A Novel, Rapid Chip-Based iCIEF-MS Analysis of Therapeutic mAb Charge Variants Under Forced Degradation Conditions and Comparison to Traditional Methods

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Background and Challenge Statement

- Charge heterogeneity analysis is essential for the successful development and production of therapeutic antibodies.
- Traditionally, charge variant analysis and peak identification are performed by two separate offline assays (cIEF/IEX and Mass Spectrometry).
- MS identification of a charge variants requires a tedious, multi-step process including fractionation, sample preparation, and off-line MS analysis. Additionally, IEX (which is the current preferred charge variant method at Merck) can suffer from extensive method development, column robustness, and long run times.





- A cIEF-MS platform can turn a multistep (3-4 steps) workflow into a one step analysis
- Following CQA determination by more in-depth methods (RPM) monitoring levels of deamidation or oxidation for extended timepoint studies (e.g., stability, forced degradation) can be done more efficiently



Intabio proprietary microchip technology integrates key iCIEF-MS analytical functions



Blaze real-time Imaged cIEF separation

- Whole column, UV absorbance during focusing & mobilization
- Stepped voltage and current control during focusing enable realtime feedback
- Enables rapid cIEF method development & peak tracking during mobilization





Major degradation pathways of recombinant monoclonal antibodies and Traditional Assays to Monitor them¹



¹Nowak et al (2017) Forced degradation of recombinant monoclonal antibodies: A practical guide, mAbs, 9:8, 1217-1230



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★ LC/MS ★ cIEF

Combining cIEF and MS analysis can successfully cover all degradation pathways of interest in a single shot.



Outline of cIEF-MS Evaluation of Forced Degradation Study





Traditional cIEF Analysis of Merck mAb using in-house methods following forced degradation conditions.





¹Du, Yi et al. (2012) Chromatographic analysis of the acidic and basic species of recombinant monoclonal antibodies." *mAbs* vol. 4,5, 578-85.











Major Modification Sites Identified by Reduced Peptide Mapping

| | N terminal Pyroglutamat e | Deamidation | Oxidation | Oxidation | C-terminal Lysine Retention |
|-------------------|---------------------------------|---------------|---------------|---------------|-----------------------------------|
| | N-terminus (Q) | Asparagine #1 | Methionine #1 | Methionine #2 | C-terminus (K) |
| No Stress (411) | 97.9 | 3.2 | 0.5 | 2.2 | 9.2 |
| H50 –D0 (412) | 97.9 | 3.4 | 0.7 | 2.0 | 8.3 |
| H50 – D1 (413) | 97.8 | 3.9 | 0.8 | 2.2 | 8.3 |
| H50 – D3 (414) | 97.4 | 3.6 | 0.9 | 2.4 | 8.6 |
| H50 – D7 (415) | 97.6 | 4.8 | 1.0 | 3.2 | 9.1 |
| LG – 0X (417) | 97.5 | 4.0 | 1.0 | 2.8 | 8.6 |
| LG – 0.5X (418) | 97.5 | 3.8 | 3.6 | 17.2 | 8.7 |
| LG – 1X (419) | 97.5 | 4.1 | 7.2 | 39.7 | 7.4 |
| OT001 – H0 (420) | 97.6 | 4.0 | 0.9 | 2.2 | 7.0 |
| OT001 – H6 (421) | 97.2 | 3.9 | 1.1 | 5.2 | 7.8 |
| OT001 – H24 (422) | 97.5 | 4.3 | 2.3 | 19.2 | 7.6 |
| P10 – D0 (423) | 97.6 | 4.4 | 1.0 | 3.6 | 7.7 |
| P10 – D1 (424) | 97.3 | 11.0 | 1.0 | 3.3 | 7.7 |
| P10 – D3 (425) | 97.5 | 23.7 | 1.0 | 3.9 | 7.4 |
| P10 – D7 (426) | 97.4 | 42.8 | 1.0 | 4.0 | 7.0 |



>95% sequence coverage



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| H50 – D3 (414) | 97.4 | 3.6 | 0.9 | 2.4 | 8.6 |
| H50 – D7 (415) | 97.6 | 4.8 | 1.0 | 3.2 | 91 |
| LG – 0X (417) | 97.5 | 4.0 | 1.0 | 2.8 | 8.6 |
| LG – 0.5X (418) | 97.5 | 3.8 | 3.6 | 17.2 | 8.7 |
| LG – 1X (419) | 97.5 | 4.1 | 7.2 | 39.7 | 7.4 |
| OT001 – H0 (420) | 97.6 | 4.0 | 0.9 | 2.2 | 7.0 |
| OT001 – H6 (421) | 97.2 | 3.9 | 1.1 | 5.2 | 7.8 |
| OT001 – H24 (422) | 97.5 | 4.3 | 2.3 | 19.2 | 7.6 |
| P10 – D0 (423) | 97.6 | 4.4 | 1.0 | 3.6 | 7.7 |
| P10 – D1 (424) | 97.3 | 11.0 | 1.0 | 3.3 | 7.7 |
| P10 – D3 (425) | 97.5 | 23.7 | 1.0 | 3.9 | 7.4 |
| P10 – D7 (426) | 97.4 | 42.8 | 1.0 | 4.0 | 7.0 |



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| LG – 1X (419) | 97.5 | 4.1 | 7.2 | 39.7 | 7.4 |
| OT001 – H0 (420) | 97.6 | 4.0 | 0.9 | 22 | 7.0 |
| OT001 – H6 (421) | 97.2 | 3.9 | 1.1 | 5.2 | 7.8 |
| OT001 – H24 (422) | 97.5 | 4.3 | 2.3 | 19.2 | 7.6 |
| P10 – D0 (423) | 97.6 | 4.4 | 1.0 | 3.6 | 7.7 |
| P10 – D1 (424) | 97.3 | 11.0 | 1.0 | 3.3 | 7.7 |
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| P10 – D7 (426) | 97.4 | 42.8 | 1.0 | 4.0 | 7.0 |



Change in acidic variant abundance as seen is cIEF analysis can most likely be attributed to deamidation at the PENNY peptide



Blaze iCIEF-MS Analysis of Charge Variants – Non-stressed Control



Blaze iCIEF-MS Analysis of Charge Variants – Non-stressed Control

- Mass shifts of 162 Da denote the galactose series of GOF, G1F and G2F glycans
- Basic charge variants with a mass addition of 128 Da correspond to unprocessed lysine(s)
- Acidic charge variants correspond to sialic acid additions

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Blaze iCIEF-MS Analysis of Non-stressed Control Main Peak Putative Modifications



Blaze analysis of the 3 light stressed conditions shows comparable cIEF profiles to Merck cIEF analysis



Comparable Quantitation of Blaze ciEF with Merck In-House Data





Blaze iCIEF-MS Analysis OX Control



Blaze iCIEF-MS Analysis of 0.5X Light Stress Shows 1-2 Oxidation Sites



Blaze iCIEF-MS Analysis of 1X Light Stress Shows 3-4 Oxidation Sites



Comparison of Oxidation Levels by iCIEF-MS vs. Peptide Mapping

| Light Stressed samples | % of oxidized protein measured with blaze CiEF-MS analysis | % of oxidized amino acid residue with blaze CiEF- MS analysis | Average % oxidized Methionine (Methionine #1 & Methionine #2) with RPM | % of oxidation of Methionine #1 (RPM) | % of oxidation of Methionine #2 (RPM) |
|------------------------------|---|---|--|---|--|
| OX | 2.81% | 2.53% | 1.90% | 1.00% | 2.80% |
| 0.5X | 20.16% | 13.59% | 10.40% | 3.60% | 17.20% |
| 1x | 36.65% | 25.56% | 23.45% | 7.20% | 39.70% |



Increase in Dimer Species is observed by iCIEF-MS in Light Stress Material and Verified with Size Exclusion Chromatography



iCIEF Analysis of High pH Samples on Blaze vs. in-house iCIEF





Blaze iCIEF-MS Analysis of all Charge Variants - High pH Day 7

| Common Acidic Modifications | | | | | |
|------------------------------------|-------------|--|--|--|--|
| Sialic Acid | | | | | |
| Deamidati | Deamidation | | | | |
| Trisulfide bo | onds | | | | |
| Thiosulfide Mod | ification | | | | |
| Cysteinylat | ion | | | | |
| Free thio | s | | | | |
| Fragment | S | | | | |

- Succinimide 2X higher in high pH day 7, compared to control.
- No additional shift in mass from acidic to main peak identified suggesting presence of deamidation (+1 Da, unresolved on TOF instrument), agreeing with RPM data.



Conclusions

- 1. Complete Blaze iCIEF-MS analysis in < 20 minutes
- 2. Rapid method develop and easy method transfer from legacy iCE instruments, <1 day
- 3. iCIEF electropherograms from Intabio Blaze analysis and in-house analysis are comparable throughout control and all stressed conditions.
- 4. Analysis of non-stressed material identifies expected modifications including glycosylation series (G0F/G0F, G0F/G1F, etc), unprocessed lysines, loss of GlcNAc, and sialic acid additions.
- 5. iCIEF-MS analysis of light-stressed samples identifies increasing oxidative states of mAb (1-4 oxidation sites). Total amount of oxidation observed generally matches RPM analysis. Increase in the presence of dimer is observed across all electrophoretic peaks of 1X light-stressed material.
- 6. RPM identified one main deamidation site in high-pH stress (Day 7). cIEF data shows an increase in acidic peaks but intact mass unable to resolve +1 Da mass shift. Analysis on higher resolution MS may better resolve this mass shift.

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