



European Federation of Pharmaceutical  
Industries and Associations

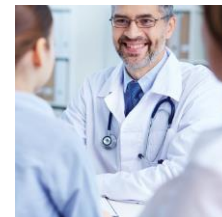


# Moving Towards ICH Q5A(R2): Use of Next Generation Sequencing for Viral Safety Testing of Biotechnology Products

Elodie Charbaut Taland, Merck KGaA, on behalf of EFPIA Supportive Group  
“Clonality, Characterisation and Viral Safety of Cell Lines”



CASSS CMC Strategy Forum EU  
October 16, 2023



# Presentation Outline

1. CMC Applications of Next Generation Sequencing (NGS)
2. Biomanufacturing Supportive Group: Clonality, Characterisation and **Viral Safety** of Cell Lines
3. Objective of the Position Paper on Next Generation Sequencing for Viral Safety Testing of Biotechnology Products
4. Status of the Position Paper

# CMC Applications of Next Generation Sequencing

## Next Generation Sequencing

A massively parallel technology used to determine DNA or RNA sequences offering ultra-high throughput, scalability, and speed for a variety of applications

## Main CMC Applications

Cell Banks  
Viral Seeds  
In-Process Controls  
Drug Substance

**Viral Safety Testing**  
Genotypic Characterisation  
Clonality Evidence  
Identity, Purity  
Stability

Recombinant Proteins, Vaccines, ATMPs

## ICH Q5A(R2) Concept Paper (2019)

Technological advances since the publication of the original ICH Q5A(R1) Guideline have occurred that require additional discussion. Specifically, nucleic acid-based assays such as Polymerase Chain Reaction (PCR) and **Next Generation Sequencing (NGS)** may provide rapid and sensitive detection of adventitious and endogenous viruses

# Clonality, Characterisation and Viral safety of Cell Lines

The EFPIA MQEG Biomanufacturing Supportive Group was initiated in 2019.

## Project Aim

- Facilitate the implementation of advanced analytical **NGS technologies** in relation with clonality, cell line characterization and viral safety for **biotechnological products** by sharing and leveraging industry expertise / experience and best practices.
- As a first priority, develop, promote and provide guidance for implementation of NGS as an alternative for current standards on **viral safety testing**
- Present industry position in perspective of shaping regulatory landscape for smoother implementation and acceptance by Health Authorities

## Scope

- Technology of interest: NGS
- Applicable for viral safety testing on Biotechnological products (**ATMPs out of scope**)
- **EU, US, ICH/WHO** regulatory landscapes

## Deliverables

- Drafting of a Position Paper: acting as a practical implementation guide by providing an industry position for discussion with the Regulatory Authorities.
- Presentation of the Industry position within congresses/ conferences

# 16 Companies Participate to the NGS Supportive Group

abbvie

 NOVARTIS

AMGEN

Johnson & Johnson

 PathoQuest

MERCK

Roche

AstraZeneca 

Synthon

 Biogen.

GSK

Lilly

 MSD

 ucb

 Pfizer

sanofi

efpia  5

# Objectives of the Position Paper

Envisaged as a practical implementation guide providing a position from Industry for dialogue with Regulatory Authorities.

## Completion

100%

### Introduction: Why NGS?

Characteristics of NGS technologies and comparison with current virus safety tests

100%

### Validation strategy for NGS methods

Assessing **Analytical Comparability** of NGS with *in vivo* / *in vitro* virus safety tests

95%

### Regulatory strategy for NGS implementation

Regulatory lifecycle management of NGS methods

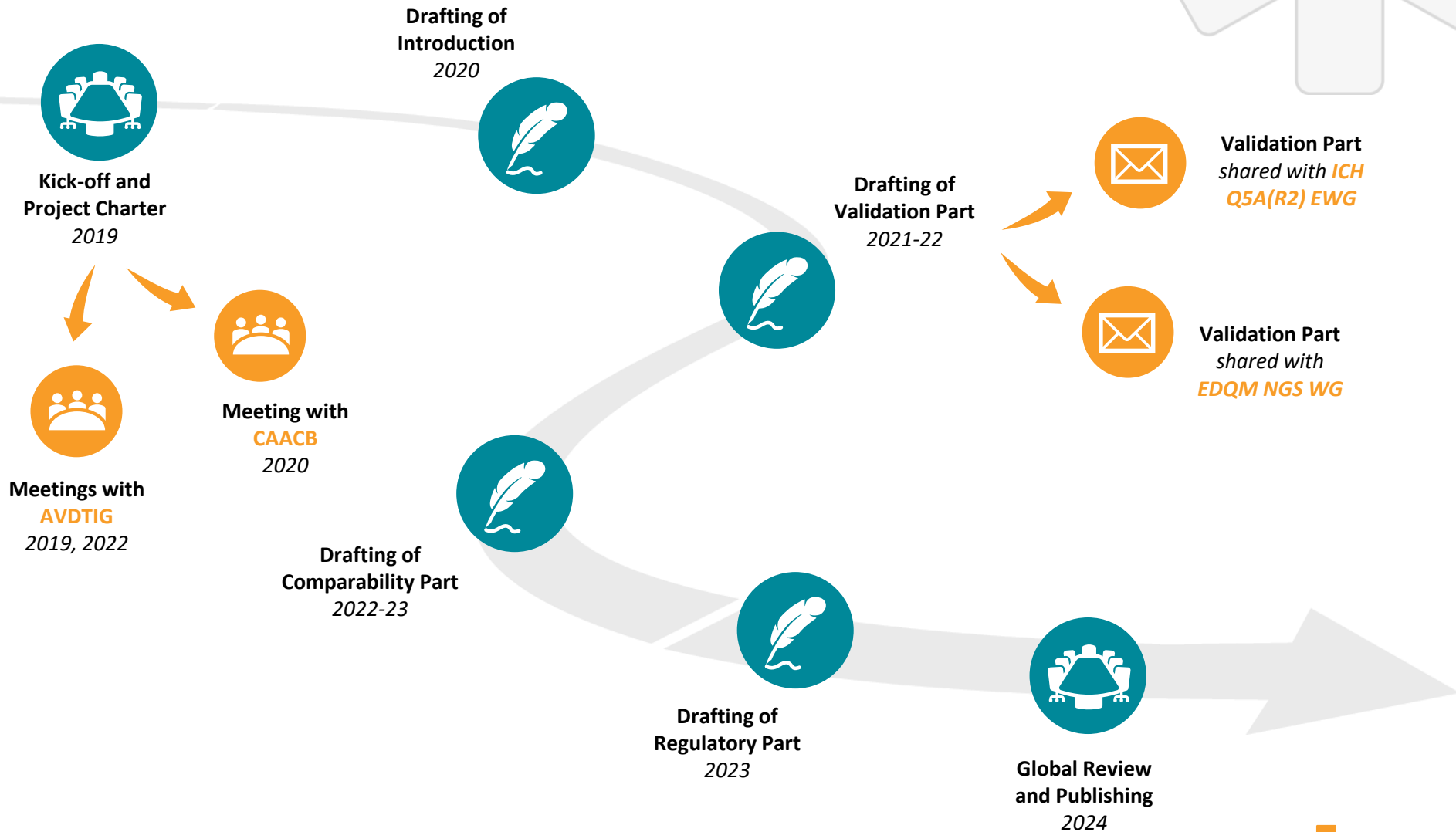


Support drafting of ICH Q5A(R2), as needed

Influence potential drafting of a Compendial General Chapter

Support creation of an ICH Q2(R2) Annex for NGS

# Progress of the NGS Supportive Group Work



# Status of the Position Paper

## Introduction

- Key **features** and expected performance of **conventional virus detection assays** (*in vivo*, *in vitro*, retrovirus), as well as molecular assays such as **PCR and NGS** for the purpose of virus detection
- Comparative table highlighting their **complementarity and orthogonality**
- Initial considerations for introducing NGS-based methods for virus detection, possibly as replacement of routine assays
- **To add:** the description of different **NGS method formats** (genomics, transcriptomics, viromics)

## Validation of NGS-based Methods

- **Validation parameters:** Specificity, Limit of Detection, (Breadth of detection)
- Specific challenges related to specificity and LOD determination
- Limitations of validation studies and necessity to use **representative model viruses**
- Validation strategies: end-to-end vs step-by-step, **reference materials** depending on the NGS method format
- Elements coming from **method development:** robustness, bioinformatic analysis, System Suitability Test
- **Life-cycle management** considerations: e.g. viral genome database update, partial revalidation



# Status of the Position Paper

## Comparability with Current Tests

- Discussion on significance and limitations of an **analytical comparability study** when replacing a conventional virus test by NGS
- **General methodology** to identify potential challenges for a comparability study
- **Specific examples** are presented in more detail: in vivo, in vitro infectivity tests, PCR
- **Key challenges to analytical comparability**: differences in Critical Quality Attributes, absence of validation data or limitations to their comparison, result interpretation

## Regulatory Implications

- Existing **regulatory pathways and requirements** to register NGS for viral safety testing (clinical vs commercial, IPC vs Cell Banks vs DS)
- **Regulatory strategies** that can be used to facilitate acceptance of NGS (depending on product lifecycle stage and intended purpose of the method) e.g. PACMP, HA interactions
- Opportunities to use **technology-driven regulatory mechanisms**, e.g. Innovative Technology Forums (ETT, QIG), future Platform Technology pathways



**Group Lead**

Lionel  
Randon

**Drafting Subteam Leads**

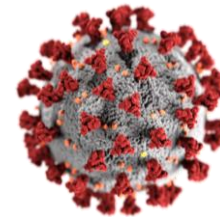
1. Christopher Frye, Luhong He, Elodie Charbaut Taland
2. Alessia Bachmann, Simone Olgiati
3. Elodie Charbaut Taland
4. Morgane Rochemont

J. Auer  
C. Azimpour  
S. Balaji  
S. Bartels  
M. Bednar  
P. Beurdeley

A. Bhogle  
C. Braxton  
K. Cai  
J.P. Cassart  
C. Cecil



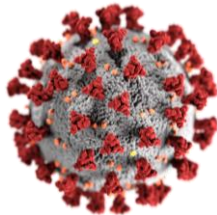
A.S. Colinet  
C. Cote  
K. Daris  
N. Deneyer  
Z. Dragic  
M. Eloit



L. Fan  
L. Grassi  
T. Hartman  
D. Hatton  
D. Hickman

M. Hinchliffe  
S. Jacques  
F. Lay  
C. Lambert  
S. Lemaire  
C. Logvinoff

H. Newton  
S.M. O'Donnell  
N. Salehi  
K. Smith  
J. Stevens



Thank you



A. Stokes  
O. Vandeputte  
M. Wisher  
B. Yang  
Y. Zhu