

Analytics for Process-related Impurities in Viral Vector Manufacturing

Alla Zilberman, Ph.D.

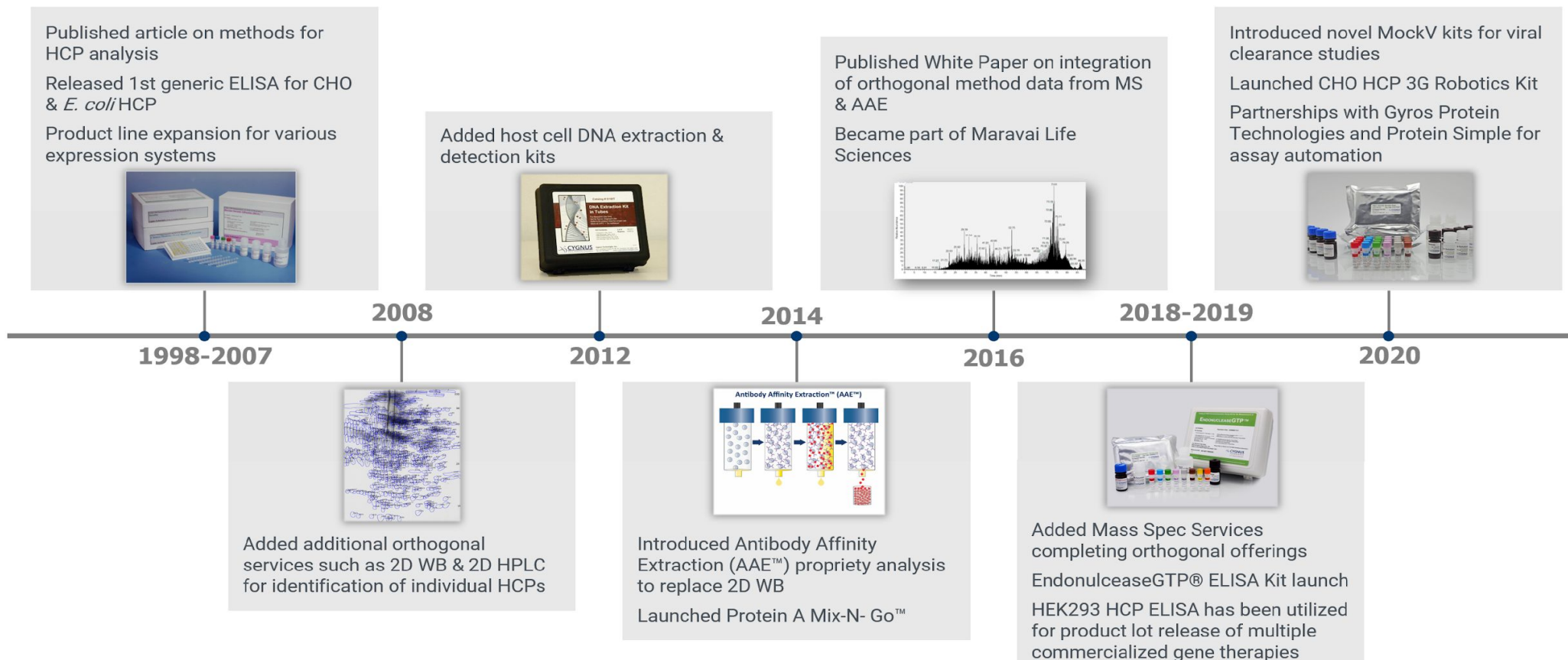
Vice President of Technical Marketing

CASSS CGTP 2023



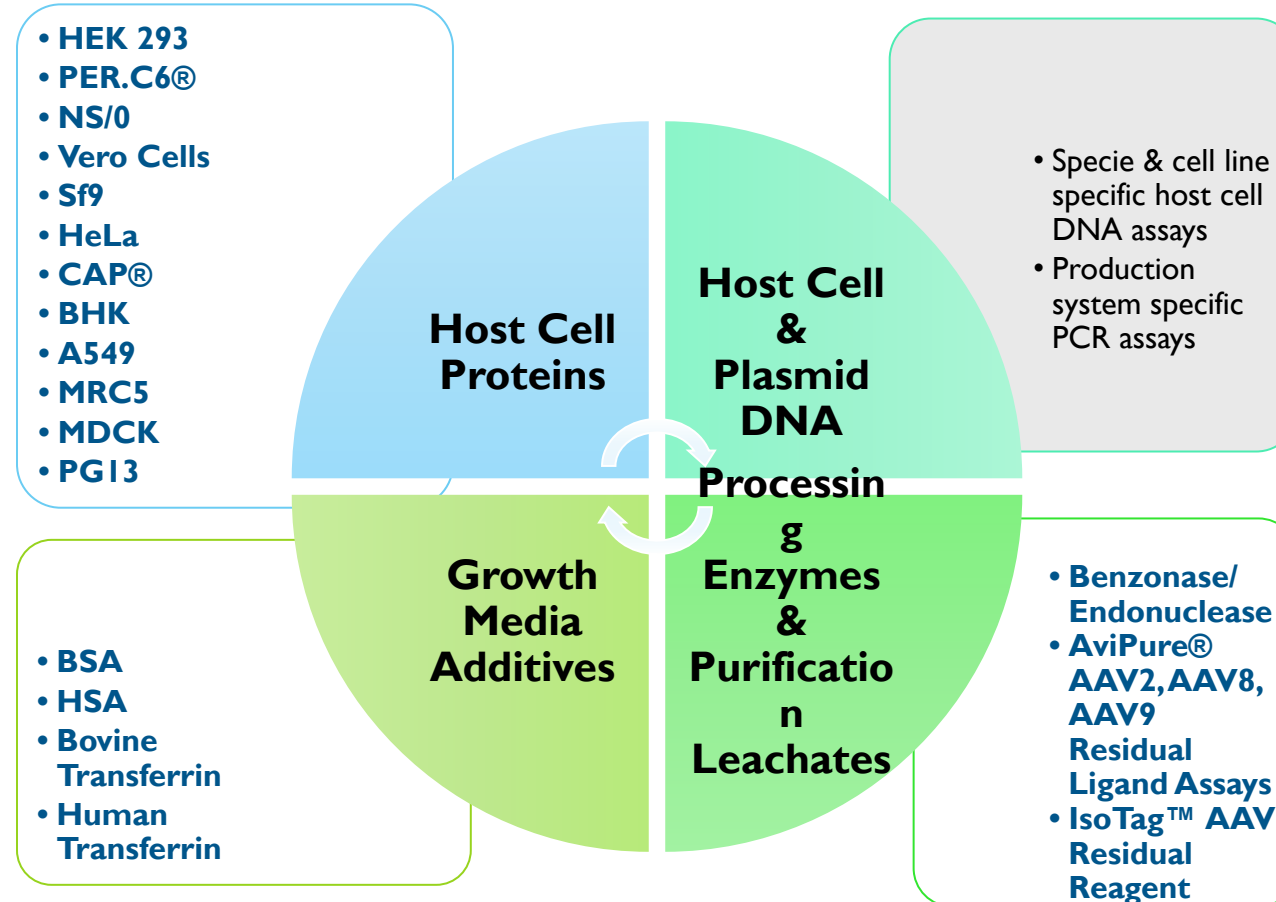
CYGNUS
TECHNOLOGIES

part of Maravai LifeSciences



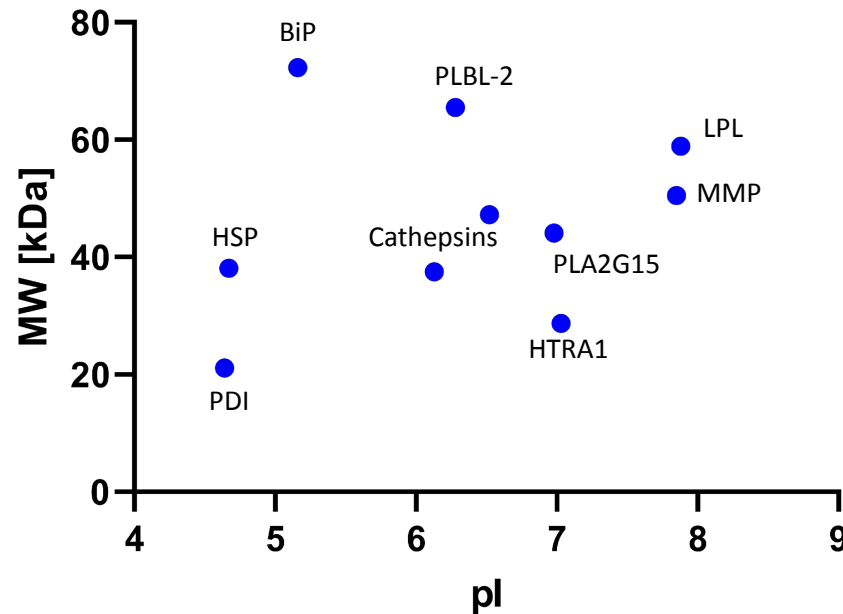
Driving your success at every stage.

- **What did we learn from mAb bioprocess?**
 - HCP Impurities are a real safety concern [immunogenicity, DS stability]
 - Updated USP and European guideline are more harmonized for mAb and rProteins (<100 pm with a trend to <10 ppm for mAbs)
 - Regulators are fully aware of limitations of current methods and now reference AAE and Mass Spec as acceptable orthogonal methods
 - HCP assessment is not a “Check the box ” exercise
- **FDA Guidance for Industry:** CMC Information for Human Gene Therapy IND
Applications require testing for process impurities such as HCP, HC + “production system” DNA, and other process-related impurities (BSA, Benzonase/Endonuclease, AAV affinity resins leachates)



Developed by and available from Cygnus = blue font

Potentially Problematic HCPs



Drug Degradation

Serine Protease
HTRA1
Cathepsins
Matrix
Metalloproteinase

Drug Aggregation

Protein Disulfide
Isomerase
Endoplasmic reticulum
chaperone BiP
Heat shock proteins

Lipases

Lysophospholipase 2
Lysophospholipase-like I
Lysosomal acid lipase A

Immunogenicity and Loss of ELISA dilutional linearity

Phospholipase B domain
containing 2

PS20 PS80 Degradation

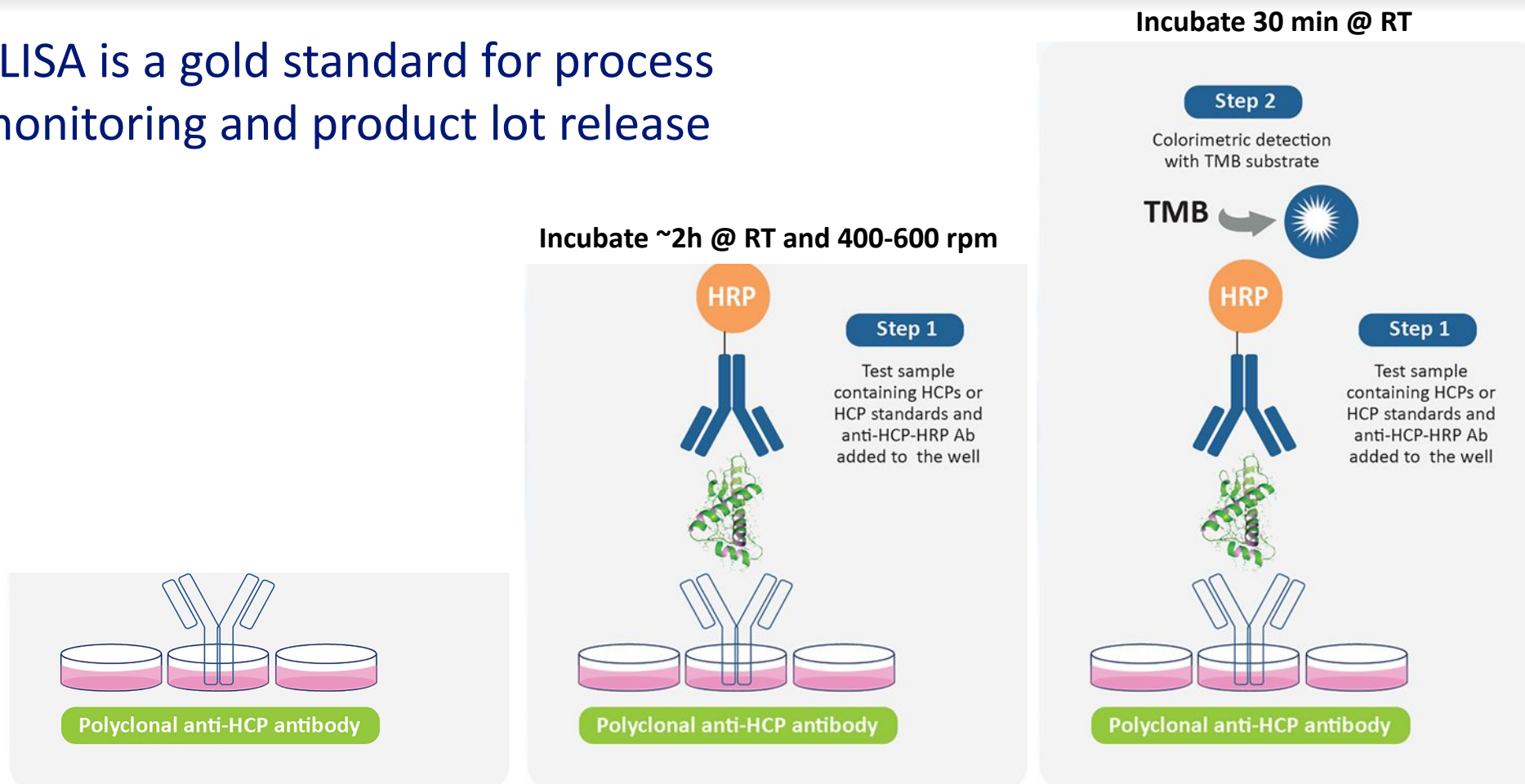
Lipoprotein lipase
Phospholipase A2 Group XV
Phospholipase A2 Group VII
Phospholipase A I

Jones, M. et al *Biotechnol. Bioeng.* 2021
Li, X. et al *bioRxiv.* 2020
Wang, F. et al *Bioprocess Intl.* 2018
Vanderlaan, M. et al *Biotechnol. Prog.* 2018
Durocher, V. et al. *J. of Biotech.* 2017
Valente, K. et al *Biotechnol. Prog.* 2015
Levy, N. et al. *Biotechnol Bioeng.* 2014



HCP Immunoassays

- ELISA is a gold standard for process monitoring and product lot release



Typical Cygnus HCP ELISA – Simultaneous Protocol

- ELISA is a gold standard for process monitoring and product lot release
 - Semi-quantitative measurement of a statistical sample of HCPs in a sample
 - Excellent sensitivity and selectivity
 - Does not require special expertise to run
 - Effective in relatively high levels of product protein
 - Easily transferable, reliable, robust

 - Reports total HCP equivalents (not ng/mL)
 - Results subject to change with change in kit/reagents
 - Can take 12-months to generate new reagents



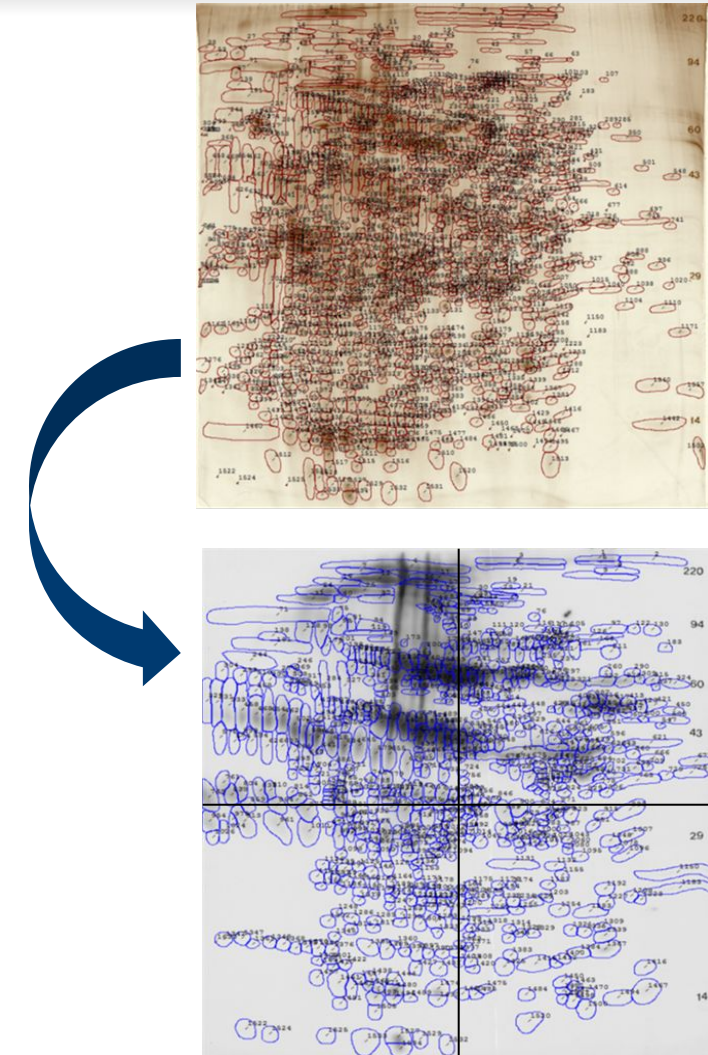
- Dilution Linearity
- Accuracy
- Precision
- Range: LOD/LLOQ/ULOQ
- Coverage by AAE™ /2D-PAGE
- Coverage & Characterization by AAE-MS™

- ICH Q11 “Development and Manufacture of Drug Substances (Chemical Entities Biotechnological/Biological Entities)”, 2012
- ICH Q6B “Test Procedures and Acceptance Criteria for Biotechnological/Biological Products,” 1999
- USP Chapter 1132 “Residual Host Cell Protein Measurement in Biopharmaceuticals,” 2016
- USP Chapter 1132.1 “Residual Host Cell Protein Measurement in Biopharmaceuticals by Mass Spectrometry”, 2023

Methods for Determining Antibody Coverage

ELISA Antibody Coverage Analysis

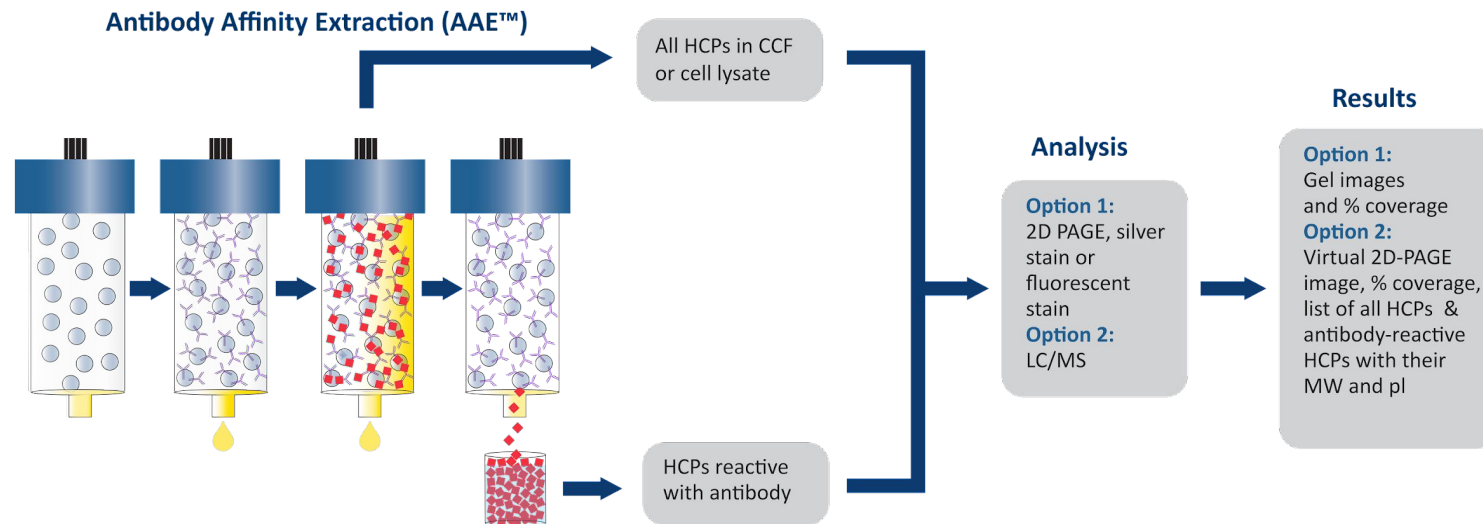
- Performed to demonstrate that the Ab used in the ELISA are broadly reactive to the HCPs in the expression platform
 - Conventional Method
 - 2D Western blot compared to Silver Stain
 - Specifications:
 - Reactive with the majority of spots found on the silver stain gel
 - Must have spots in each quadrant of the gel where spots are found on the silver stain
 - Antibody Affinity Extraction (AAE™) with Silver Stain or DIGE detection
 - AAE™ with LC-MS





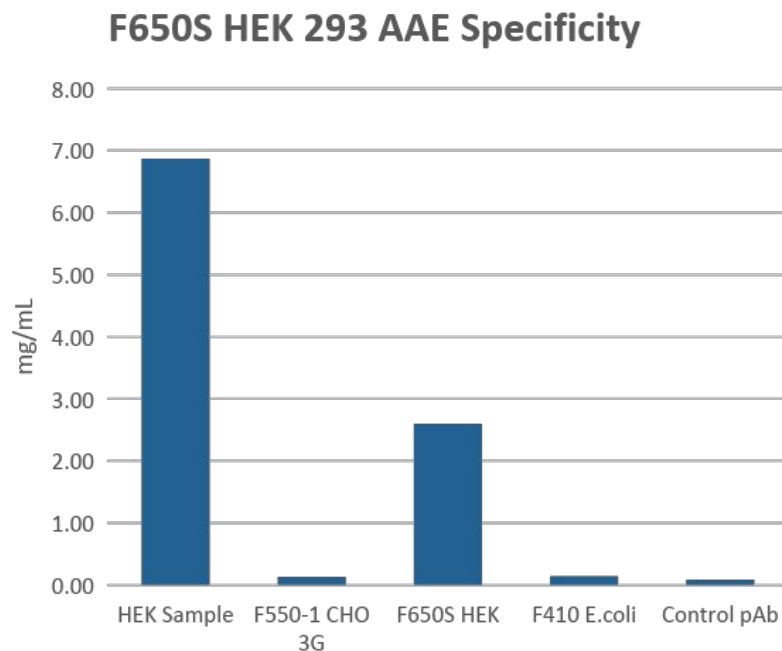
Antibody Affinity Extraction (AAE™)

Harvest Material

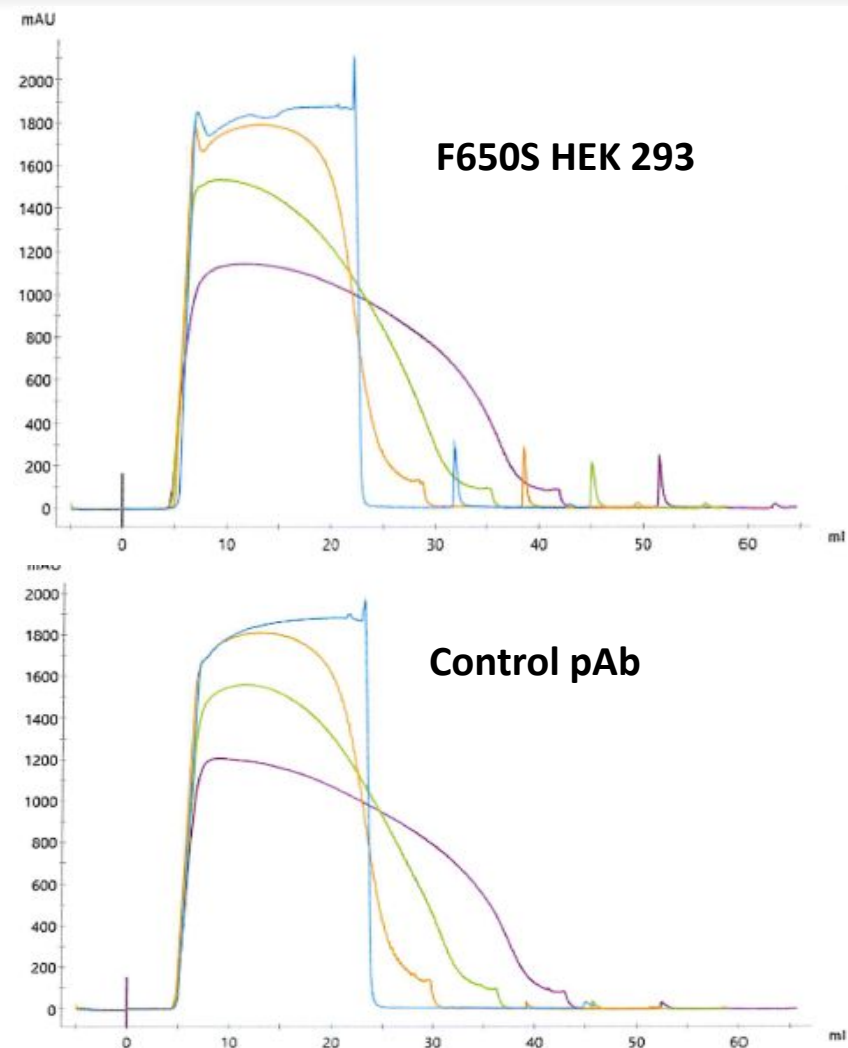


Method

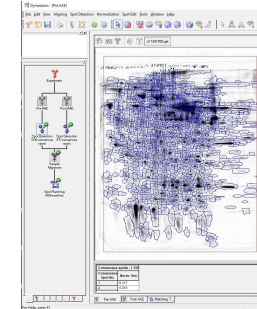
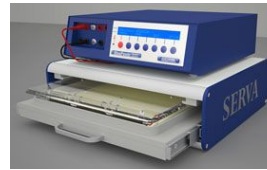
- AAE columns with antibodies from HEK 293 [F650S], CHO [F550-1], E. coli [F410] HCP ELISA Kits
- Negative controls: AAE columns with non-immune goat/rabbit IgG
- Antibody Affinity Extraction was performed with HEK 293 HCP
- LC-MS Sample Preparation:
 - the Pre- and Post-AAE HCPs were precipitated, dissolved in 8M urea, reduced, alkylated, digested with trypsin, desalted, and concentrated
- HCP Identification by LC-MS:
 - the Pre- and Post-AAE samples were analyzed with the custom LC-MS method independently in triplicate and in a randomized sequence. Blank washing runs were implemented in between sample injections to minimize sample carryover.
- HCP Quantification by LC-MS:
 - Cygnus Protein Standard (CPS) was spiked into all quantification samples prior to LC-MS sample preparation. Quantification of HCPs in ppm were calculated relative to CPS at 1000 ppm. The ng/mL of HCPs were calculated by multiplying the mg/mL of the sample to the ppm value.



HEK 293 HCP sample was passed over the F550-1 3G CHO, F650S, F410 E. coli, and IgG control columns by FPLC. Immunoreactive HCPs were quantified by LC-MS



2D-PAGE Workflow in HCP Analytics



Antibody Affinity Extraction & 2D Electrophoresis Analysis of HEK HCPs found in the Cygnus HEK Antigen using the Cygnus HEK Antibodies.

Work Performed By: Sue Babin, Luke Lockhart, and David Isaac Cygnus Technologies, LLC.

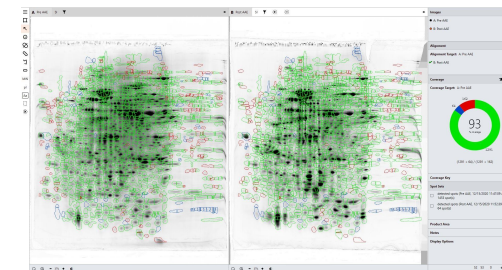
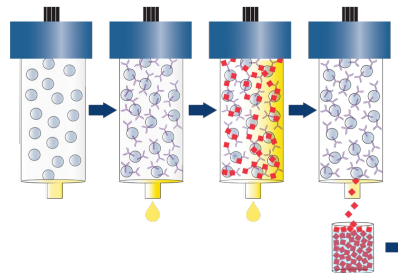
Reference:
Report dated 11 November 2019

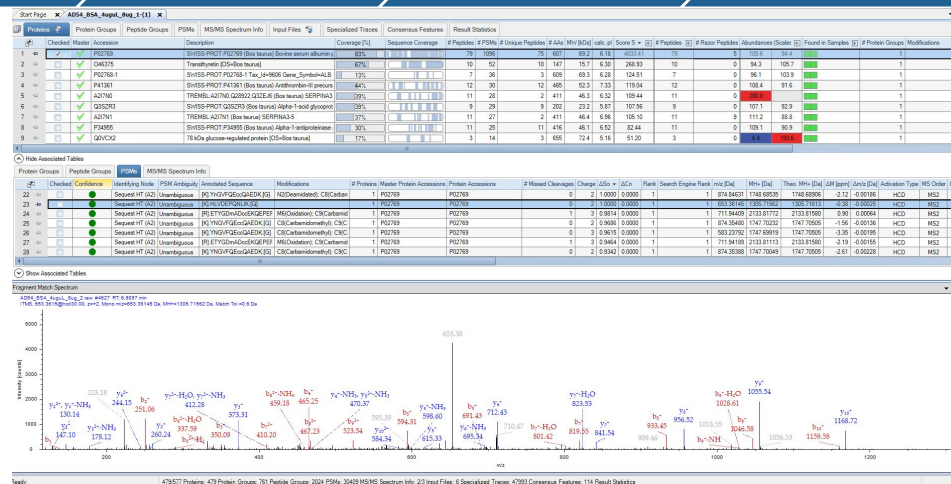
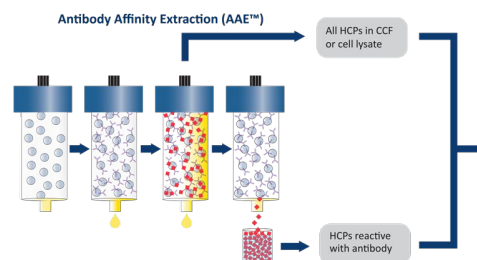
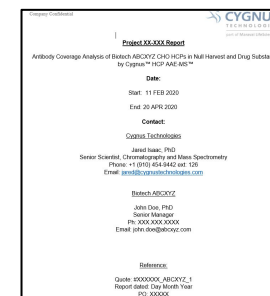
Summary:
The Cygnus HEK antigen was evaluated by Antibody Affinity Extraction (AAE). AAE was performed on the Cygnus HEK antigen using HEK antibodies. The protein in the AAE eluate was separated as described by gel and size and analyzed by silver stain. The gels were analyzed and the overall coverage was measured by the number of HCPs in the AAE eluate fraction after size exclusion as the 2D-PAGE silver stain of the starting antigen sample.

Materials and Methods:
AAE Column Preparation: The procedures used in column preparation and in the AAE protocol are proprietary to Cygnus Technologies. Standard regulatory agencies must use standard protocols. Cygnus can provide more information. The affinity purified antibody pool was used in the AAE. The HEK antigen was incubated in a 100% methanol/20% glycerol solution. The column was prepared in a manner way as described in the AAE protocol. The column was prepared in a manner way as described in the AAE protocol. The column was prepared in a manner way as described in the AAE protocol.

HCP Affinity Extraction: The sample was passed over the antibody affinity column to extract the HCPs. The column was immediately washed to remove all unbound sample and then eluted with acid. The eluted material was immediately neutralized to pH 7.0 using a base buffer system. Unbound HCPs were passed back over the column under the same conditions, eluted and combined with the first cycle. The sample was measured in this way 4 times. The eluted fractions were combined and then immediately lyophilized at 4°C for subsequent 2D-PAGE.

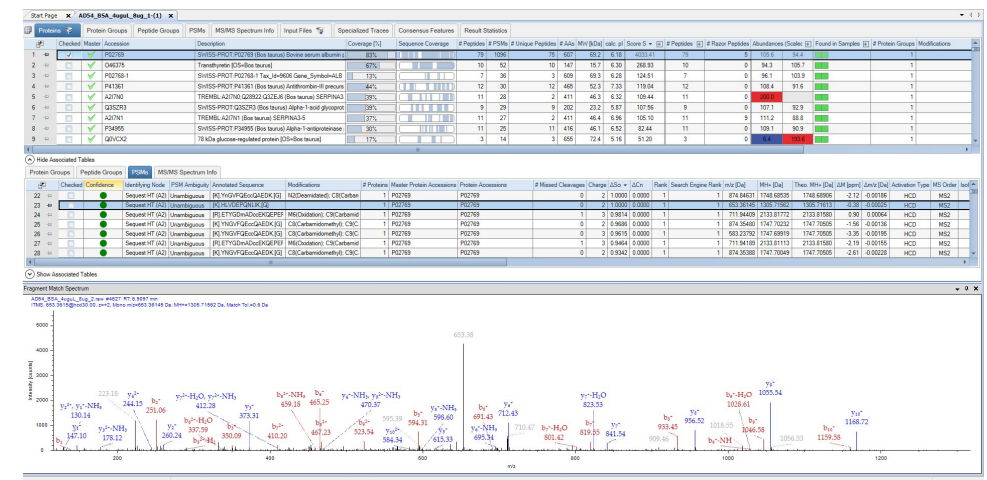
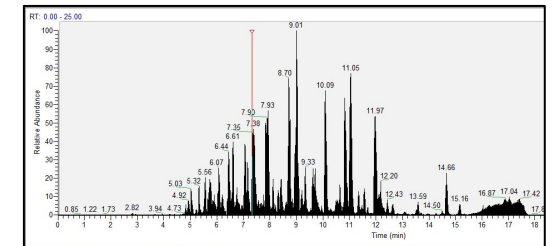
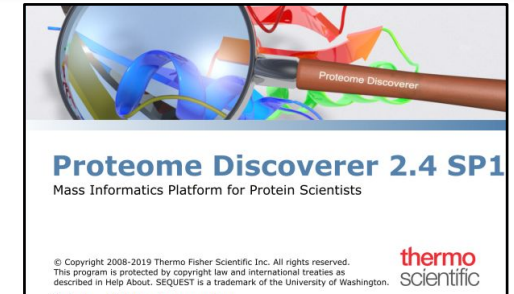
Sample Preparation:
100 µl of the sample was then diluted with 100 µl of SDS Buffering Buffer and diluted overnight at 4°C. The sample was then separated on 10% SDS-PAGE. The sample was then separated on 10% SDS-PAGE. The sample was then separated on 10% SDS-PAGE.



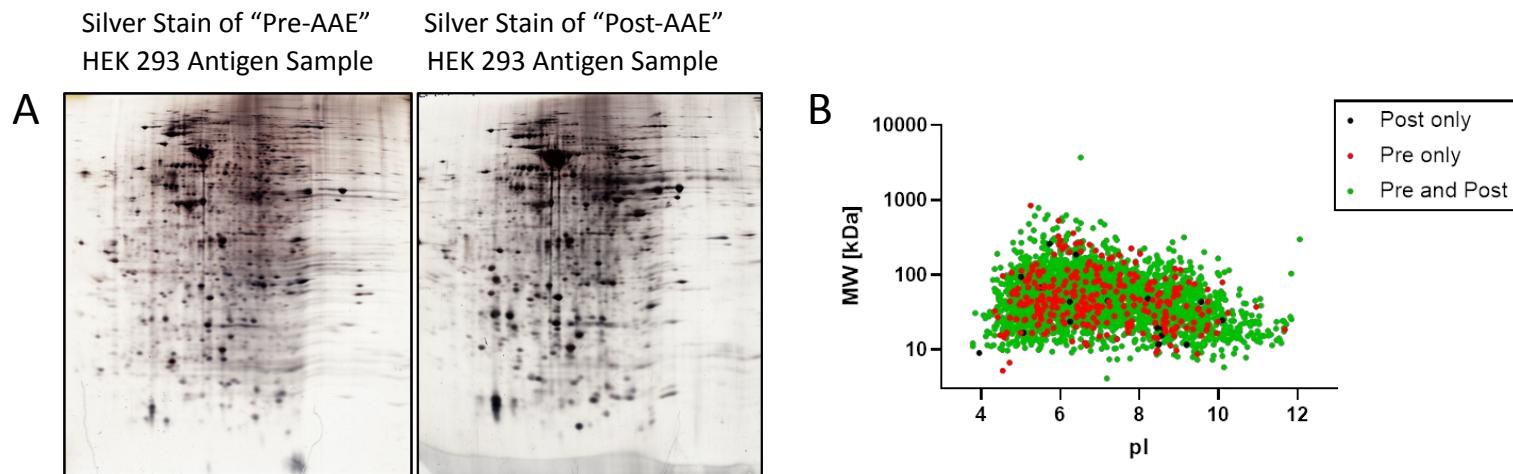


Curated Proteomic Databases

1. *Homo sapiens* (HEK293, HeLa)
2. *Cricetulus griseus* (CHO)
3. Vero
4. *Bos taurus*
5. *Mus musculus* (NS/O and PG13)
6. *E. coli*
7. Sf9
8. *S. cerevisiae*
9. *P. pastoris*
10. Goat
11. Rabbit
12. MRC5
13. *S. aureus*
14. *P. fluorescens*
15. *L. lactis*
16. Per.C6
17. BHK
18. MDCK

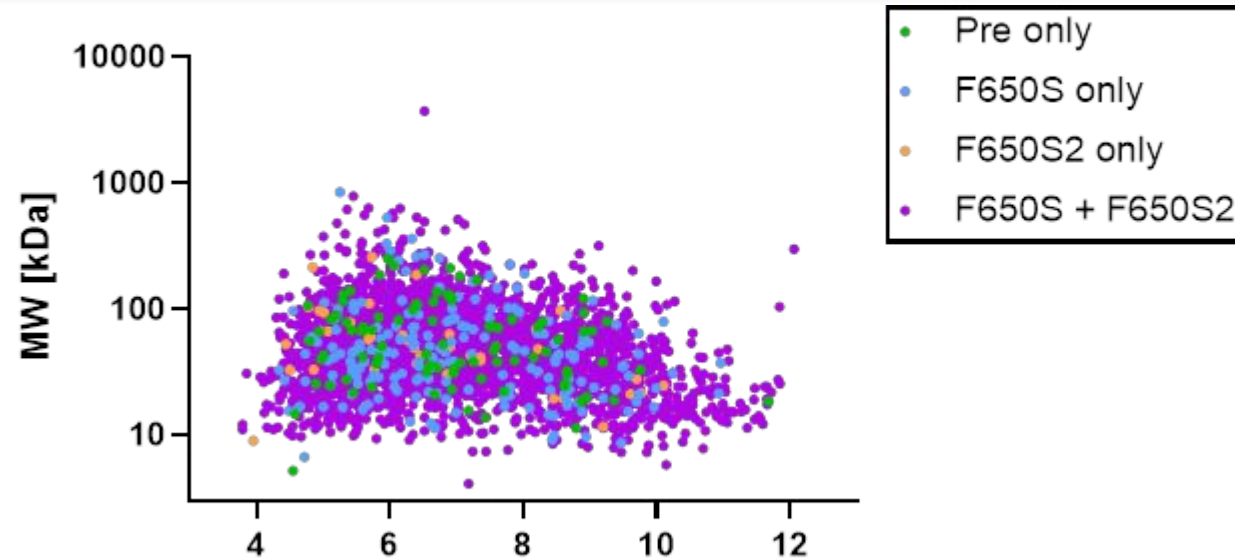


F650S Anti-HEK293 pAb Coverage of HEK 293 Harvest Sample



Detection Method	PRE-AAE (all HCPs)	POST-AAE (immunoreactive HCPs)	% pAb Coverage
2D-PAGE	1687	1604	95
MS	3075	2812	91

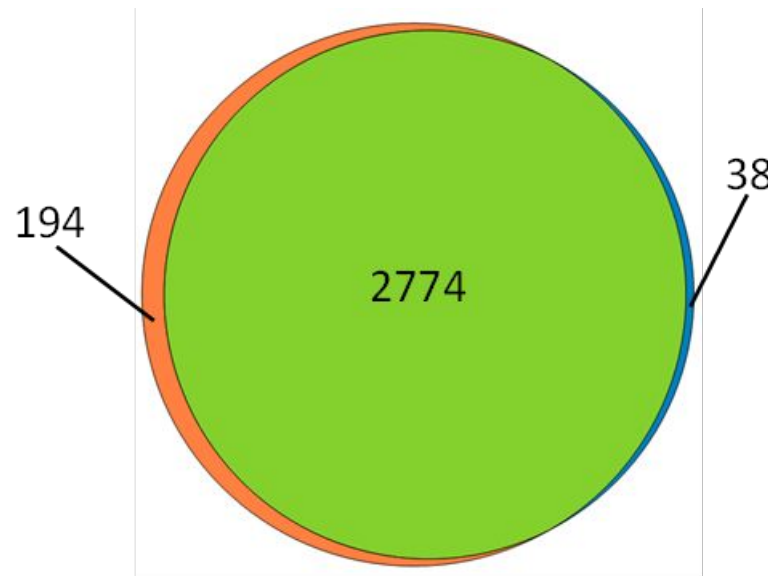
Virtual 2D Gels of Cygnus HEK 293 Master Antigen with F650S and F650S R1 HCP Antibodies: MW vs pI



Sample		AAE (number of protein ids)				% Antibody Coverage	
Name	AAE	Total	Unique to each fraction	Total Unique	Matching	Lower Boundary	Upper Boundary
F650S	Pre	3075	119	3278	2765	90.54%	96.52%
	Post	2968	12				
F650S Resupply	Pre	3075	281	3093	2794	90.91%	91.45%
	Post	2812	18				

Comparison of F650S and F650S Resupply Protein Identifications

Antibody	Total Protein ID	Matching Proteins	Unique Proteins	% Similarity
F650S	2968	2774	194	92.3%
F650S Resupply	2812		38	

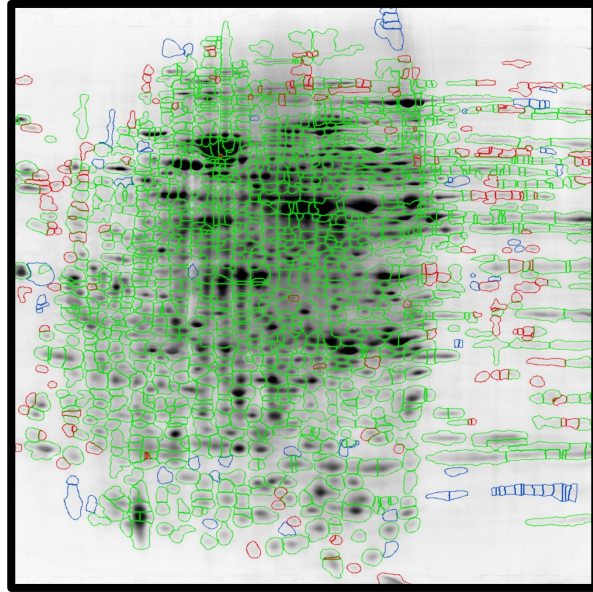


Quantitative Venn Diagram to assess the % similarity of F650S and F650S Resupply antibodies binding to the Cygnus HEK 293 antigen. Unique identifications of F650S Resupply (blue) and F650S (orange) are displayed along with proteins identified by both antibodies (green).

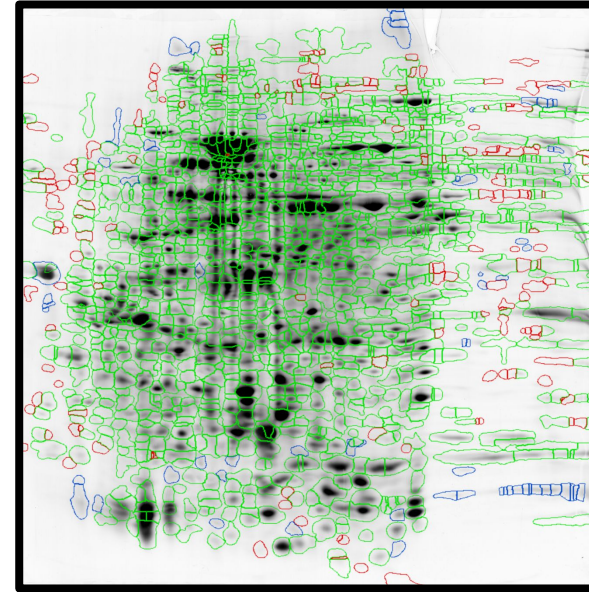
Individual HCP pAb Reactivity by MS

#	Potential High-Risk HCPs	PRE	F650S	F650S R	pI	MW
1	78 kDa glucose regulated protein (BiP)	Y	Y	Y	5.07	72332.96
2	Actin	Y	Y	Y	5.29	41736.73
3	Aldose reductase related protein 2 (Aldo-keto reductase)	Y	Y	Y	7.66	36019.6
4	Alpha enolase	Y	Y	Y	7.01	47169.0
5	Carboxypeptidase D	Y	Y	Y	5.68	152931.1
6	Cathepsin B	N	N	N	5.88	37821.6
7	Cathepsin D	Y	Y	Y	6.1	44552.2
8	Cathepsin E	N	N	N	4.69	42793.5
9	Chondroitin sulfate proteoglycan 4	N	N	N	5.27	250536.8
10	Clusterin	N	N	N	5.88	52494.6
11	Cofilin 1	Y	Y	Y	8.22	18502.5
12	Elongation factor 1a1	Y	Y	Y	9.1	50140.9
13	Elongation factor 2	Y	Y	Y	6.41	95338.1
14	Flagellin (Salmonella paratyphi A (strain ATCC 9150 / SARB42)	N	N	N	5.03	51950.2
15	Galectin 3 binding protein	Y	Y	Y	5.12	65331.0
16	Glutathione S transferase P	Y	Y	Y	5.43	23355.8
17	Glyceraldehyde 3 phosphate dehydrogenase	Y	Y	Y	8.57	36053.2
18	G-protein coupled receptor 56	N	N	N	8.79	57473.2
19	Heat shock cognate 71 kDa protein	Y	Y	Y	5.37	70898.1
20	Heat shock protein HSP 90	Y	Y	Y	4.94	84659.7
21	Lipoprotein Lipase	N	N	N	8.37	53162.5
22	Lysosomal protective protein	N	N	N	6.16	54466.1
23	Matrix Metalloproteinase 19	N	N	N	7.22	57357.0
24	Metalloproteinase inhibitor 1	N	N	N	8.46	23170.9
25	Monocyte chemoattractant protein 1 (C-C motif chemokine 2)	N	N	N	9.4	11025.0
26	Nidogen-1	N	N	N	5.12	136377.0
27	Peptidyl-prolyl cis-trans isomerase	Y	Y	Y	8.27	19257.2
28	Peroxiredoxin 1	Y	Y	Y	8.27	22110.4
29	Phosphoglycerate kinase 1	Y	Y	Y	8.3	44614.7
30	Phospholipase A2 (Group XV lysosomal)	Y	Y	Y	6.26	46657.8
31	Phospholipase B like 2	Y	Y	Y	6.34	65471.8
32	Procollagen C endopeptidase enhancer 1	N	N	N	7.41	47972.5
33	Procollagen-lysine, 2-oxoglutarate 5-dioxygenase 1	Y	Y	Y	6.46	83550.19
34	Procollagen-lysine (Procollagen-lysine 5-dioxygenase)	Y	Y	Y	6.54	83504.2
35	Protein disulfide isomerase	Y	Y	Y	4.76	57116.37
36	Pyruvate kinase	Y	Y	Y	7.96	57936.9
37	Serine protease HTRA1	N	N	N	8.09	51287.0
38	SPARC (Osteonectin)	N	N	N	4.73	34632.2
39	Sulfated glycoprotein 1 (Prosaposin)	Y	Y	Y	5.06	58112.6
40	T-complex protein 1 subunit alpha	Y	Y	Y	5.8	60343.6
41	Thioredoxin 1	Y	Y	Y	4.82	11737.5

PRE AAE

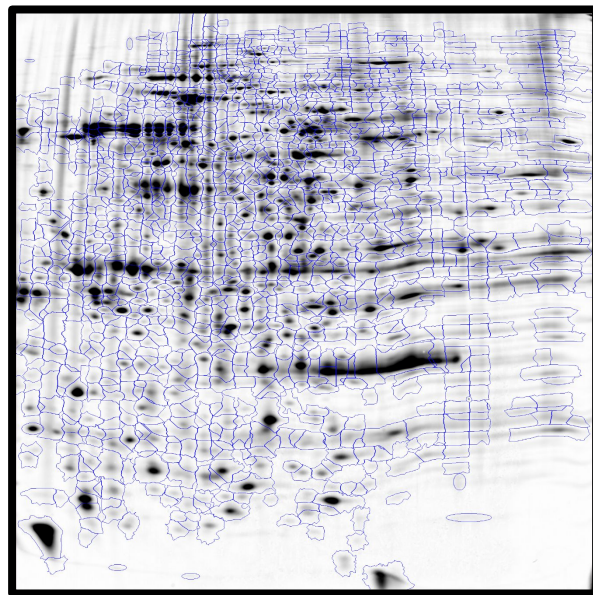


POST AAE

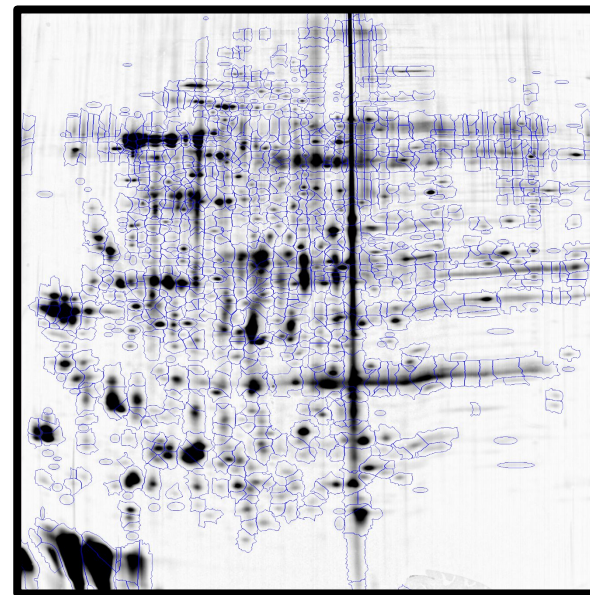


There were 1454 spots in the sample and 1356 spots in the AAE elution fraction. **Based on our assessment the coverage of the antibodies is 92%.**

PRE AAE

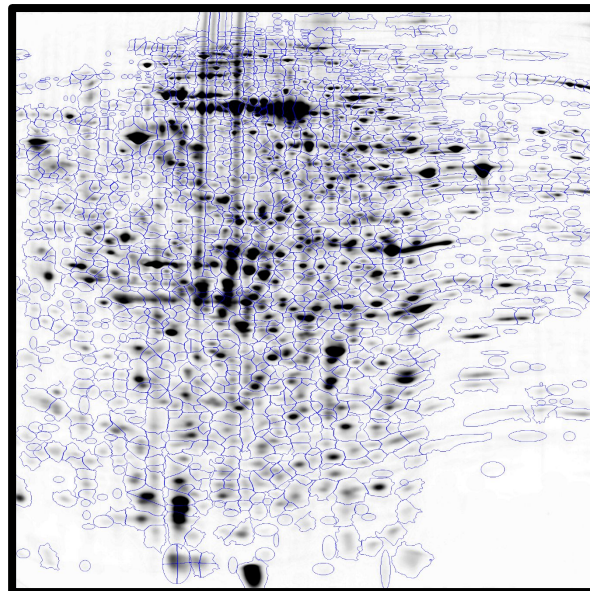


POST AAE

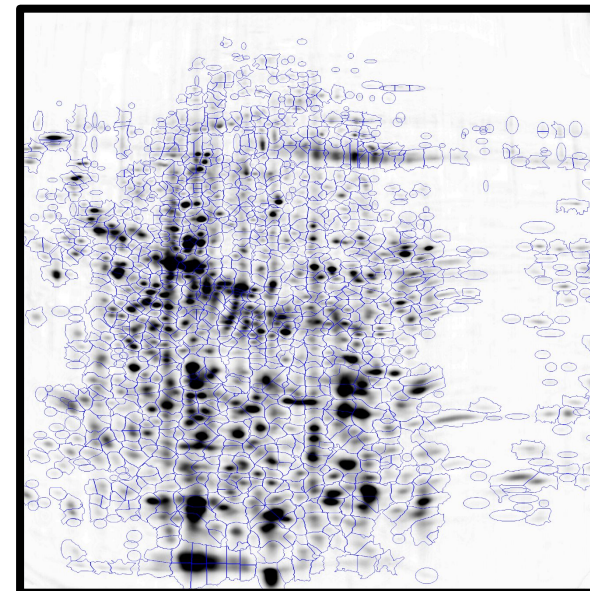


There were 1585 spots in the sample and 1214 spots in the AAE elution fraction. **Based on our assessment the coverage of the antibodies is between 64% - 78%.**

PRE AAE

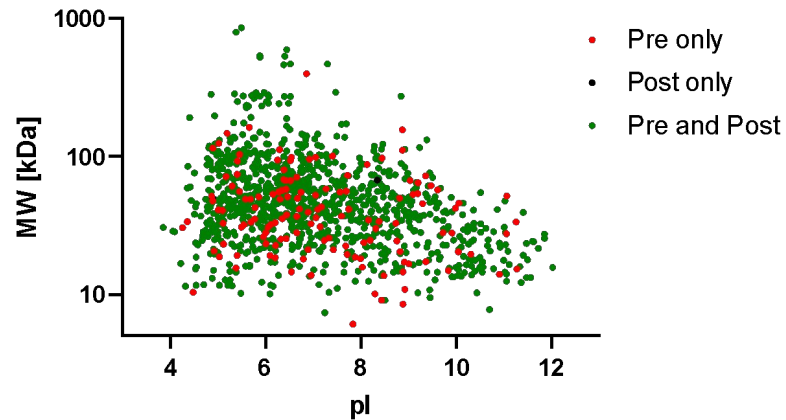


POST AAE



There were 1517 spots in the sample and 1193 spots in the AAE elution fraction. Based on our assessment the coverage of the antibodies is between 62% - 79%.

Vero F975 AAE-MS



	AAE		
	PRE	POST	% Antibody Coverage
no of HCPs	1129	993	88%

#	Potential High-Risk HCPs	Pre AAE	Post AAE	IEP	MW
1	Protein 1	N	N	5.07	72379.1
2	Protein 2	Y	Y	5.29	41736.7
3	Protein 3	Y	Y	5.85	46698.3
4	Protein 4	Y	Y	5.98	50011.7
5	Protein 5	N	N	5.3	48265.1
6	Protein 6	Y	Y	5.73	35646.9
7	Protein 7	Y	Y	6.54	44110.9
8	Protein 8	N	N	4.61	42726.4
9	Protein 9	N	N	5.4	252012
10	Protein 10	N	N	5.58	51557.5
11	Protein 11	Y	Y	8.22	18532.5
12	Protein 12	Y	Y	9.39	55106
13	Protein 13	Y	Y	6.41	95324.1
14	Protein 14	N	N	4.5	51295
15	Protein 15	N	N	5.05	63802.2 2
16	Protein 16	Y	Y	7.64	23638.2
17	Protein 17	Y	Y	8.49	35747.9
18	Protein 18	N	N	9.06	77370.5
19	Protein 19	Y	Y	5.23	70804.9
20	Protein 20	Y	Y	4.94	83166.1
21	Protein 21	Y	Y	7.94	52900.2 9
22	Protein 22	N	N	5.64	56110.7
23	Protein 23	N	N	7.71	58942
24	Protein 24	N	N	8.84	22401
25	Protein 25	N	N	9.32	15858.4
26	Protein 26	N	N	4.72	83103
27	Protein 27	Y	Y	9.59	23634.4
28	Protein 28	Y	Y	8.22	22262.6
29	Protein 29	Y	Y	8.02	44562.5
30	Protein 30	N	N	6.16	87100
31	Protein 31	Y	Y	5.63	61824.4
32	Protein 32	N	N	8.16	50446.5
33	Protein 33	Y	Y	6.46	83550.2
34	Protein 34	N	N	6.57	83327.9
35	Protein 35	Y	Y	5.98	56796.4
36	Protein 36	Y	Y	6.88	57893.8
37	Protein 37	Y	Y	6.62	34404.5
38	Protein 38	Y	Y	7.1	51081.6
39	Protein 39	N	N	5.31	65758.2
40	Protein 40	Y	Y	5.7	60338.6

Coverage is Dependent on Method



2D WB = 50-60%

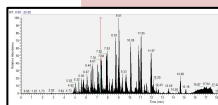


AAE-2D PAGE

Silver = 60-80%

Fluorescent = 70-80%

Cy3 and Cy5 = 70-85%



AAE-MS = 70-90%



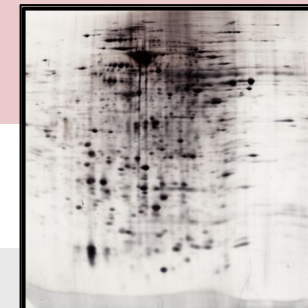
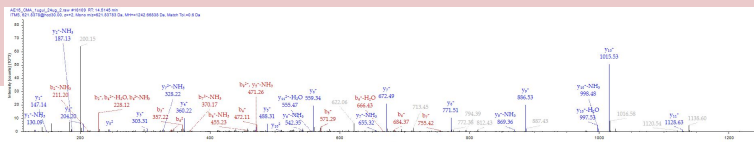
How is ELISA pAb Antibody Coverage Determined?

Lower boundary = (Post AAE spots/Unique spots)

Upper boundary = (Post AAE spots/Pre AAE-spots)

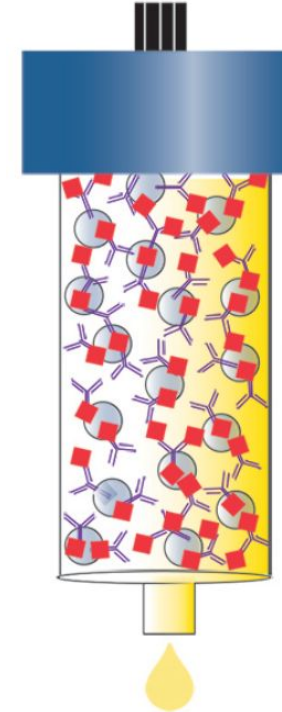


Spots and Ions



AAE in Completing the HCP Picture

- Antibody coverage analysis
 - 2D WB = 50%
 - AAE and 2D SDS PAGE= 65-75% (Silver Stain)
 - AAE and MS > 80-85%
- Coverage is dependent upon detection method
- AAE is more predictive of how the Ab will perform in ELISA
- Increased sensitivity and specificity as compared to 2D WB
- Sufficient sensitivity to evaluate individual HCPs that persist through purification process

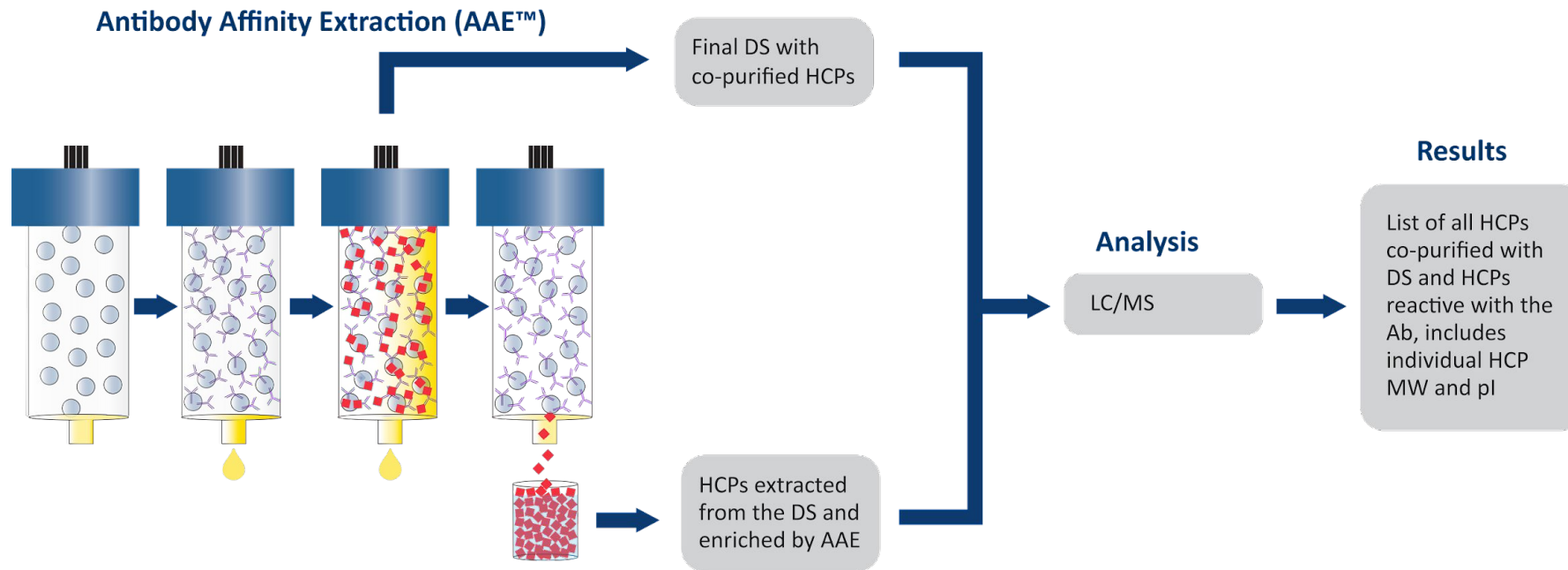




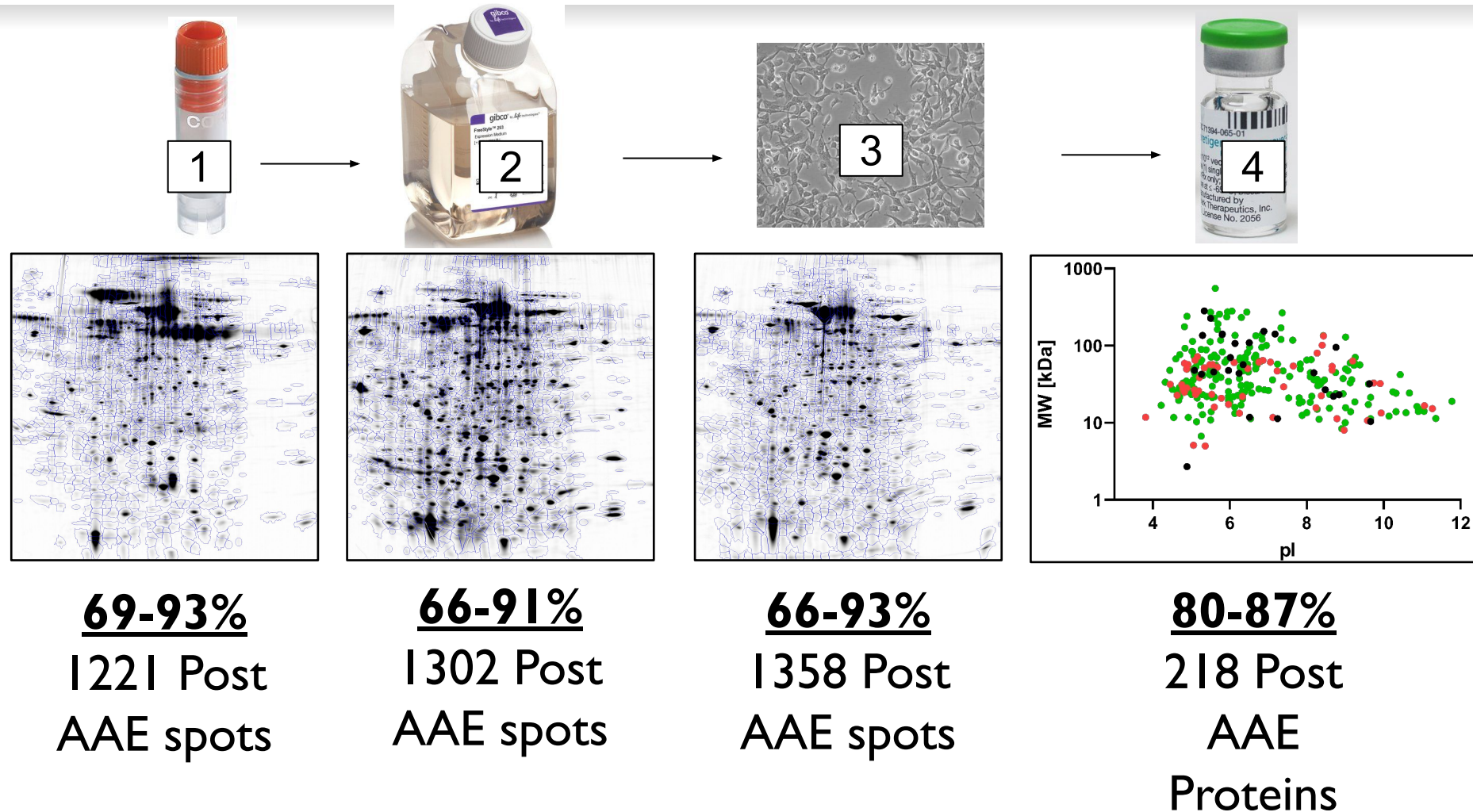
HCP Identification in DS and IP Samples by AAE-MS™

HCP Identification in Drug Substances

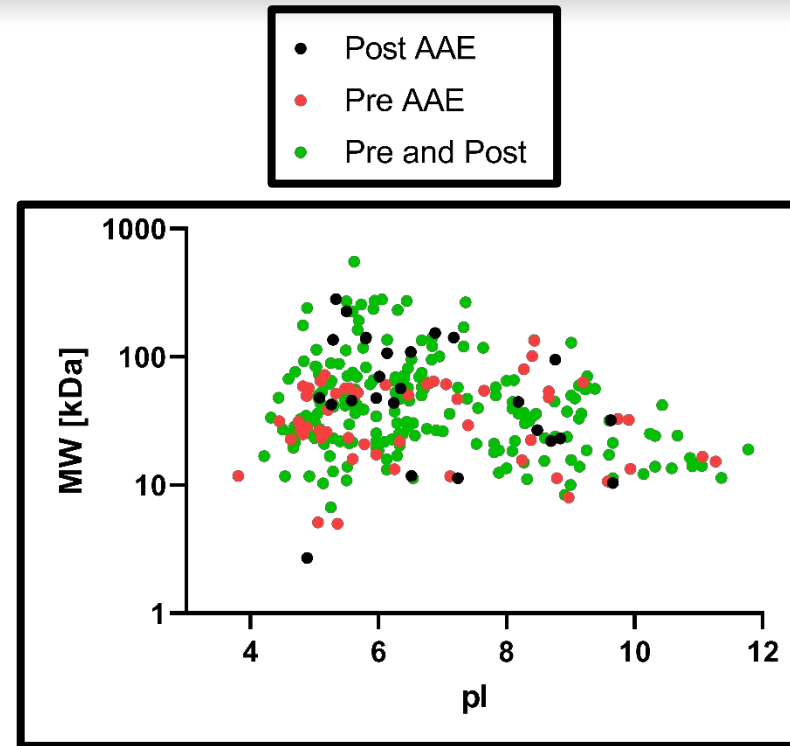
DS Material



Gene Therapy HEK 293 F650S Coverage by AAE 2D-PAGE and MS



MS is well suited for antibody coverage of DS



F650S: 87%

- 251 Pre AAE spots
- 218 Post AAE spots

HCP Antibody Coverage Analysis and Assay Qualification

- Comprehensive qualification of in-house assays or generic kits by orthogonal methods (**AAE, Mass Spec**)
- AAE method to demonstrate coverage to those individual HCPs that co-purify with drug substance & identify those HCPs by MS
- ELISA testing including dilution linearity and spike recovery analysis

Process-specific antibody and assay development:

- Proven antibody generation & purification
- Cygnus develops, qualify, & manufacture a robust kit that meets customer specific monitoring, release testing & regulatory needs

techsupport@cygnustechnologies.com

Acknowledgments



- Eric Bishop
- Jared Isaac
- Bryan Lanning
- Jacob Stubbs
- Stephen Stahlschmidt
- Johanna Barnhill



Thank You

Driving your success at every stage



CYGNUS
TECHNOLOGIES

part of Maravai LifeSciences