



Relative Infectivity as a Reliable Alternative to the TCID₅₀ Assay

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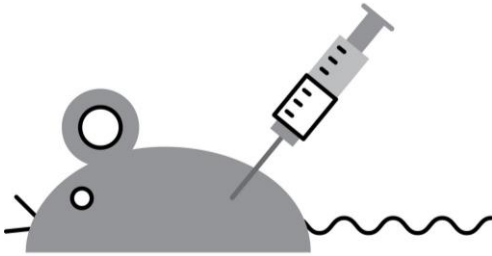
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AAV gene therapies for rare diseases

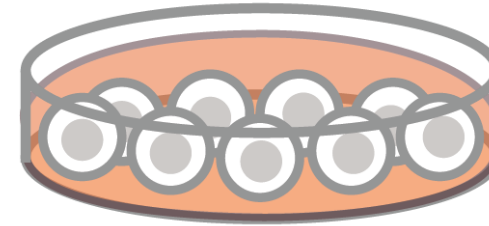
- The recent approvals of Luxturna™ and Zolgensma® have provided validation for the use of AAV as a gene delivery platform, especially in the treatment of rare genetic diseases
- For rare diseases, early clinical success may lead to expedited product development and approvals
- **Product approval requires a potency assay that:**
 - Measures the biological activity or activities specific to the product
 - Is validated to be accurate, sensitive, specific, and reproducible
 - Includes appropriate reference materials, standards, and/or controls
 - Provides quantitative data that can indicate stability, in order to establish dating periods
- Development of a potency assay is also critical for supporting product comparability and facilitating product development

Comparison of *in vivo* vs. *in vitro* approaches for AAV potency assay development



In Vivo

- ↔ Can be designed to represent most aspects of the product's mechanism of action, depending on the selected animal model
- ↓ **High biological variability**
 - Limited quantitative ability
 - Limited sensitivity for detecting changes upon stability
- ↓ **Long time to results**
 - Results can take months

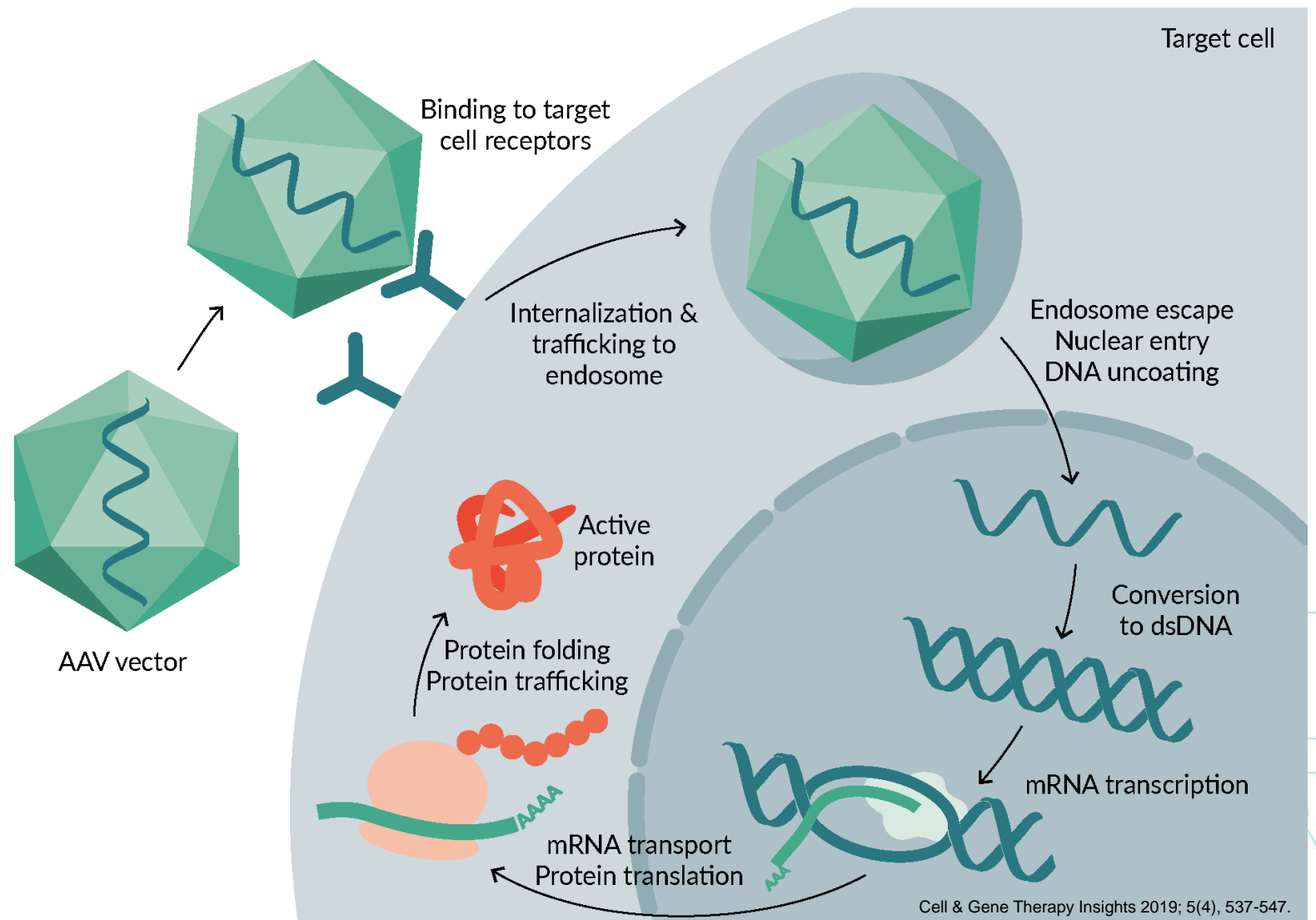


In Vitro

- ↔ Can be designed to represent many aspects of the product's mechanism of action, including transfer and biological effect of the target transgene
- ↑ **Lower biological variability**
 - Capable of providing quantitative results, relative to well-characterized reference standard
 - More sensitive to changes upon stability
- ↑ **Shorter time to results**
 - Results may be turned around in a week

Multiple coordinated steps are required for AAV transduction

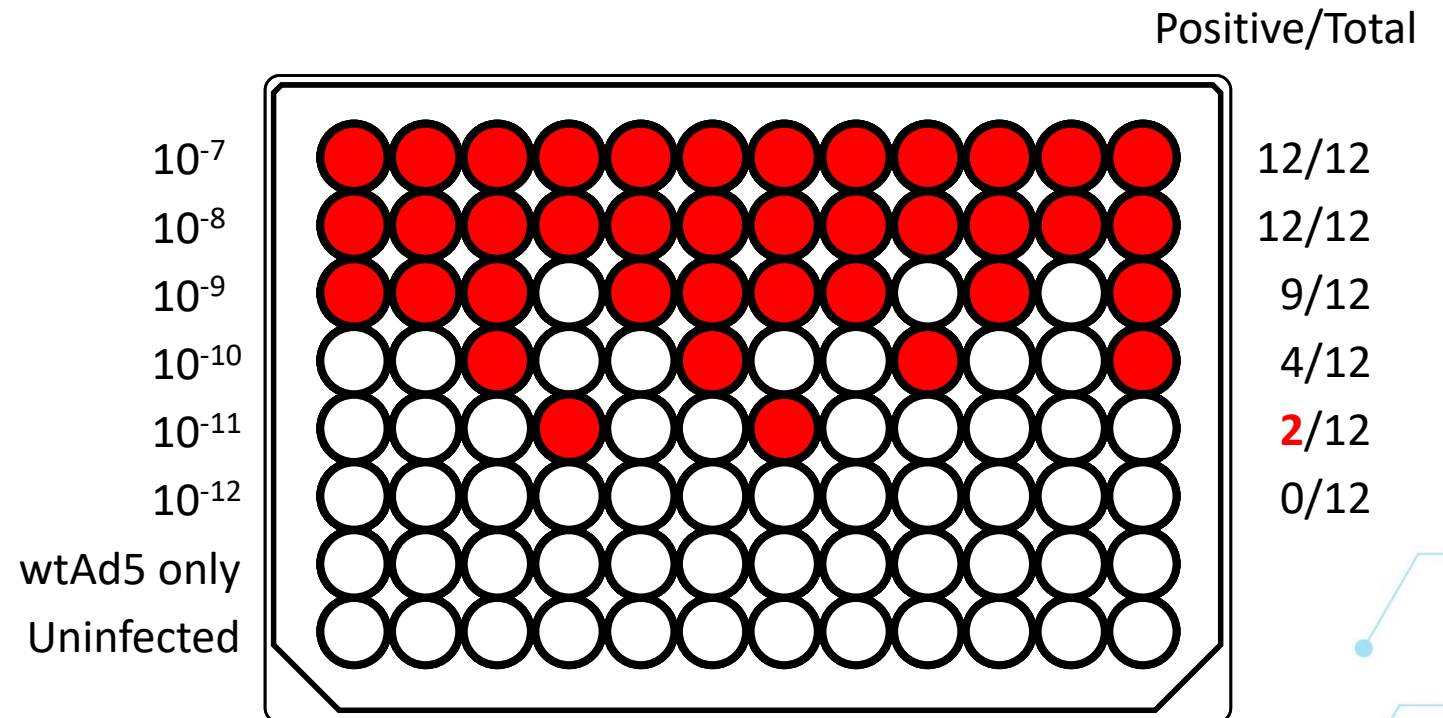
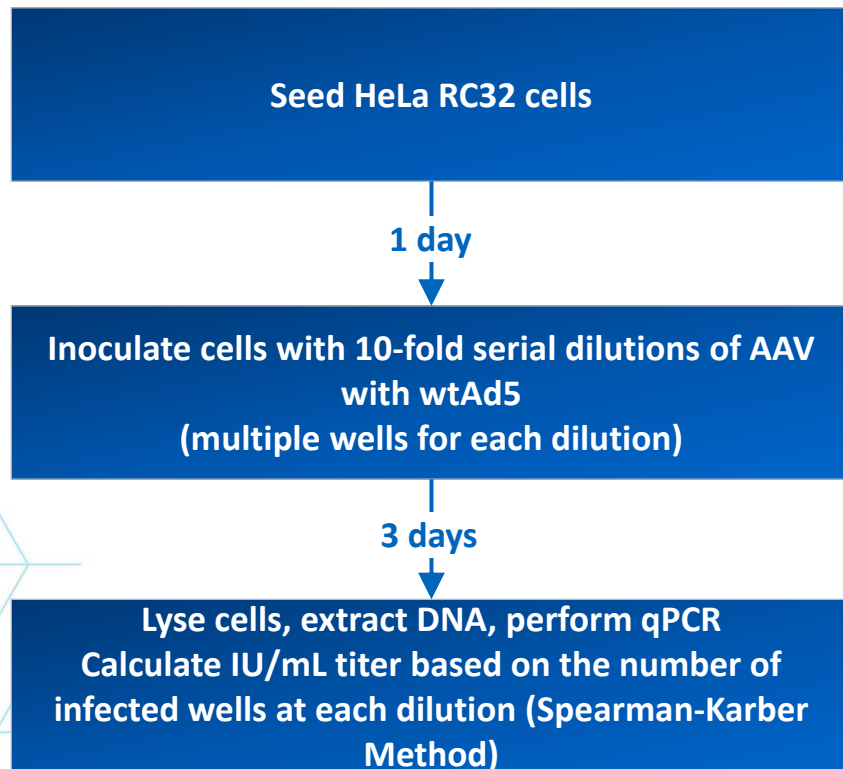
- **Problem:** *In vitro* methods that measure delivery of the transgene as well as the biological effect of the expressed sequence can take extensive time and effort to develop, which can slow the pace of product development
- **Solution:** Use a quantitative platform *in vitro* method that measures transgene delivery (infectivity) during early product development, along with less quantitative methods that confirm biological activity
 - Allow time for development of the *in vitro* potency method



Measuring infectivity for rAAV

- **Virus infectivity:** The capacity of viruses to enter the host cell and exploit its resources to replicate and produce progeny infectious viral particles
- **Traditional virological methods were developed 60-100 years ago to quantify infectious virus particles using cells permissive to infection *in vitro***
 - Infectious center assays (i.e., plaque, focus forming assays)
 - Endpoint dilution assays (i.e., median tissue culture infectious dose or TCID₅₀, most probable number or MPN)
- **However:**
 - wtAAV requires a helper virus for replication
 - rAAV gene therapy products lack the *rep* and *cap* genes and are incapable of replication, even in the presence of helper virus
 - Different AAV serotypes display differences in tissue and cell tropism
- **The TCID₅₀ assay was modified for AAV in order to accommodate the traditional definition of virus infectivity:**
 - HeLa RC32 cells stably expressing AAV2 *rep* and *cap* genes
 - Co-infection with wild-type Adenovirus 5 helper virus
 - Instead of cytopathic effect (CPE), wells are scored as infected or uninfected based on the measurement of vector genome replication

The TCID₅₀ infectious titer method



$10.75 \log_{10} \text{IU/mL}$

$5.62 \times 10^{10} \text{ IU/mL}$

(20% Difference)

The TCID₅₀ infectious titer method has very high assay variability

- ~200% geometric coefficient of variation (CV) is typical for TCID₅₀
- TCID₅₀ is an unreliable tool for measuring differences in infectivity across different vector preparations or changes as a result of degradation

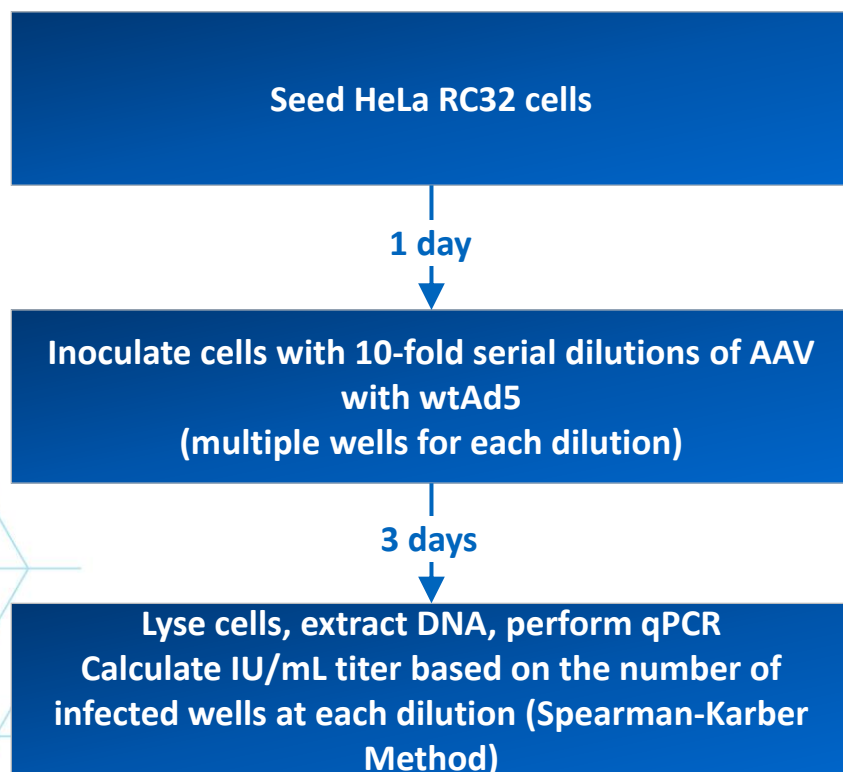


TABLE 5. FINAL rAAV2 REFERENCE STANDARD MATERIAL TITER ESTIMATES AFTER TRANSFORMATION AND MODELING

Titer units (method)	Transformation ^a	Mean	Lower 95% confidence limit for the mean	Upper 95% confidence limit for the mean	± 2 SD	± 3 SD
Particles/ml (ELISA)	Untransformed	9.18×10^{11}	7.89×10^{11}	1.05×10^{12}	3.73×10^{11} – 1.45×10^{12}	1.04×10^{11} – 1.78×10^{12}
Vector genomes/ml (qPCR)	Square root	3.28×10^{10}	2.70×10^{10}	4.75×10^{10}	9.00×10^8 – 1.04×10^{11}	0 – 1.66×10^{11}
Transducing units/ml (green cells)	Square root	5.09×10^8	2.00×10^8	9.60×10^8	0 – 2.47×10^9	0 – 4.00×10^9
Infectious units/ml (TCID ₅₀)	Log ₁₀	4.37×10^9	2.06×10^9	9.26×10^9	5.15×10^8 – 3.71×10^{10}	1.77×10^8 – 1.08×10^{11}

SD = 0.46 log₁₀ IU/mL → 191% Geometric CV

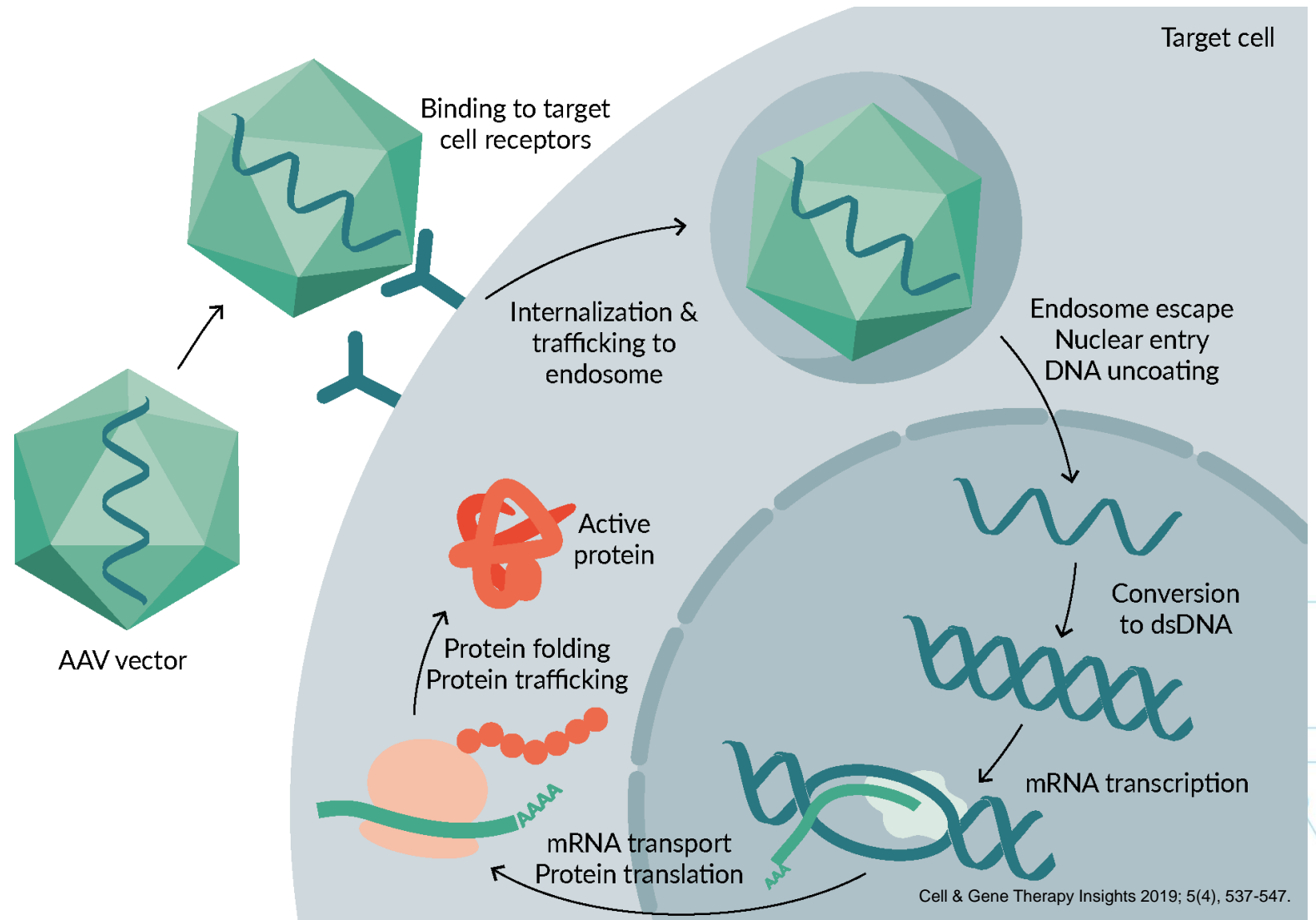
TABLE 4. FINAL rAAV8 REFERENCE STANDARD MATERIAL TITER ESTIMATES AFTER TRANSFORMATION AND MODELING

Titer units	Transformation	Mean	Lower 95% confidence limit for the mean	Upper 95% confidence limit for the mean	± 2 SD
Particles (pt)/ml	None	5.5×10^{11}	4.26×10^{11}	6.75×10^{11}	1.06×10^{11} to 9.94×10^{11}
Vector genomes (vg)/ml	Log ₁₀	5.75×10^{11}	3.05×10^{11}	1.09×10^{12}	4.57×10^{10} to 7.24×10^{12}
Infectious units (IU)/ml	Log ₁₀	1.26×10^9	6.46×10^8	2.51×10^9	1.32×10^8 to 1.20×10^{10}

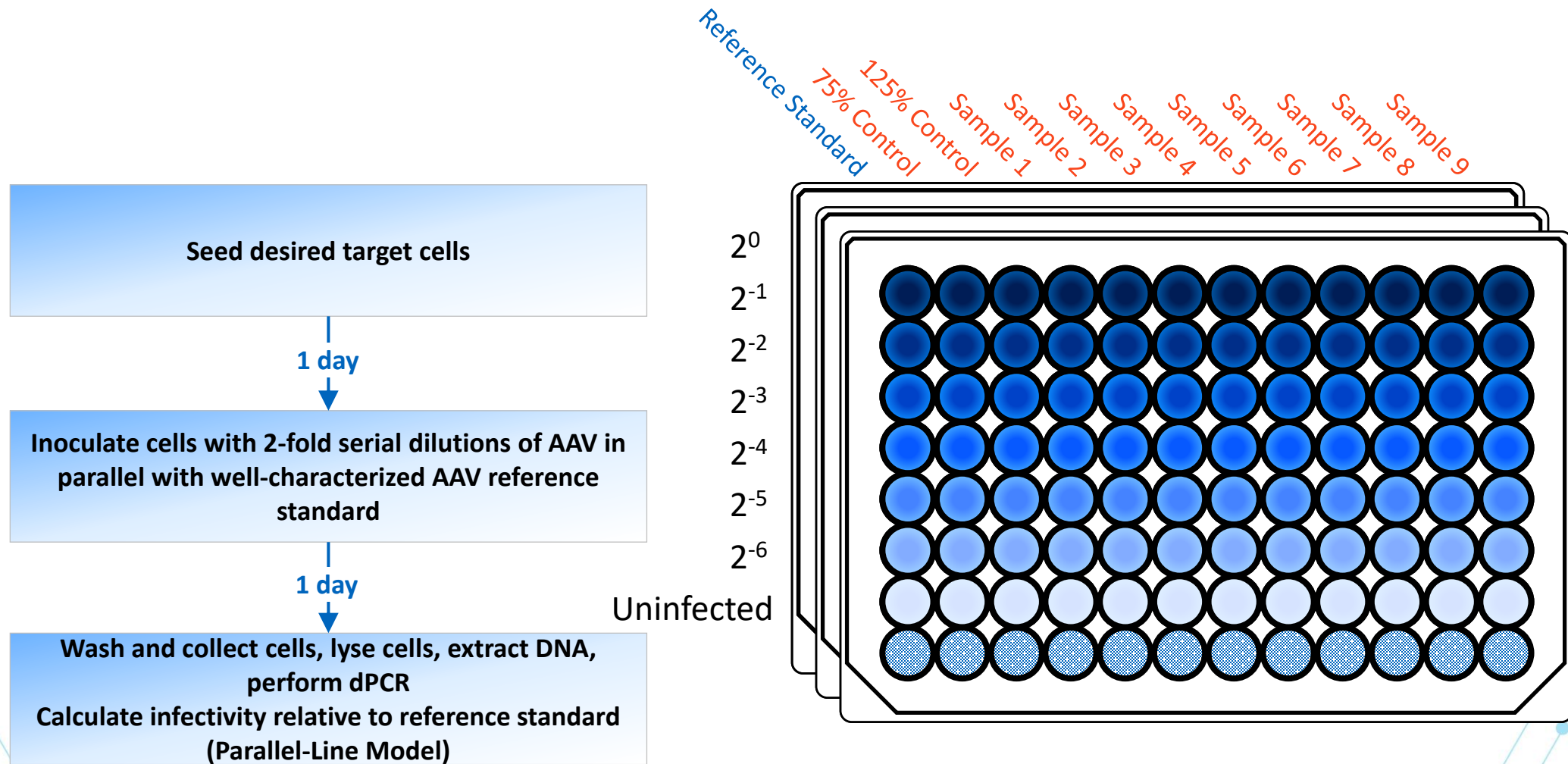
SD = 0.49 log₁₀ IU/mL → 209% Geometric CV

How can you measure rAAV infectivity with better precision?

- Since rAAV cannot replicate, rAAV infectivity should be defined as the capacity of rAAV to enter the target cell and deliver its genome
- Measure delivery of the AAV vector genome to target cells relative to a reference standard

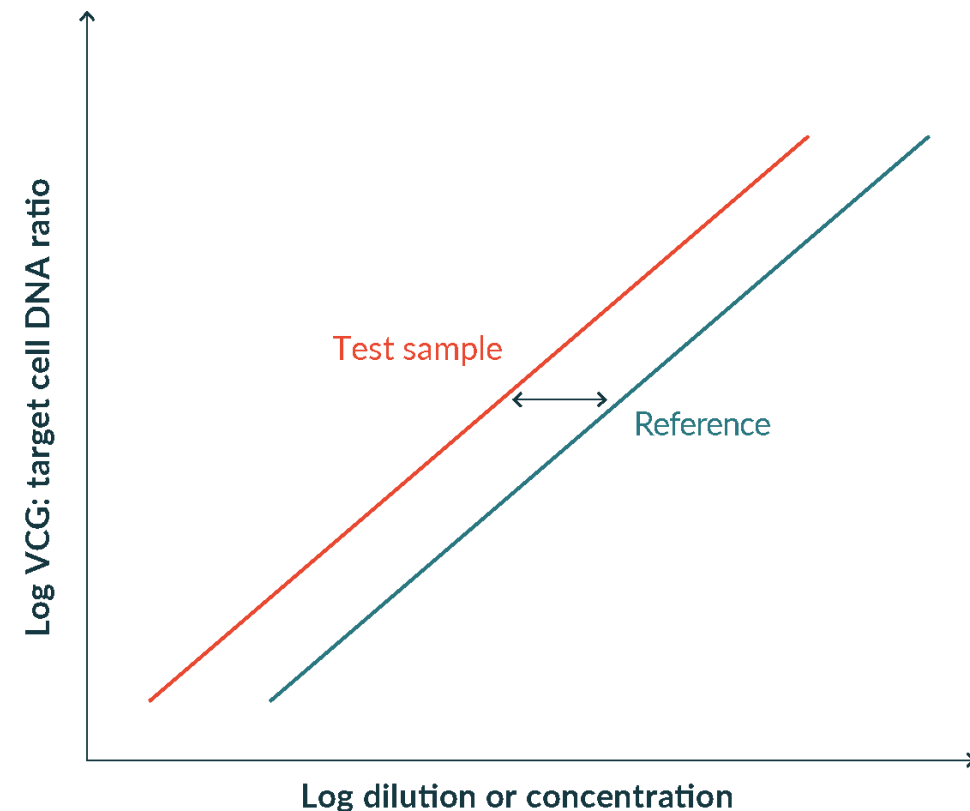
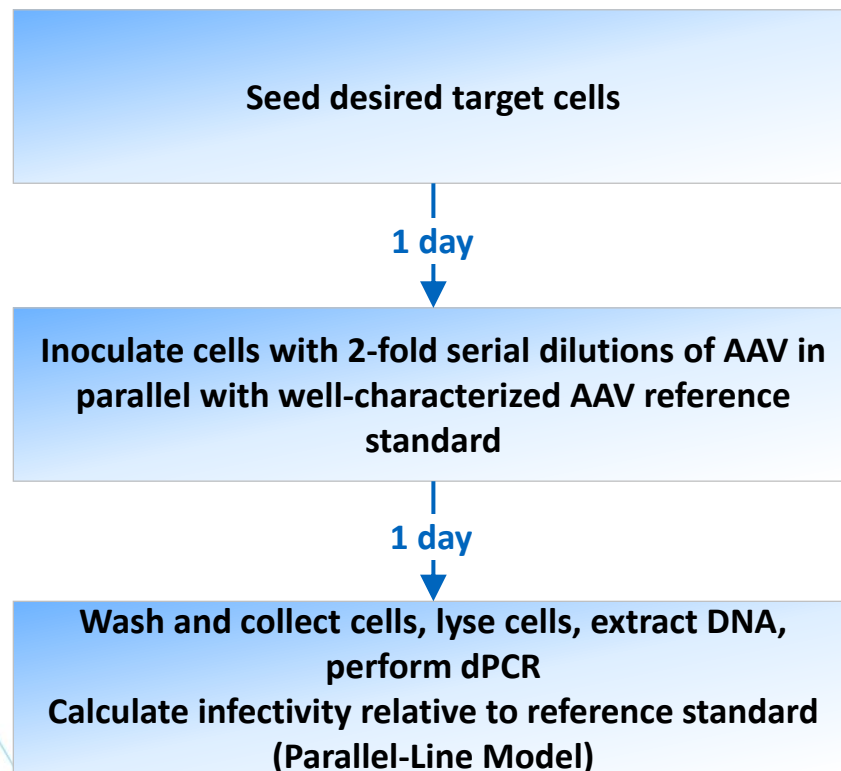


The relative infectivity method as a reliable alternative to the TCID₅₀ method



The relative infectivity method as a reliable alternative to the TCID₅₀ method

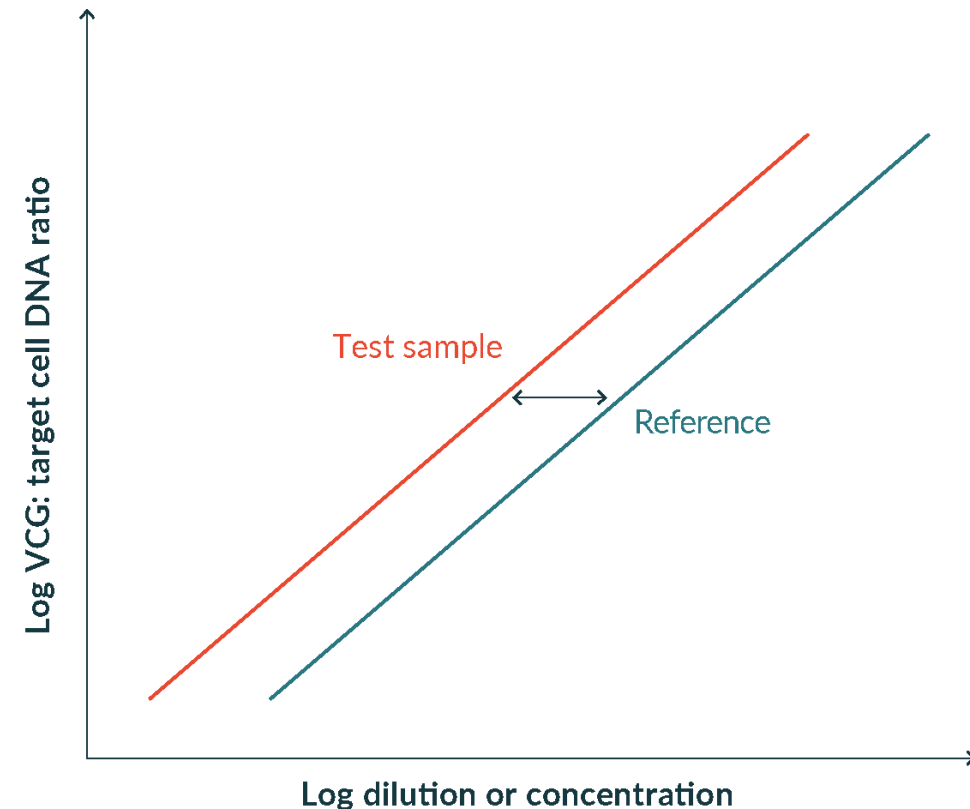
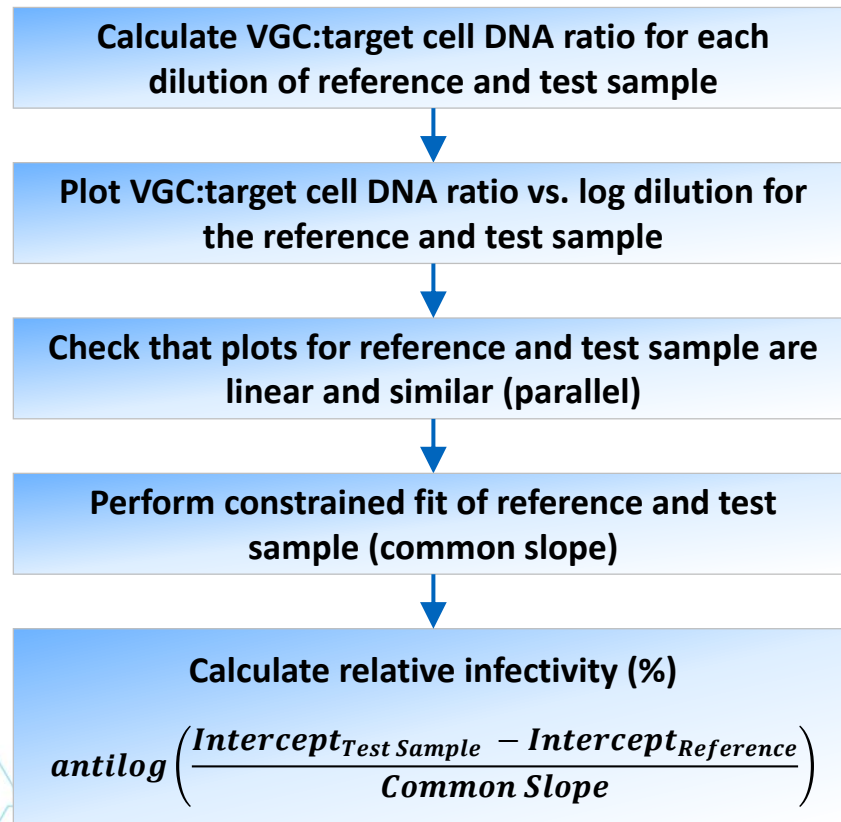
- Faster time to result than TCID₅₀ (2 days vs 4 days)
- Requires a well-characterized AAV reference standard that is known to be infectious
- Does not require co-infection with wild-type Adenovirus



VGC: Vector genome concentration
Source: Cell & Gene Therapy Insights 2019; 5(4), 537-547.

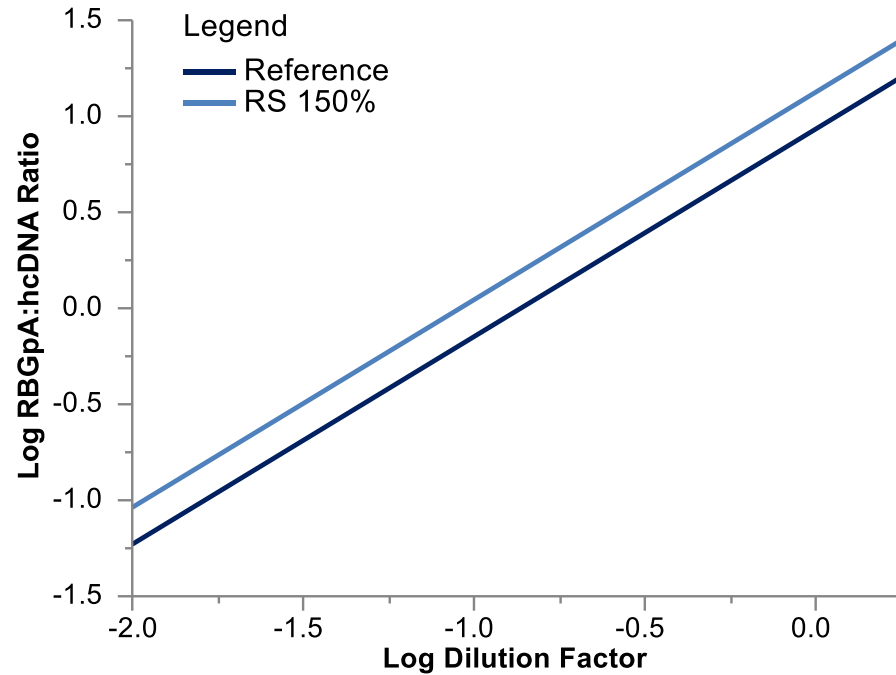
The relative infectivity method as a reliable alternative to the TCID₅₀ method

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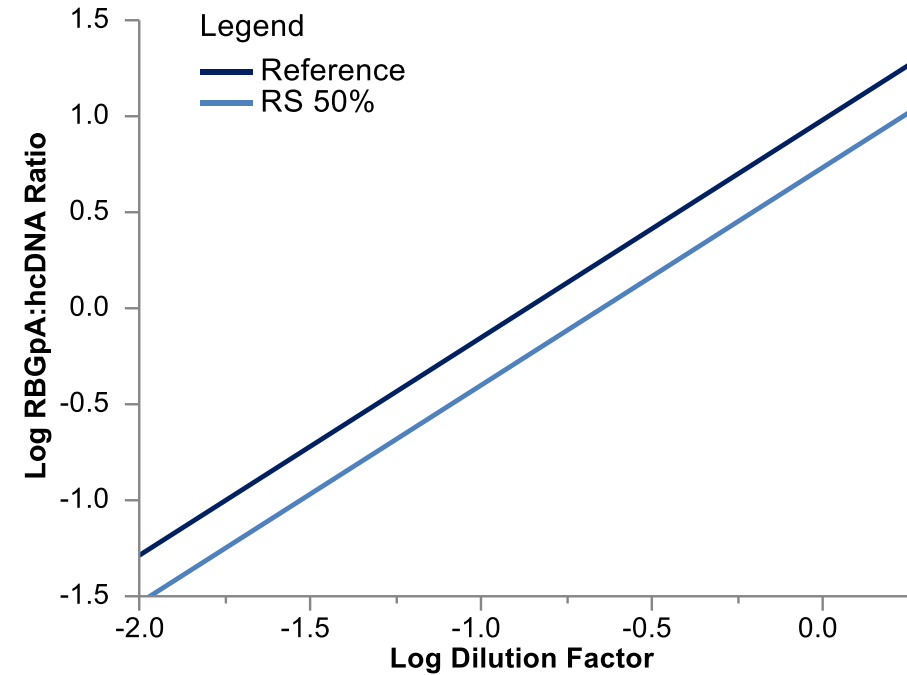


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Example relative infectivity method results using reference material



Expected Value = 150%
Measured Result = 150.4%



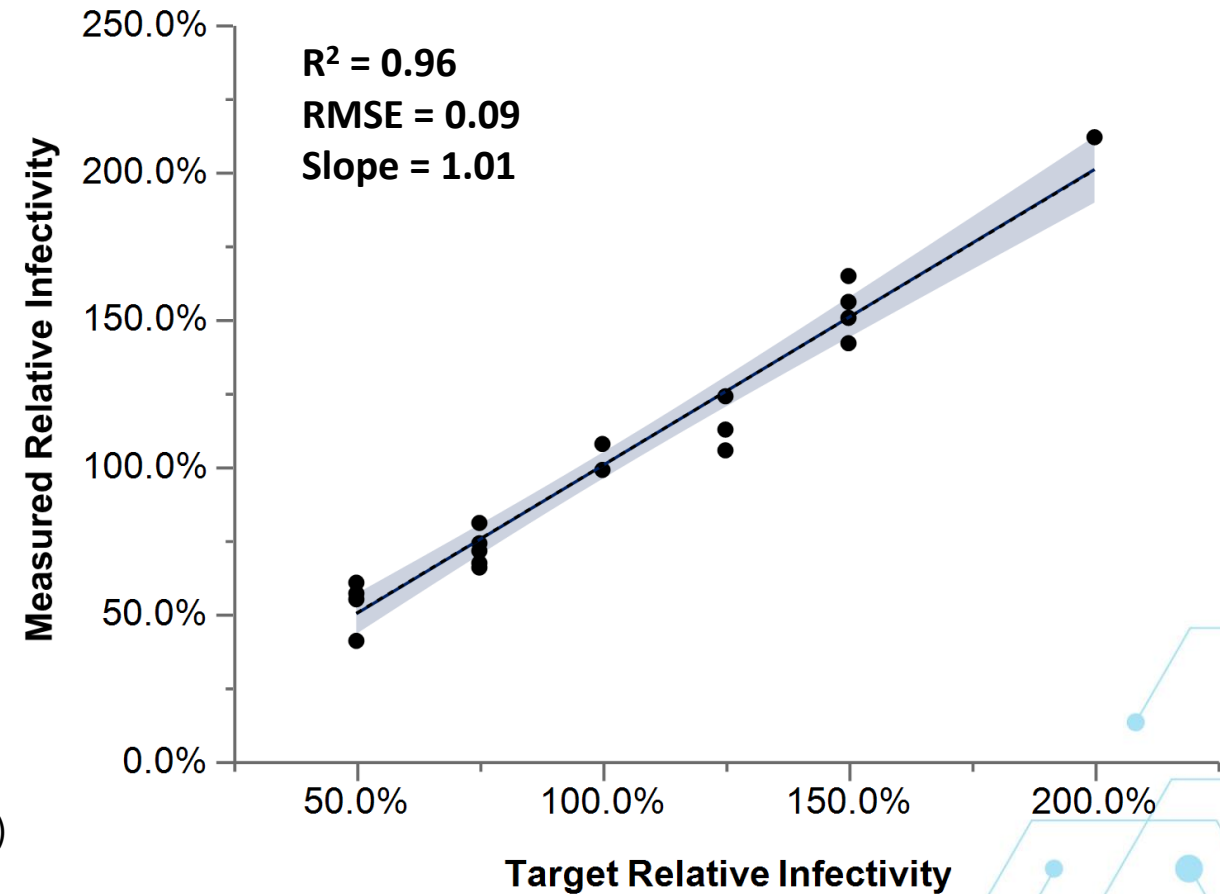
Expected Value = 50%
Measured Result = 56.9%

The relative infectivity method is linear, accurate, and precise

- The relative infectivity method is capable of quantifying relatively small differences in the *in vitro* infectivity of AAV vectors

Target Relative Infectivity	Measured Relative Infectivity	% Accuracy
200%	212%	106%
150%	153%	102%
125%	114%	91%
100%	103%	103%
75%	72%	96%
50%	55%	109%
Overall %CV		11%

Tested using rAAV8 material (N = 20, collected over nine different runs)



Applications of the relative infectivity method

- Comparisons across different products

	Serotype	Relative Infectivity
Product A	AAV9	124%
Product B	AAV9	75%
Product C	AAV9	78%
Product D	AAV8	89%

Note: Different dPCR methods were used for the different products

- Comparisons across multiple batches of the same product (i.e., product comparability)

	Batch / Lot	Relative Infectivity
Product D	1	103%
Product D	2	108%
Product D	3	81%
Product D	4	102%
Product D	5	90%

Applications of the relative infectivity method

- Detect changes upon stress & stability

Condition		Relative Infectivity
Untreated Control	-80°C	99%
Thermal Stressed	60°C for 10 minutes	375%

- Compare the ability of vectors to infect different cells & conditions

Target Cells	Relative Infectivity
HEK293 (Reference)	100%
HEK293 with Modification A	147%
HEK293 with Modifications A and B	6,968%

- Assess improvements in infectivity for engineered AAV capsid variants
- Probe AAV infection kinetics

Limitations of the relative infectivity method

- **Accurate quantitation of the vector genome concentration is required for the test samples and the reference standard**
- **The use of a well-characterized reference standard with known biological activity or infectivity is critical**
- **The method is intended to measure intracellular vector genomes, and therefore does not provide a measure of target protein expression or biological activity of the transgene**

Conclusions

- We have developed a platform-based *in vitro* relative infectivity method that is capable of detecting differences as low as 25% in the infectivity of AAV vectors, representing a significant improvement over TCID₅₀
- The relative infectivity method is linear, accurate, and precise from at least 50-200% relative infectivity
- The relative infectivity method is capable of detecting a change in infectivity upon forced degradation
- The relative infectivity method may be a useful tool in early development for comparing infectivity across different preparations, products, serotypes, and target cells
- In the absence of a quantitative product-specific *in vitro* potency method, the relative infectivity assay is a more reliable tool than TCID₅₀ for supporting product comparability and monitoring product stability
- The relative infectivity method may provide a measure of product potency for early phase development until a quantitative product-specific *in vitro* potency method can be developed and implemented

Acknowledgments

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Thank You