

Dual dynamic staining electrophoretic detection and characterization of dsRNA contaminants in mRNA vaccines

Adriana Coll De Peña

**Senior Scientist - Tripathi Biomedical Engineering Group
Brown University**

Collaborators:

Nina Li, Matei Vaduva, Menel Ben Frej

Lloyd Bwanali, Somdatta Goswami

Motivation - Analytical Landscape

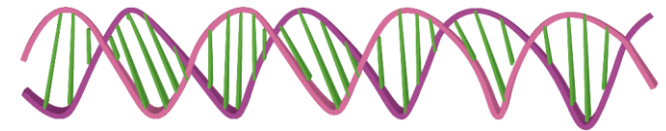
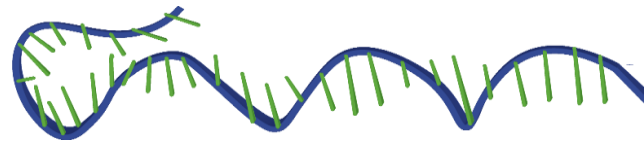
mRNA vaccines are at the forefront of the vaccine industry

- Safety and efficacy
- Shortened time between pathogen discovery and vaccine development

Problem: the *in vitro* transcription (IVT) of mRNA vaccines results in several by-products, and despite purification efforts, small concentrations of dsRNA can remain

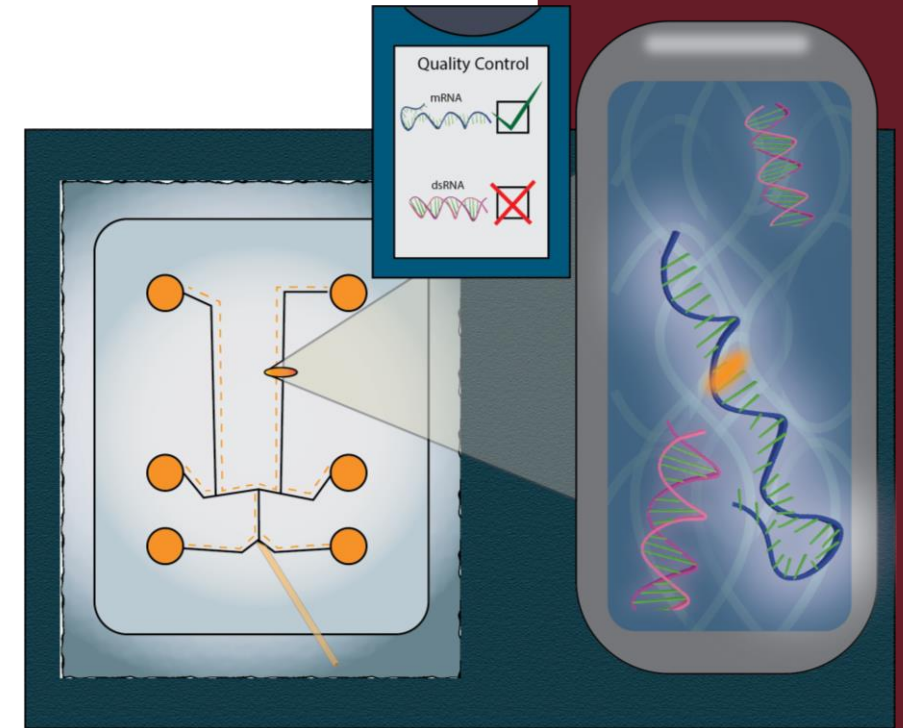
- dsRNA poses a length-dependent risk to humans

Current Analytical Technique



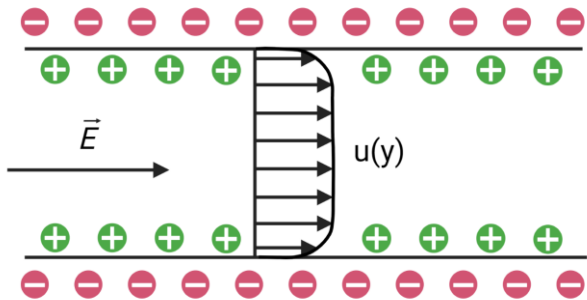
Method	Sensitivity	Resolution	Turnaround time	Throughput	Sample requirements
ELISA/Dot blot	Highest (pg/ μ L)	n/a	2-4 h/plate	High	Lowest

Goal: Develop a microfluidic electrophoresis **high throughput** and **high resolution** analytical method for the detection and characterization of dsRNA contaminants in mRNA vaccines.

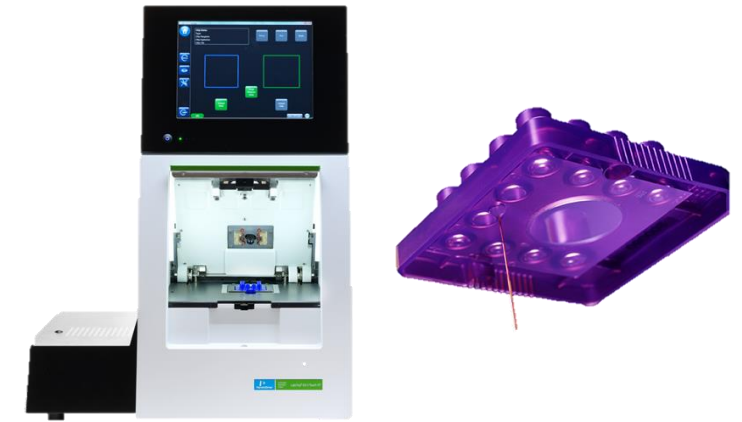


Analytical Platform

- Microfluidics: systems that manipulate small amounts of fluids (10^{-9} to 10^{-18} L)
- Usually coupled with a driving force to generate movement, here electric fields
- Electric fields were used to generate particle movement due to the ability to generate linear velocity profiles



Benefits



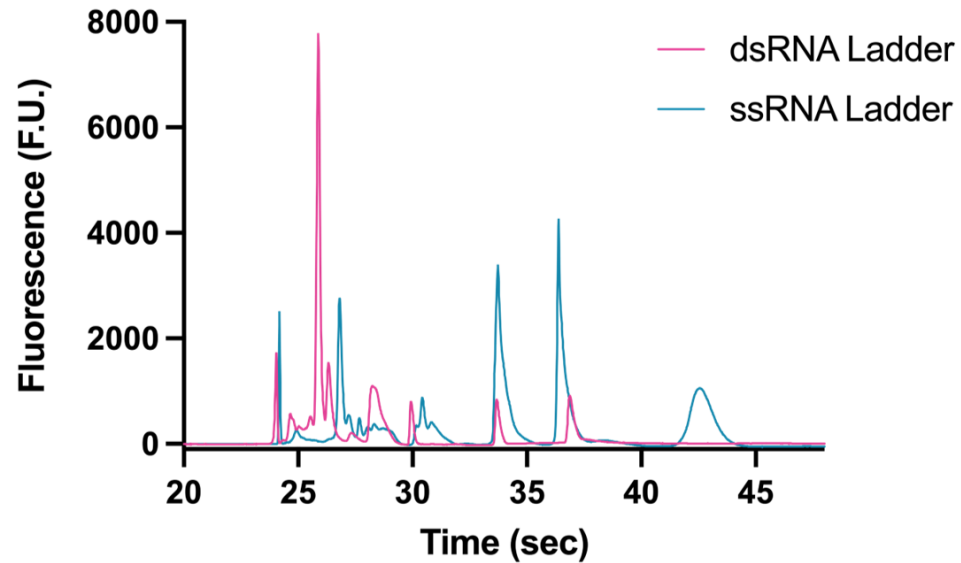
- System = high throughput
 - Scale = rapid
- Plug analysis = resolution

Work conducted in collaboration with Revvity

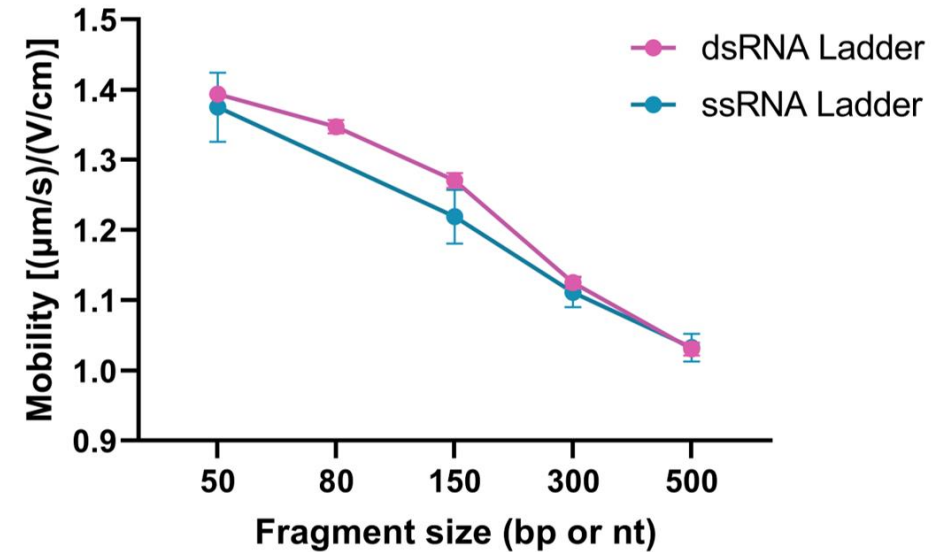


Analysis of dsRNA and ssRNA ladders

A.



B.

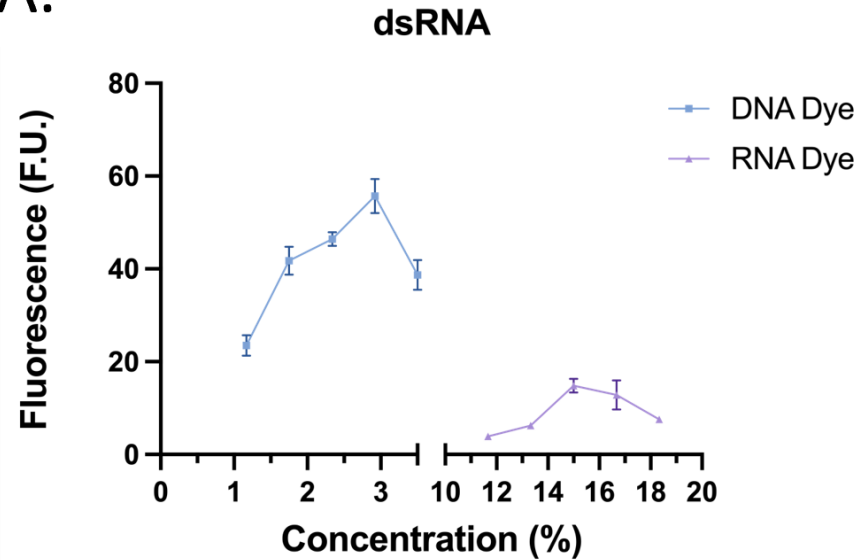


No mobility difference in the 50-500 bp or nt range - cannot differentiate based on mobility

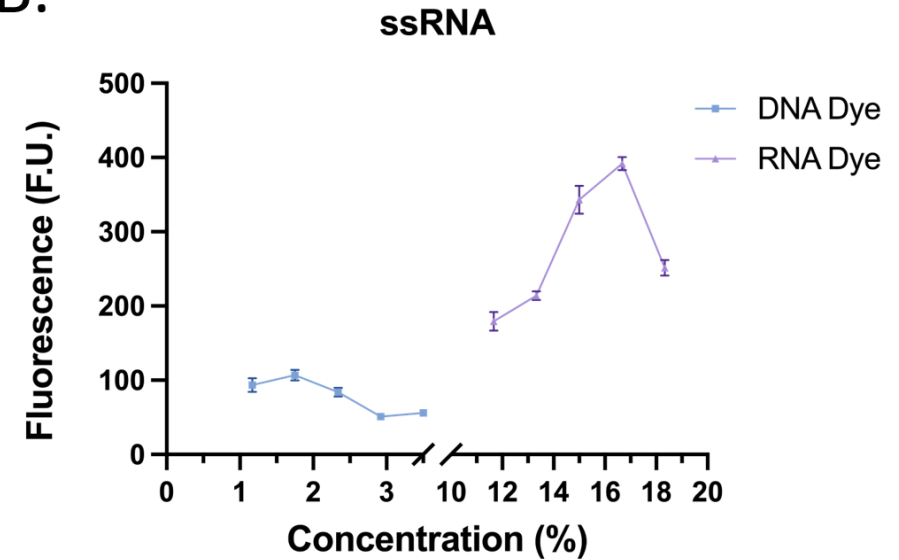
dsRNA and ssRNA Fluorescence Profiles

Dye concentration

A.



B.

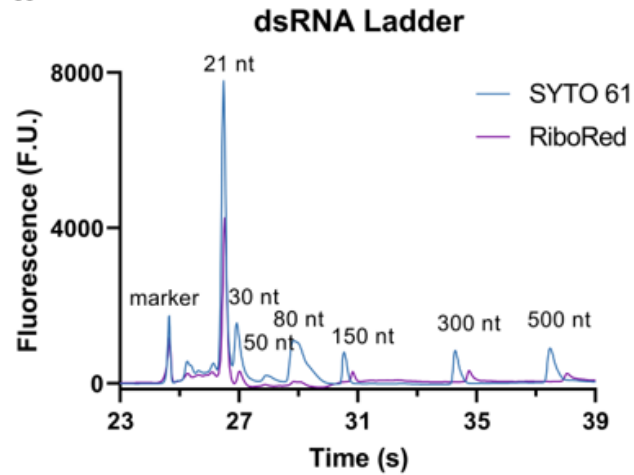


Efficiency of each stain is different for dsRNA and ssRNA

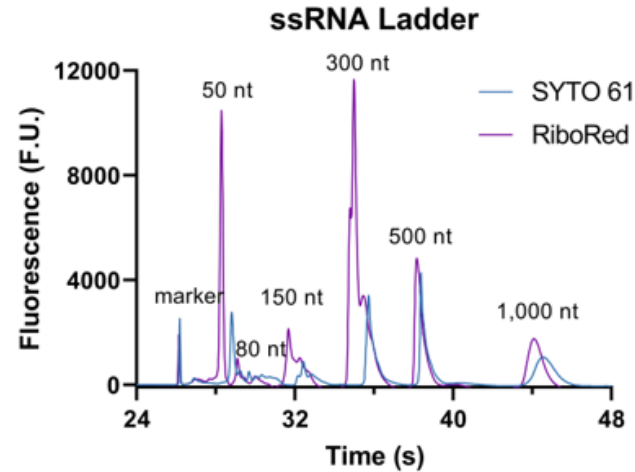


Fluorescent Staining Response

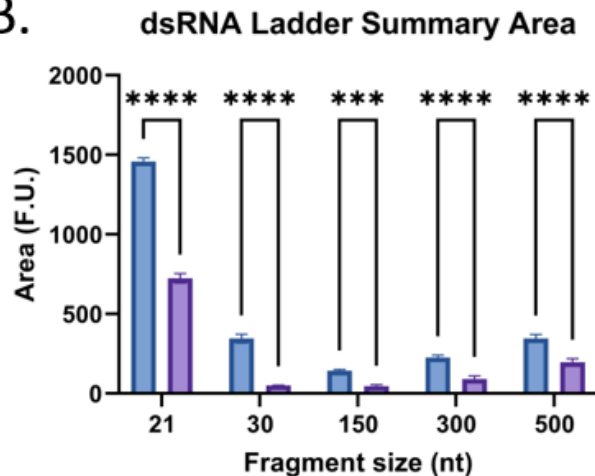
A.



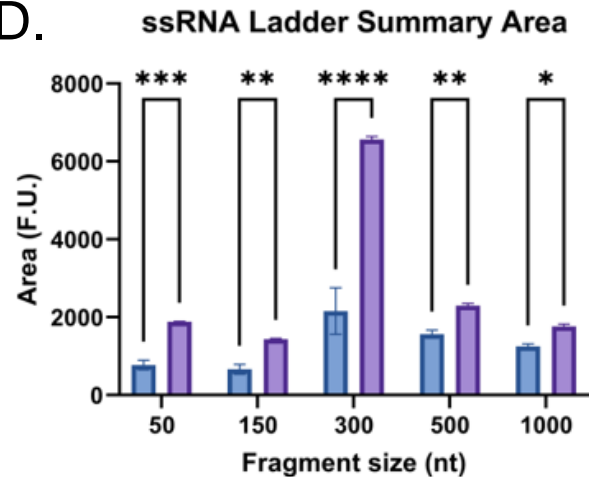
C.



B.

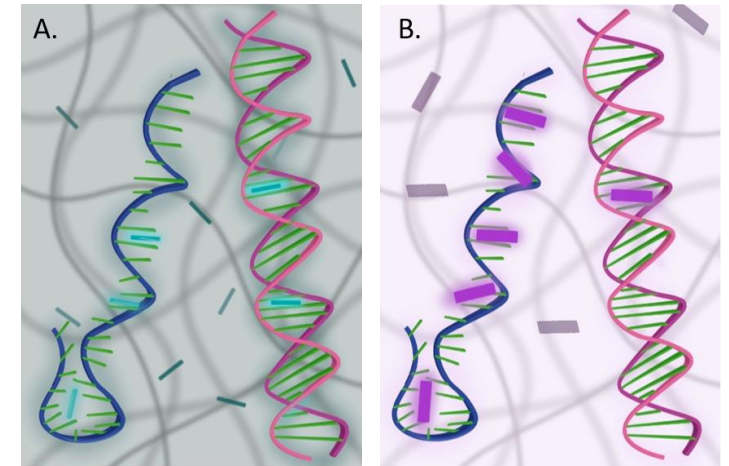


D.



DNA Stain

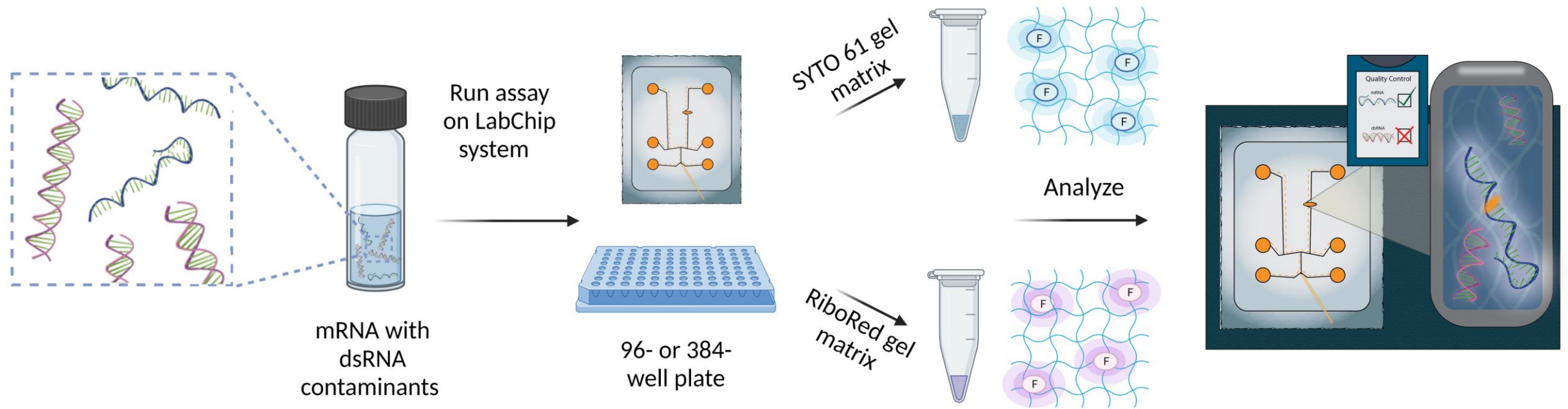
RNA Stain



dsRNA: SYTO 61 signal > RiboRed signal
ssRNA: SYTO 61 signal < RiboRed signal

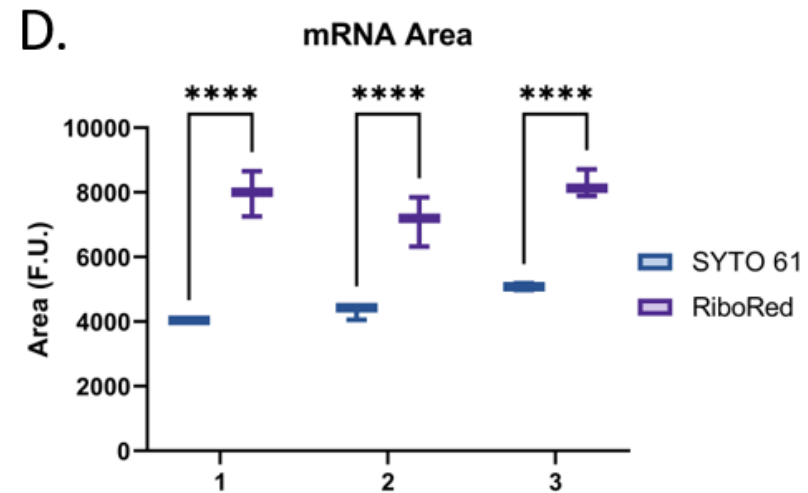
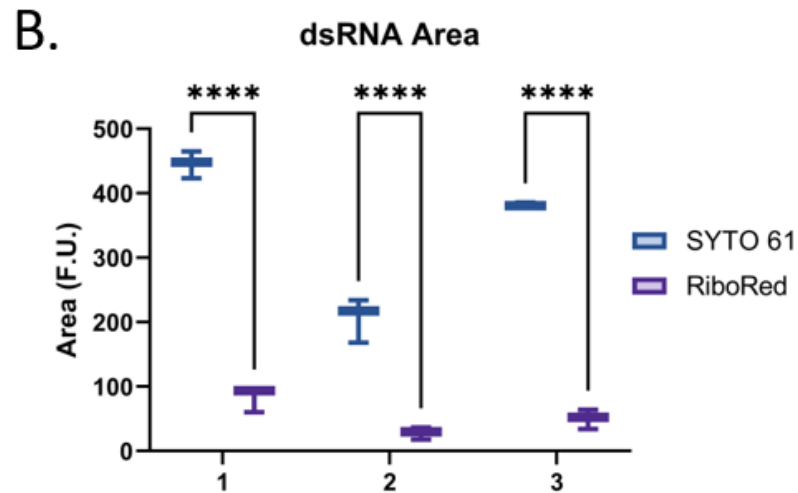
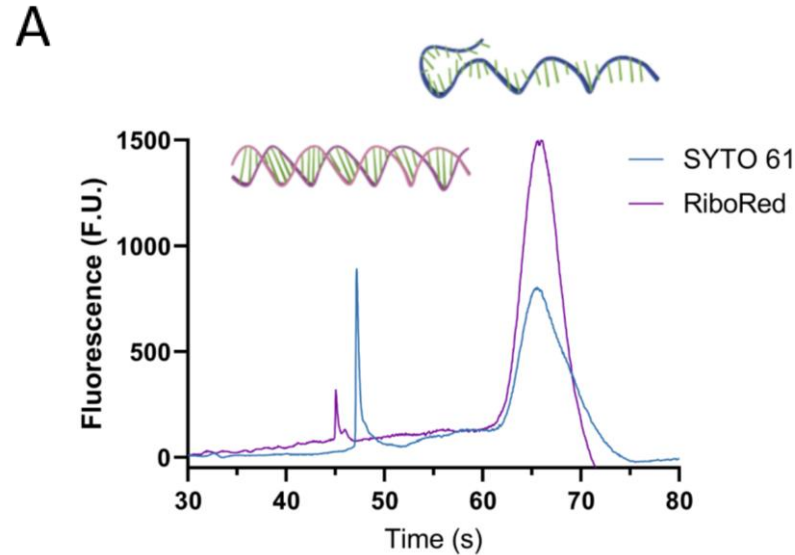


Identification Workflow



Application: dsRNA and mRNA mixture

- Same trend
- Areas vary



Peak classification/identification

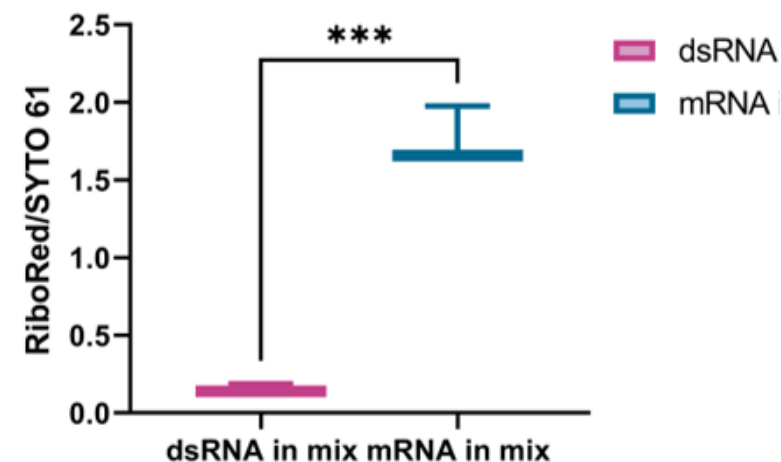
Since the total area can vary, we decided to look at the ratios instead

if:

$\frac{\text{Peak Area}_{\text{RiboRed}}}{\text{Peak Area}_{\text{SYTO 61}}} < 1$, then the peak producing molecule is dsRNA

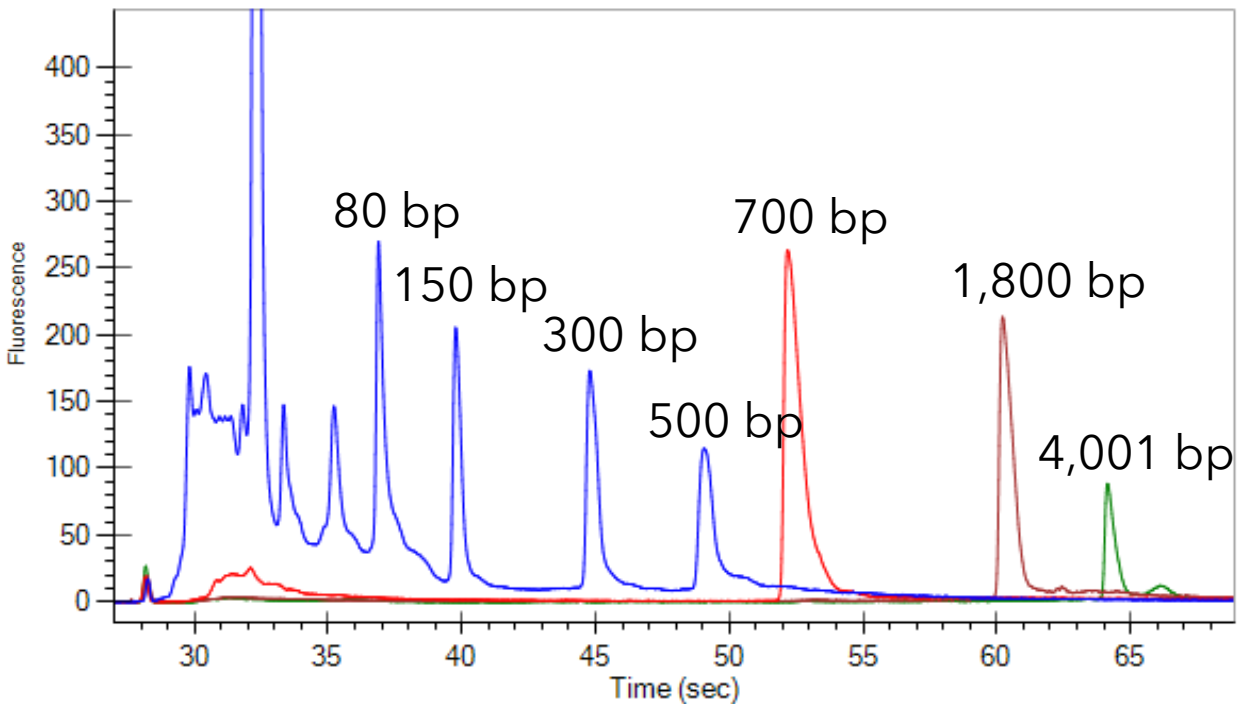
Or if:

$\frac{\text{Peak Area}_{\text{RiboRed}}}{\text{Peak Area}_{\text{SYTO 61}}} > 1$, then the peak producing molecule is mRNA (or ssRNA)

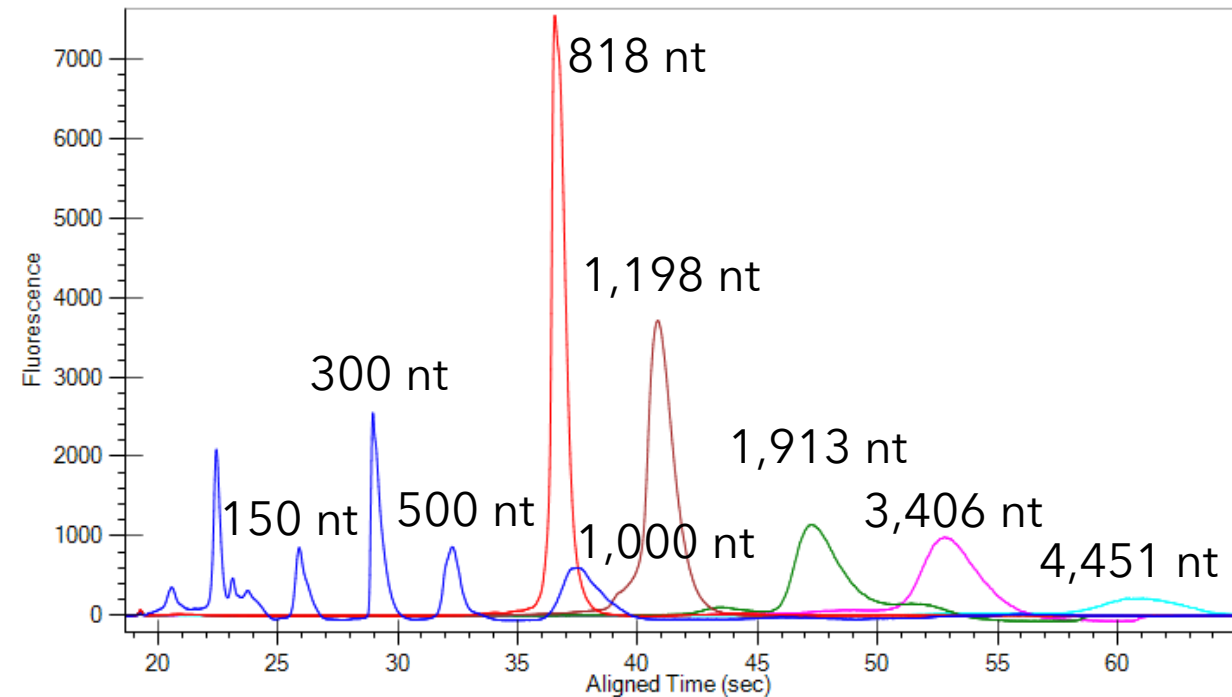


Size Calibration

dsRNA size calibration samples

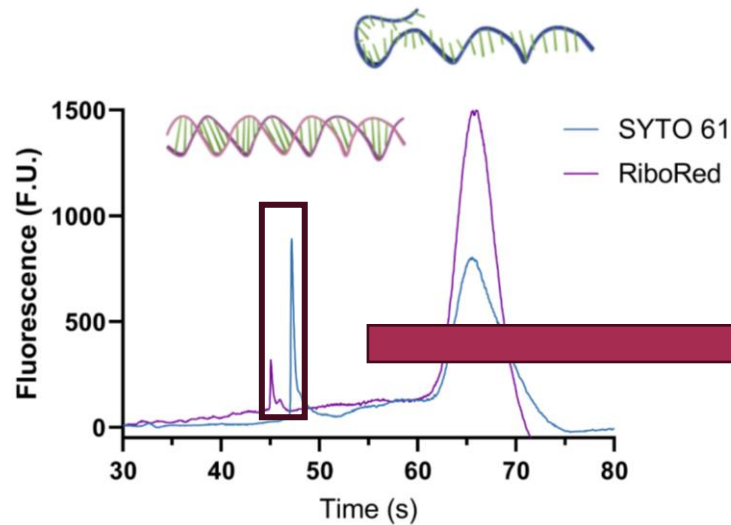


ssRNA/mRNA size calibration samples

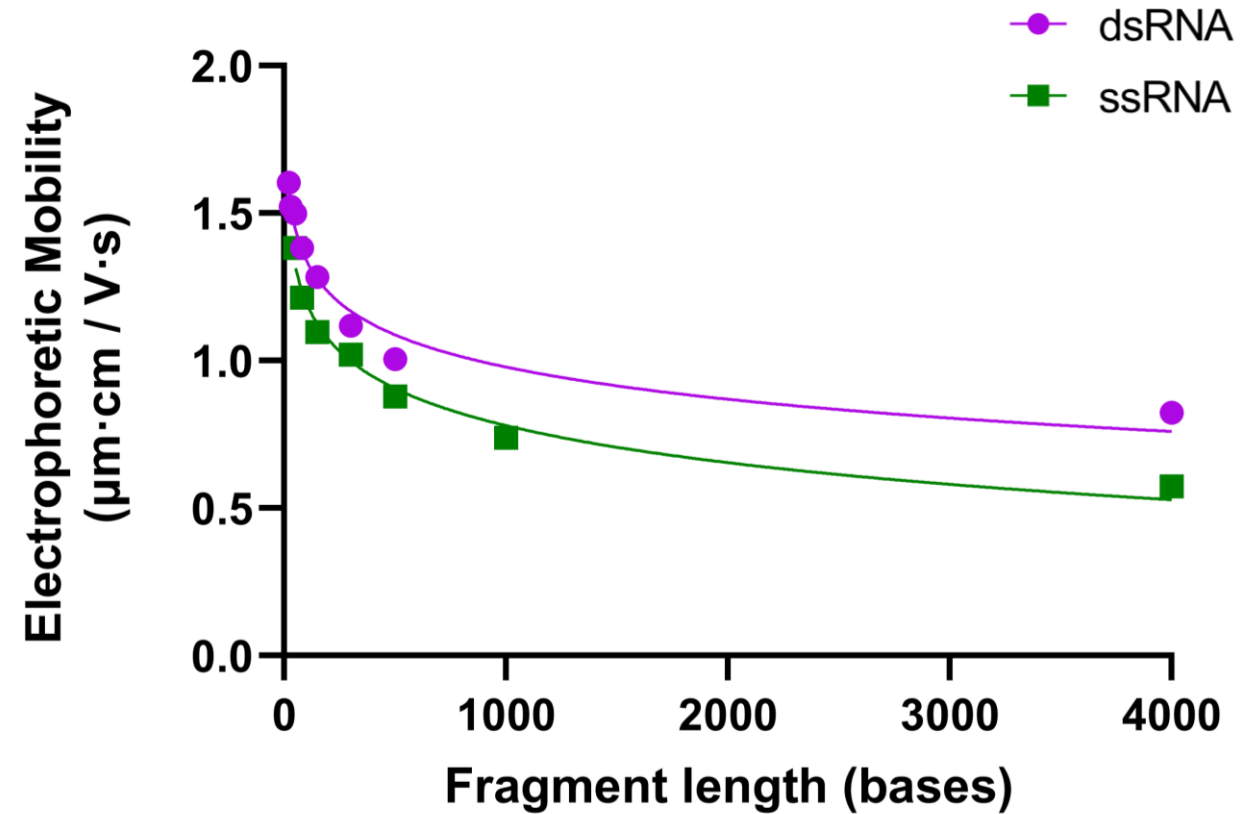


Application: Size determination

Mobility plot generated using calibration samples



Once the sample is identified, its mobility can be used to estimate the size of the contaminant.



Summary

- dsRNA and mRNA mobilities may be different but cannot be used for differentiation
- Fluorescent staining response can serve to identify molecules prior to size determination
- dsRNA ladder can then be used to estimate contaminant length

Method	Sensitivity	Resolution	Turnaround time	Scalability	Sample requirements
ELISA	Highest	n/a	2-4 h/plate	High	Lowest
Microfluidics	High (relative)	High	2-3 h/plate	High	Low



Acknowledgments



BROWN

Tripathi Lab

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Benjamin Phelps
Connor Yew
Misheel Damdinchoimbol
Sophia Esch
Aryan Narayan

Microfluidics Team

Current Members:

Nina Li

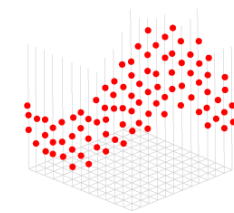
Matei Vaduva

Nicole Chen
Everett Gutterman-Johns



revvity

Lloyd Bwanali
Menel Ben Frej



**DATA
SCIENCE
INITIATIVE**

**Brown University Data
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Somdatta Goswami, Ph.D.

Assistant Professor, Applied
Mathematics

Questions?