.	1.7	3.4
G2S1F	0.9	0.6	0.2	0.2	1.7	13.8	26.5
G2S2	0.1	n.q.	n.q.	n.q.	0.5	3.9	2.8
G2S2F	0.2	0.3	n.q.	0.1	n.q.	60.3	42.9
M3	n.q.	n.q.	0.1	n.q.	n.q.	n.q.	n.q.
M3F	n.q.	n.q.	0.1	n.q.	n.q.	n.q.	n.q.
hM3	0.4	0.6	0.2	1.0	0.0	0.1	0.3
hM3F	0.6	1.2	0.3	2.2	0.0	n.q.	0.2
hM3G1S1	0.1	0.1	n.q.	0.1	0.1	1.9	2.5
hM3G1S1F	0.3	0.4	n.q.	0.6	0.5	4.4	3.7
M4	n.q.	n.q.	0.1	n.q.	n.q.	n.q.	n.q.
hM4	0.3	0.3	n.q.	n.q.	1.0	0.1	1.2
hM4F	0.0	n.q.	n.q.	0.0	2.2	n.q.	n.q.
M5	1.3	1.3	88.3	1.4	1.4	3.1	3.3
hM5	n.q.	n.q.	n.q.	n.q.	n.q.	n.q.	n.q.
M6	n.q.	n.q.	n.q.	n.q.	n.q.	n.q.	n.q.
M7	n.q.	n.q.	n.q.	n.q.	n.q.	n.q.	n.q.
not assigned	3.9	2.8	1.4	6.3	3.5	8.3	9.7

<sup>1</sup> Method reference standard; n.q. = not quantifiable.

1. **Quality attributes:** Fc glycan quantification (2-AB HILIC), size variants (SEC), microheterogeneities (LC-MS/MS)

**Table 2.** Relative quantification of chemical amino acid modifications (LC-MS peptide mapping).

Chemical Mod. (%)	Trastuzumab <sup>1</sup>	RM	Degly	Man5	G0F	G2F	G2S2F ST3	G2S2F ST6
LC N30 <sup>2</sup> deamidation	9.6	8.7	10.9	9.1	9.1	9.0	9.6	10.0
LC N30 <sup>2</sup> succinimide	0.6	0.7	0.7	0.6	0.8	0.7	1.1	1.1
HC N54 <sup>2</sup> deamidation	1.5	1.6	1.7	2.2	1.7	2.0	1.8	1.6
HC N54 <sup>2</sup> succinimide	3.9	4.0	3.9	4.1	3.9	3.9	3.7	3.8
HC D98 <sup>2</sup> isomerization	7.7	7.6	8.0	10.0	7.0	7.7	7.9	8.7
HC D98 <sup>2</sup> succinimide	3.5	4.1	3.6	2.5	4.3	3.8	4.3	4.3
HC N389/390 <sup>3</sup> deam.	2.0	1.9	2.2	1.6	1.9	1.9	2.1	2.0
HC N389/390 <sup>3</sup> succ.	1.7	1.7	1.7	2.0	1.8	1.7	1.8	1.8
HC M252 <sup>3</sup> oxidation	2.3	2.6	3.4	5.2	3.0	3.8	3.8	3.8

<sup>1</sup> Method reference standard; <sup>2</sup> Kabat numbering [59]; <sup>3</sup> EU numbering [4].

**Table 3.** Relative quantification of trastuzumab size variants (SEC-UV).

Mol. Weight Species (%)	Trastuzumab <sup>1</sup>	RM	Degly	Man5	G0F	G2F	G2S2F ST3	G2S2F ST6
Monomer	99.8	99.6	99.6	99.4	99.6	99.3	99.2	99.1
total HMW <sup>2</sup>	0.2	0.4	0.4	0.5	0.3	0.7	0.8	0.9
total LMW <sup>3</sup>	0.0	0.0	0.1	0.1	0.1	0.0	0.0	0.1

<sup>1</sup> Method reference standard; <sup>2</sup> high molecular weight species; <sup>3</sup> low molecular weight species.

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## In vitro glycoengineering (IVGE)

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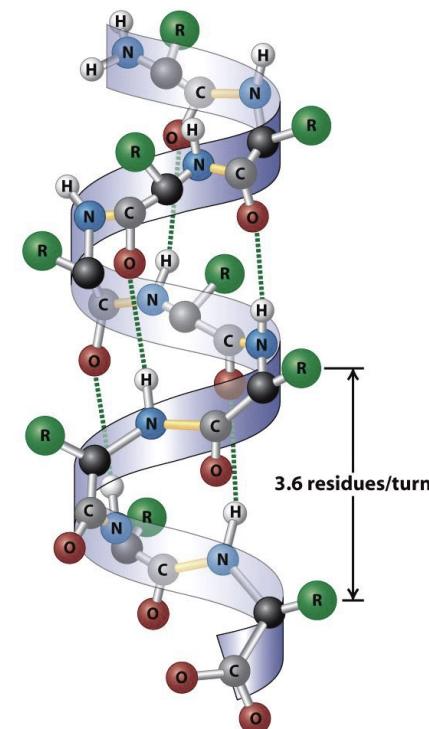
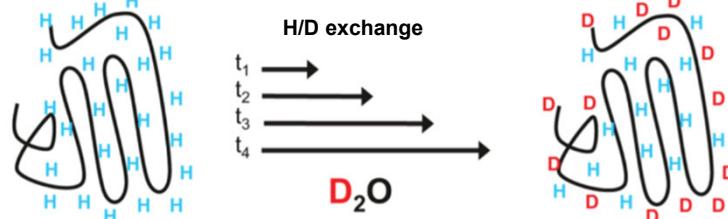
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  - *the impact of sialic acid linkages on the mAb conformation*
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- 1. Quality attributes:** Fc glycan quantification (2-AB HILIC), size variants (SEC), microheterogeneities (LC-MS/MS)
- 2. Higher-order structure:** H/DX-MS (targeted and time course approach)

- according to the protein dynamics the intramolecular H-bonds of the peptide backbone open and close more or less frequently
- dependent on molecular dynamics and interactions H-bonds can be more or less solvent-accessible

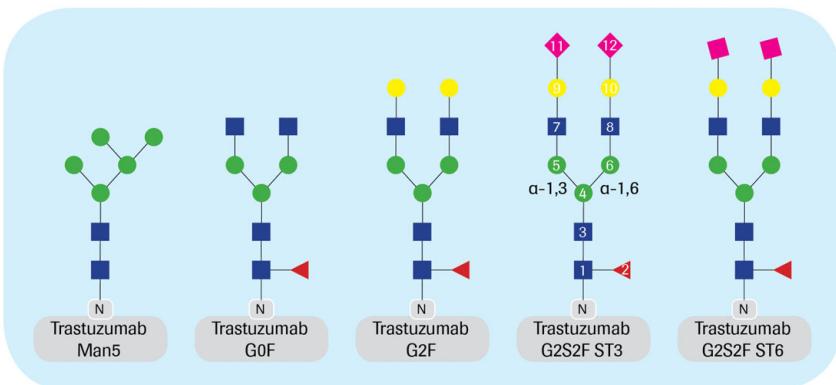
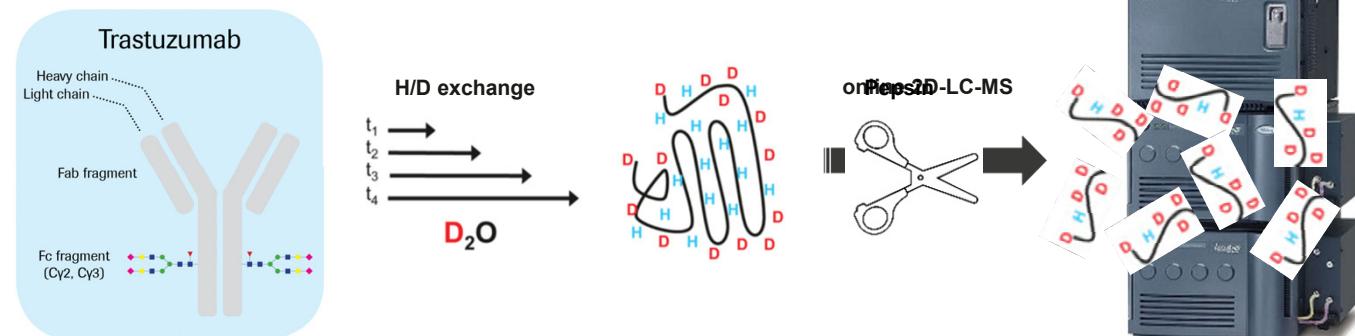
**H/D exchange correlates with the conformational dynamics.**



Rand, K. D., Zehl, M. & Jorgensen, T. J. Measuring the hydrogen/deuterium exchange of proteins at high spatial resolution by mass spectrometry: overcoming gas-phase hydrogen/deuterium scrambling. *Acc Chem Res* **47**, 3018-3027, doi:10.1021/ar500194w (2014).

Berg, J. M., Stryer, L. & Tymoczko, J. L. *Stryer Biochemie*. (Springer-Verlag, 2015)

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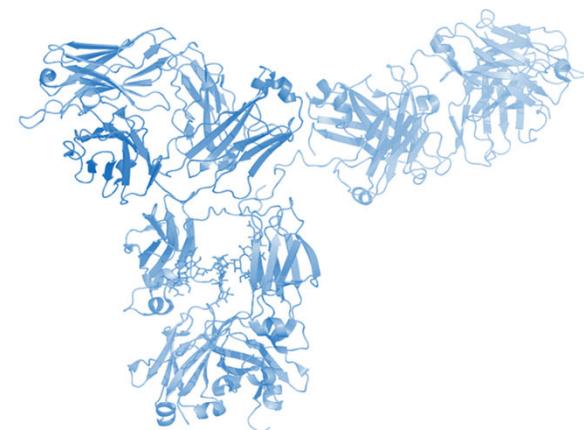
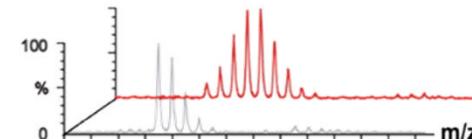
Kuhne, F. et al. The Impact of Immunoglobulin G1 Fc Sialylation on Backbone Amide H/D Exchange. *Antibodies (Basel)* **8**, doi:10.3390/antib8040049 (2019).

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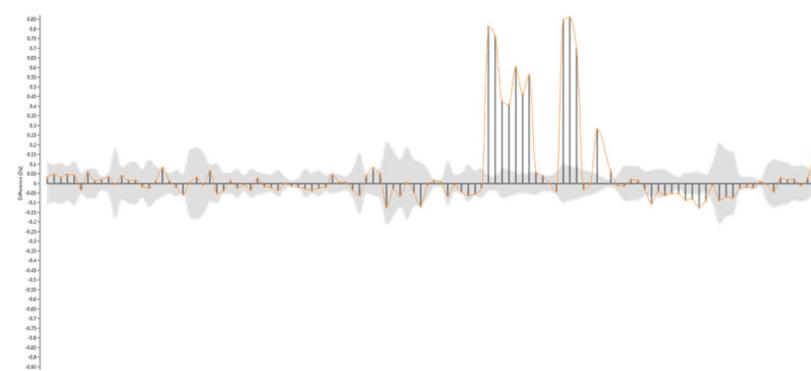
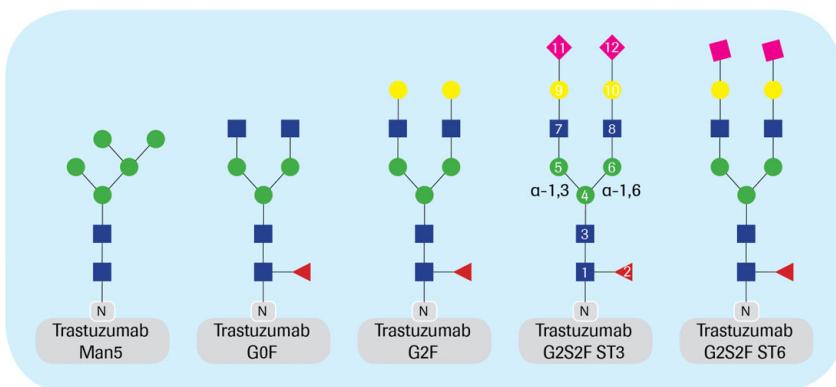
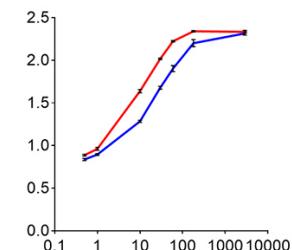
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data analysis

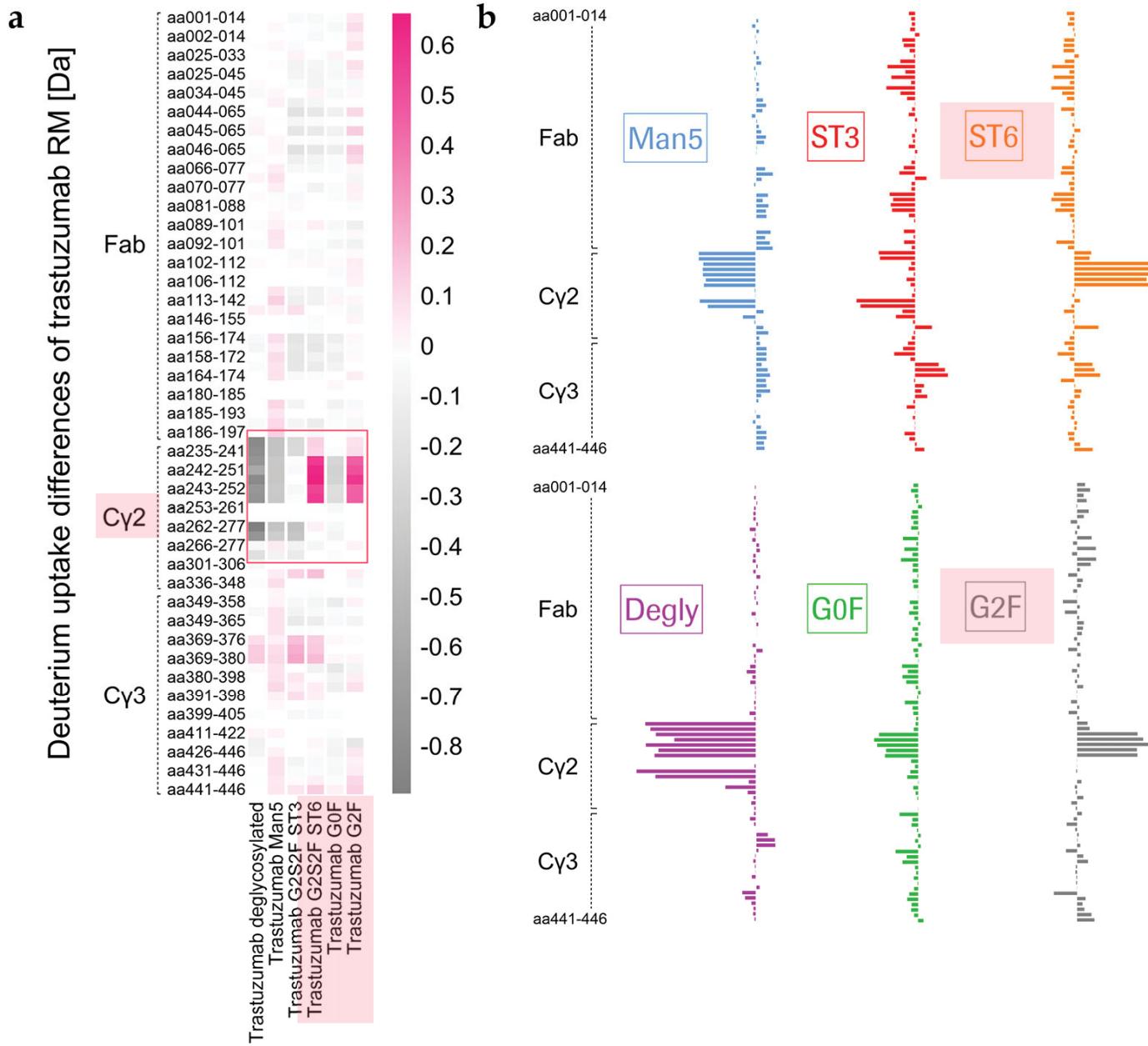


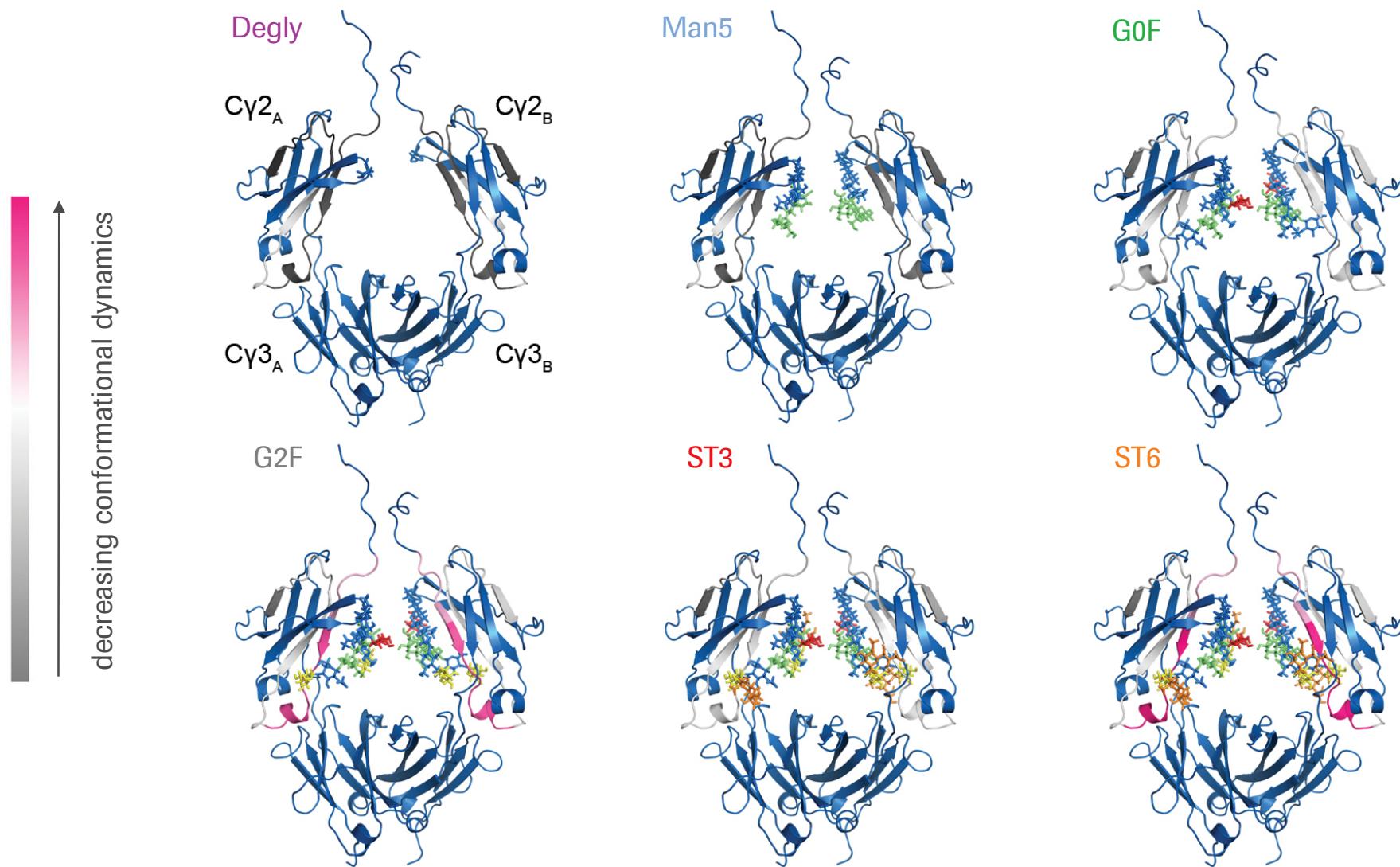
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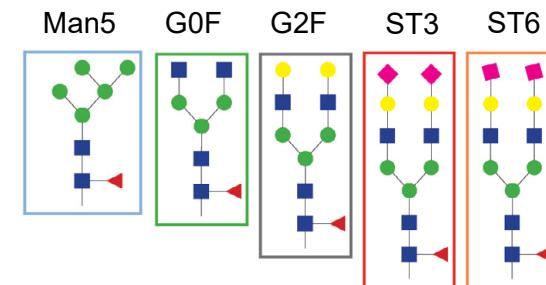
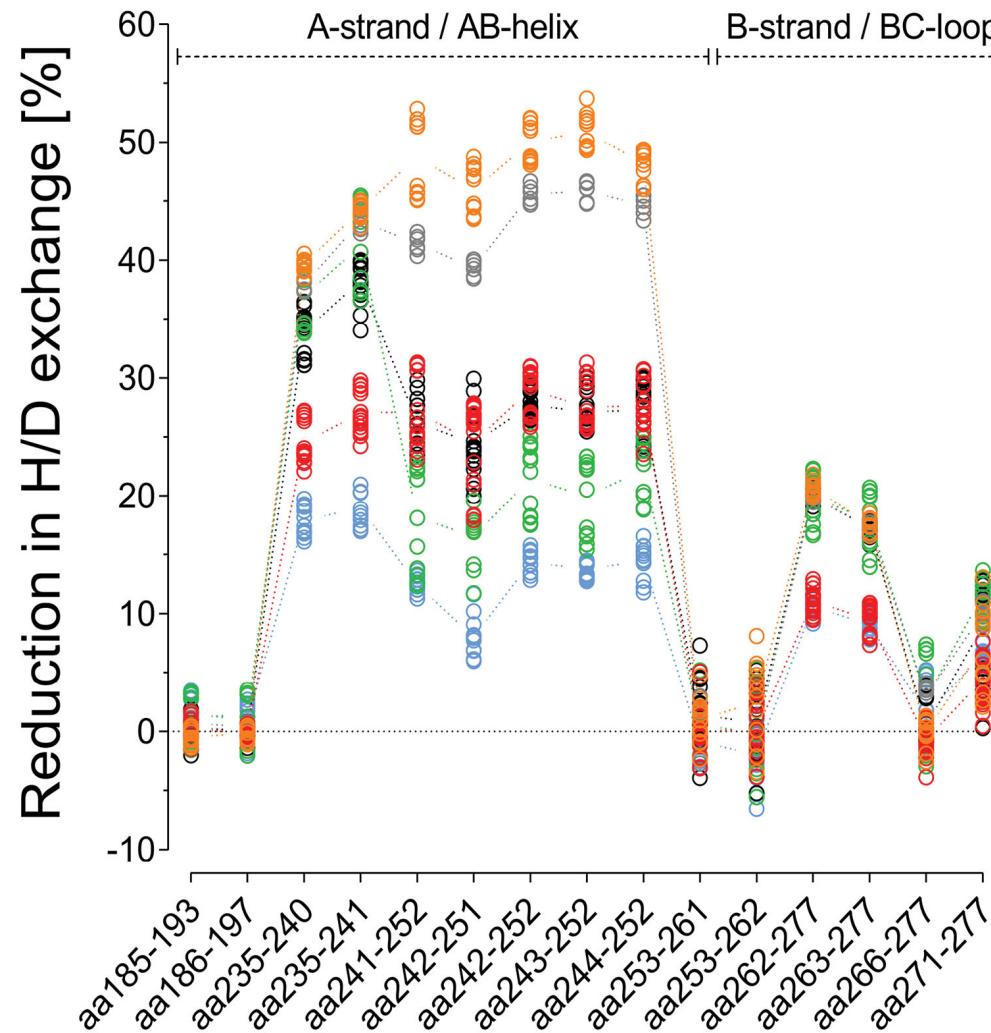
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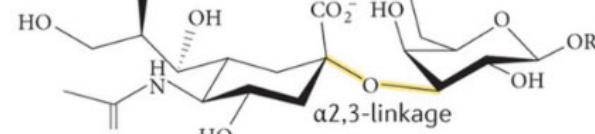




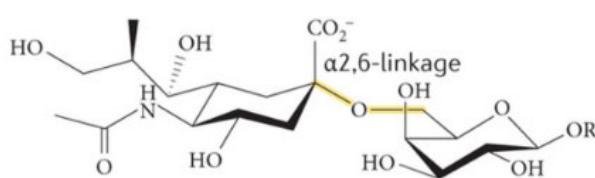
## Antibody heavy chain (Cy2 domain) dynamics vary dependent on Fc glycosylation



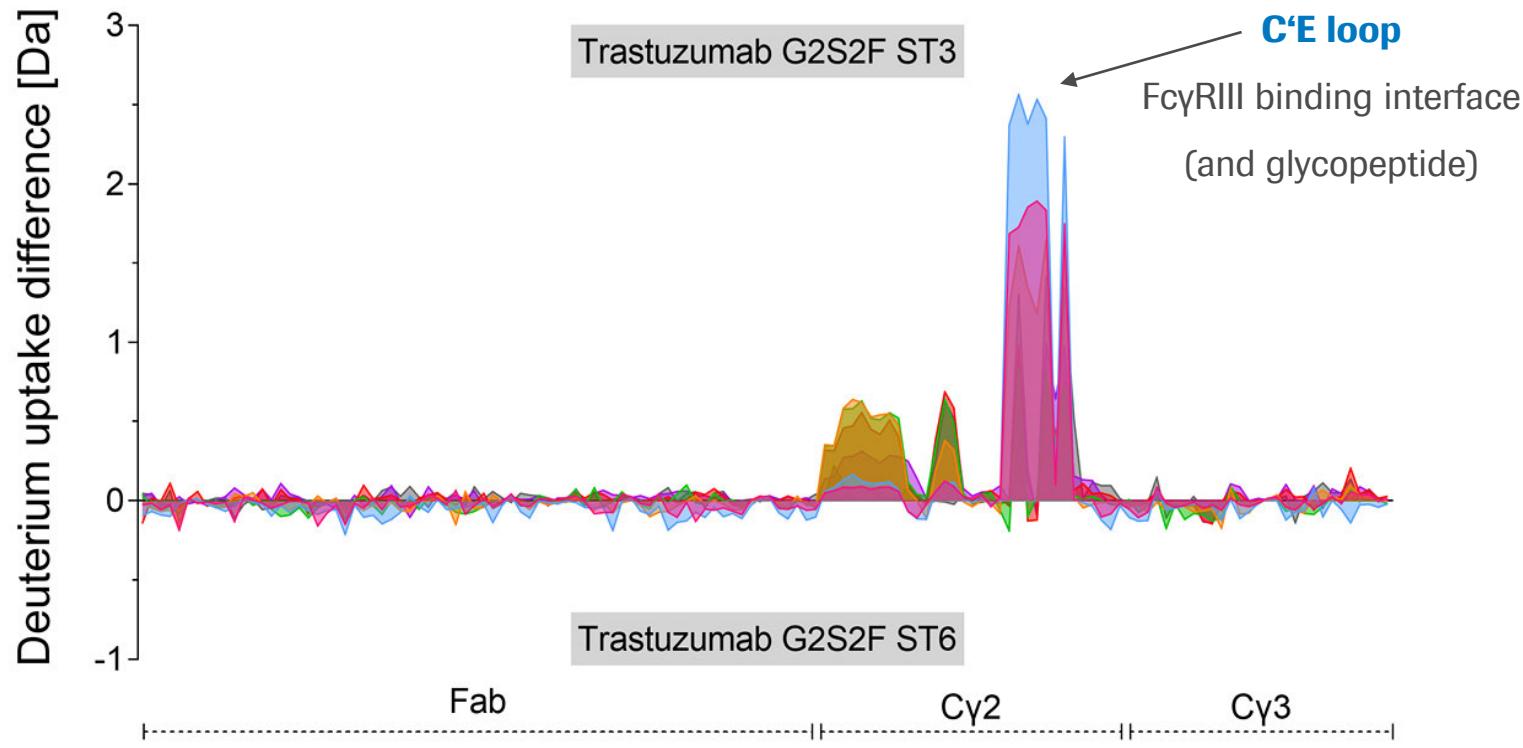
► CHO cells



► Human

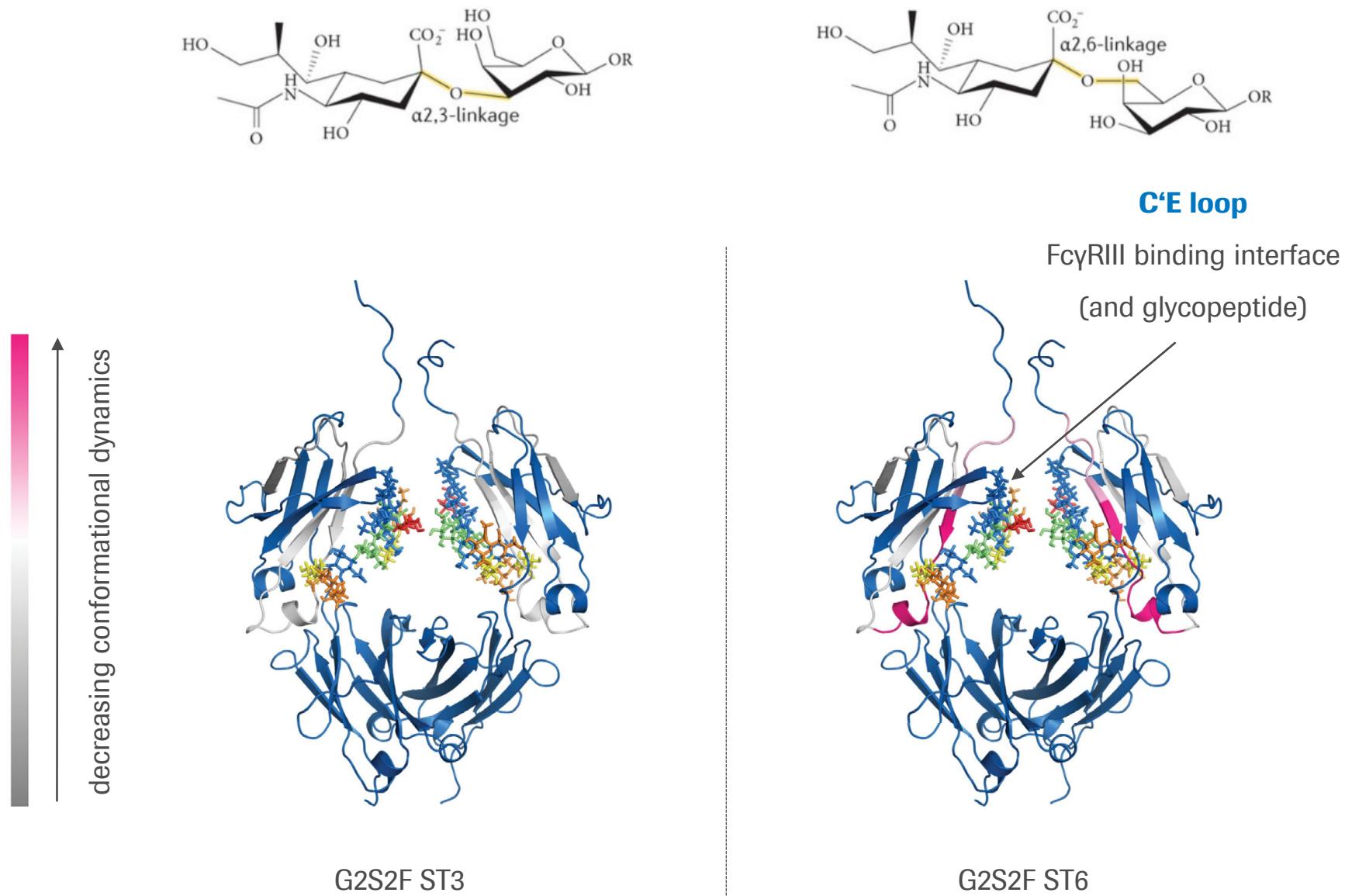


## The type of sialic acid linkage impacts the Fc (Cy2) higher-order structure

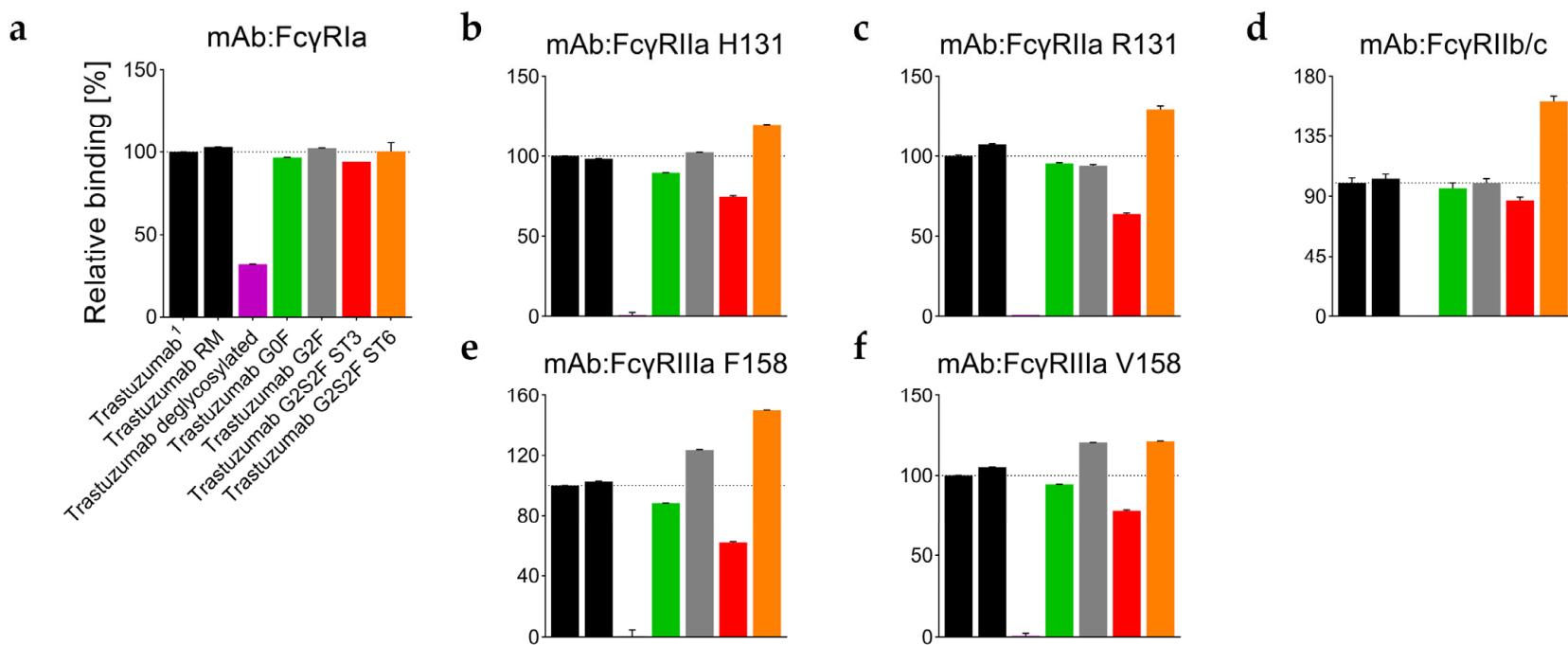


Uptake difference plot of H/DX time course experiment performed for:

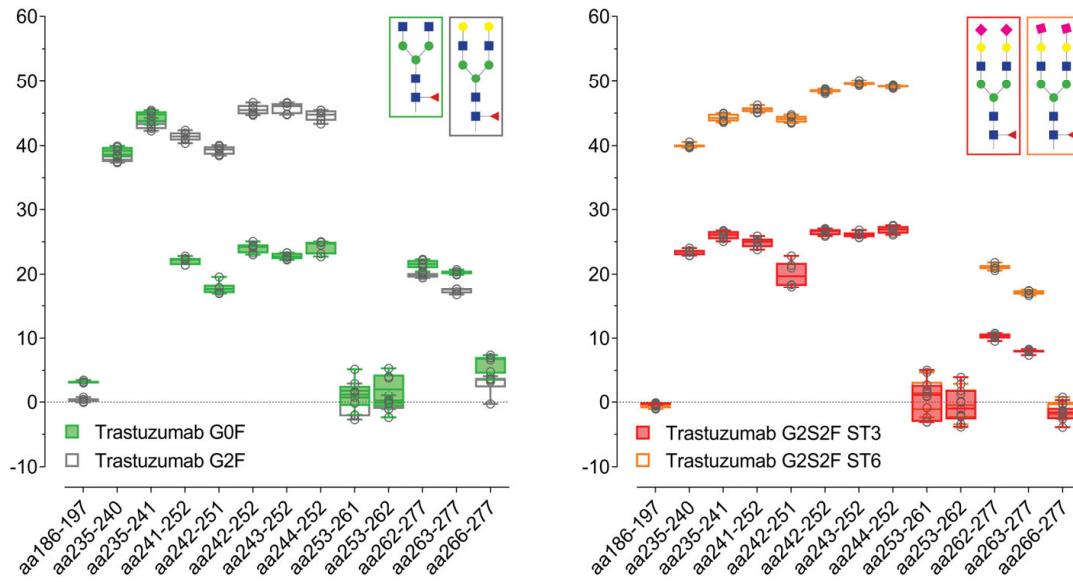
0.5 min (pink), 1 min (light blue), 10 min (orange), 30 min (green), 1 h (red), 3 h (purple), and 48 h (black).



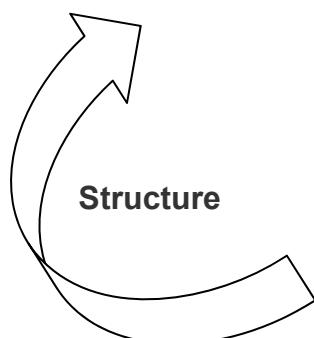
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- 3. Functional testing:** SPR (Fc $\gamma$ R $I\alpha$ , Fc $\gamma$ R $II\alpha$ , Fc $\gamma$ R $IIb/c$  and Fc $\gamma$ R $III\alpha$ )



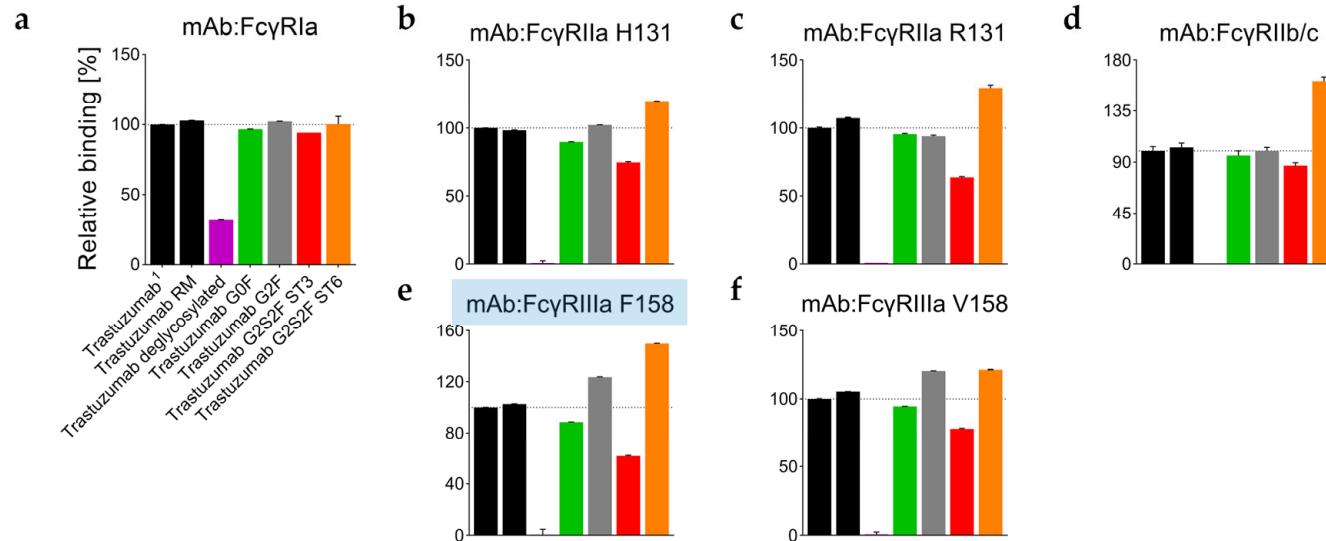
## H/DX-MS



Function

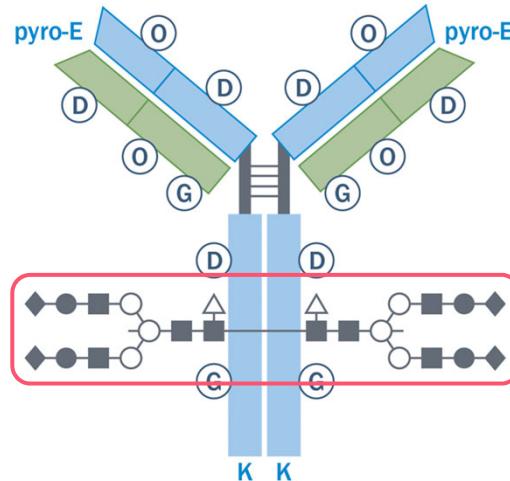


## SPR

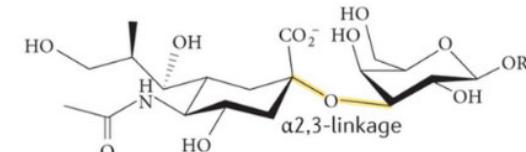


## mAb Fc glycosylation

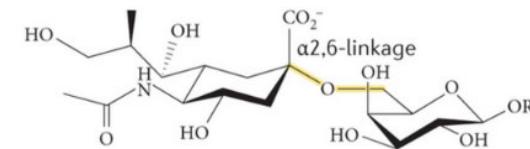
P  
conformational differences  
&  
functional differences



► CHO cells



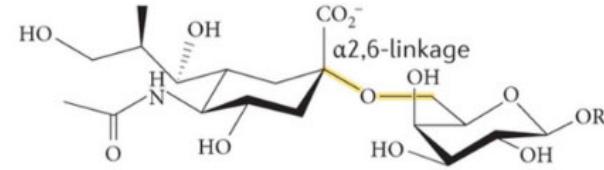
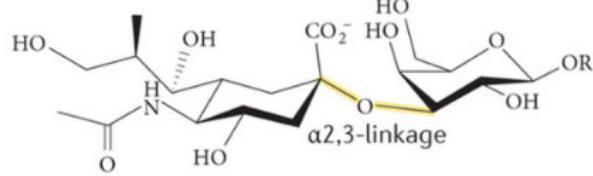
► Human



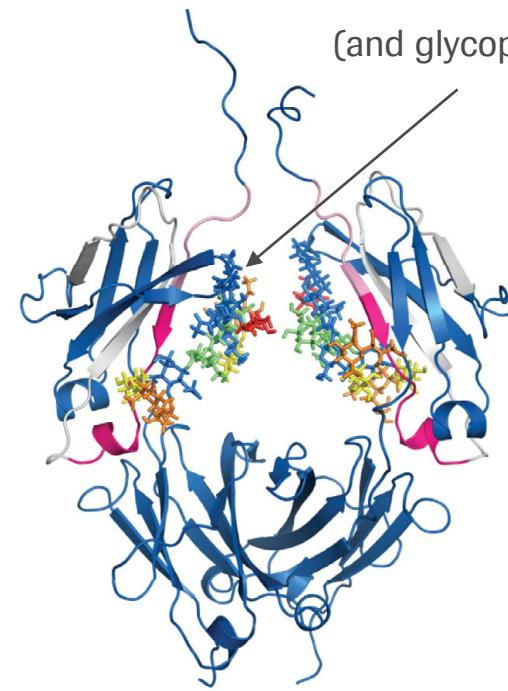
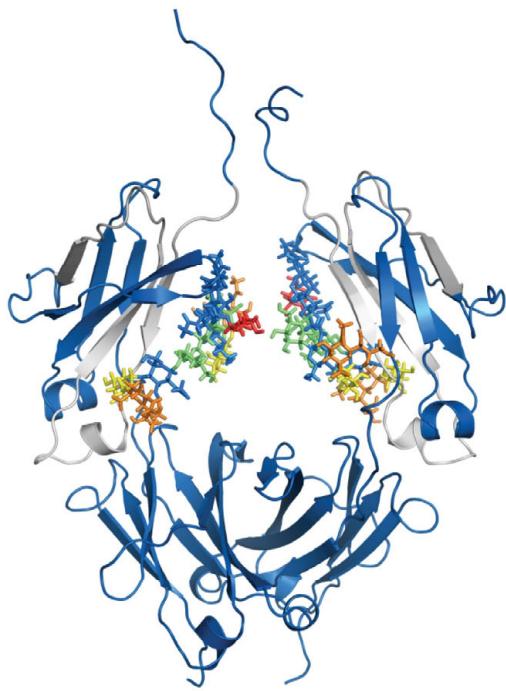
needs to be  
differentiated

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High-mannose	$\downarrow$ PK/PD; $\uparrow$ binding to $Fc\gamma RIIIA$ , $\uparrow$ ADCC; $\downarrow$ binding to $C1q$ , $\downarrow$ CDC

$\uparrow$  positive impact;  $\downarrow$  negative impact.

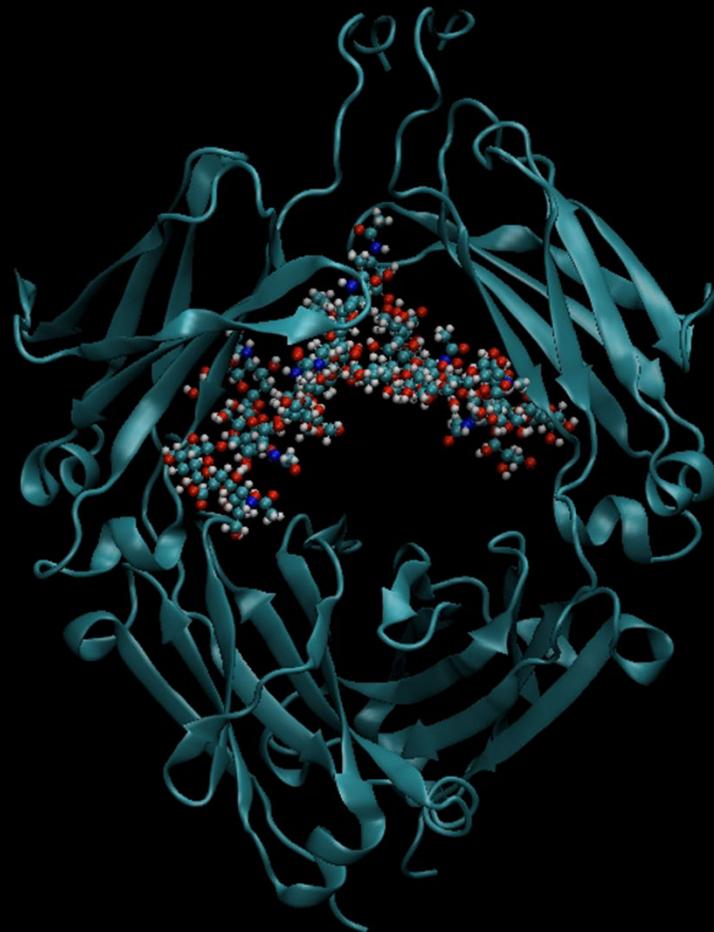


# HOW?



## Outlook

- molecular dynamics
- receptor interaction
- covalent labeling





***Doing now what patients need next***