



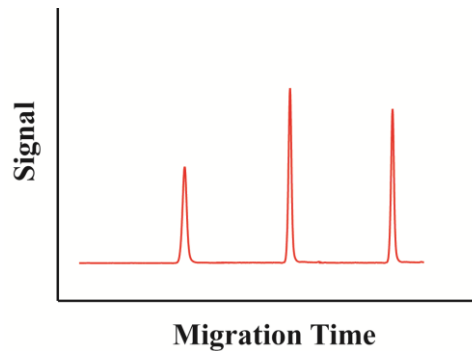
MICHIGAN STATE
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Automated and high-resolution cIEF-MS methods for delineation of proteoforms and protein complexes

Michigan State University
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CE Pharm 2021



Capillary isoelectric focusing (cIEF)

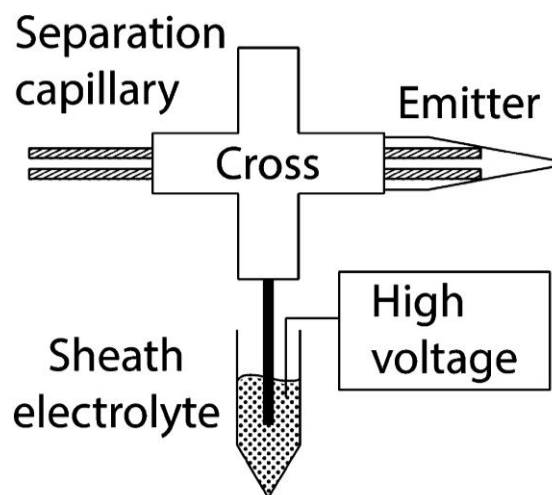
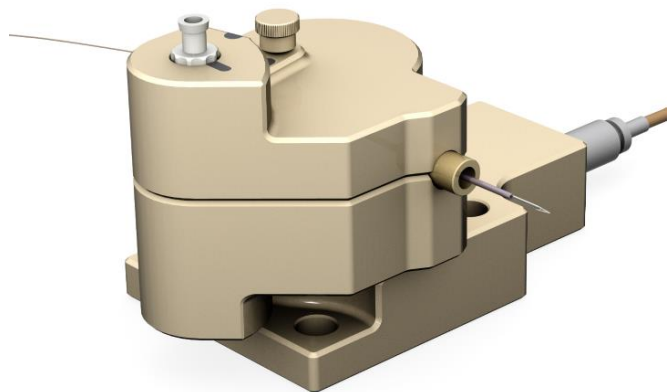


- Separation based on analytes' pIs
- 50-100 μm i.d. capillary with neutral coatings (linear polyacrylamide)
- Features
 - Ultrahigh resolution for protein separation (ΔpI : 0.001)
 - Concentrate and separate analytes simultaneously
 - High sample loading capacity (μL)
 - Accurate pI information of proteoforms



Automated cIEF-MS

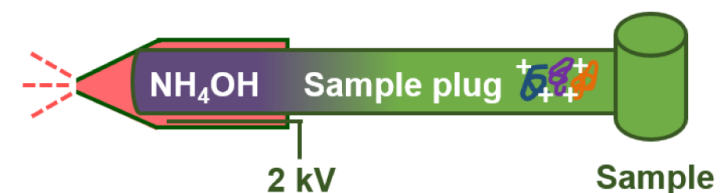
EMASS-II CE-MS interface



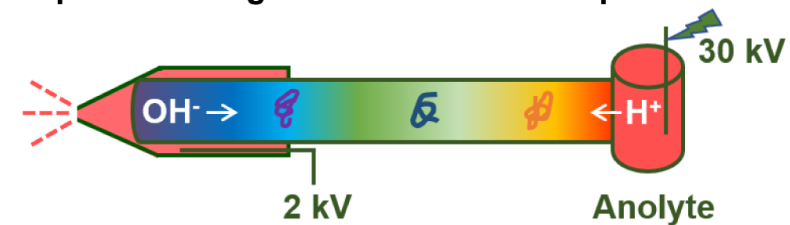
Automated cIEF-MS/MS method

“Sandwich” sample injection configuration

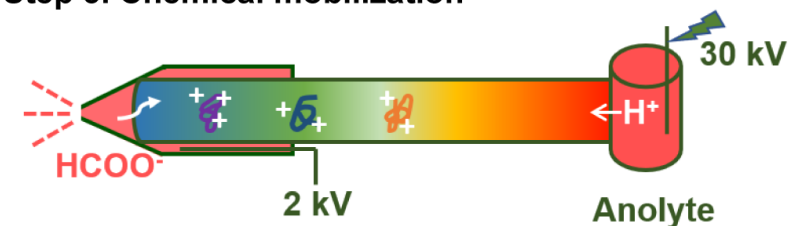
Step 1. Catholyte and sample injection



Step 2. Focusing based on isoelectric point



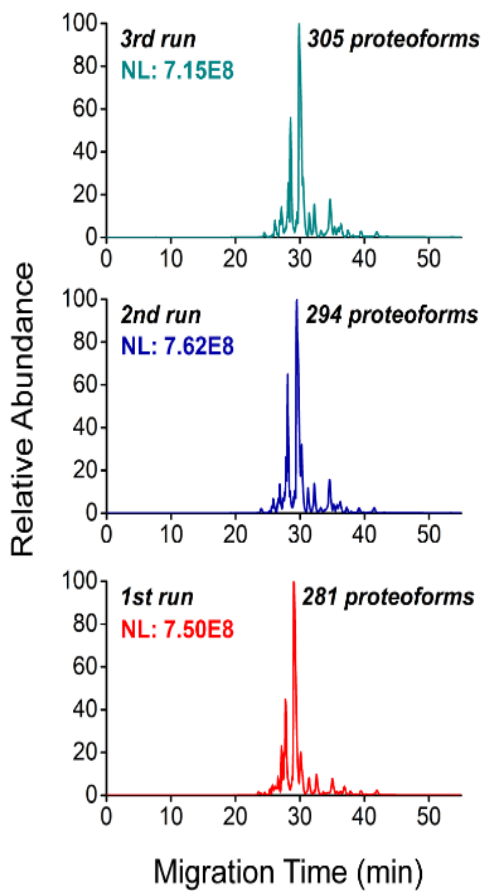
Step 3. Chemical mobilization



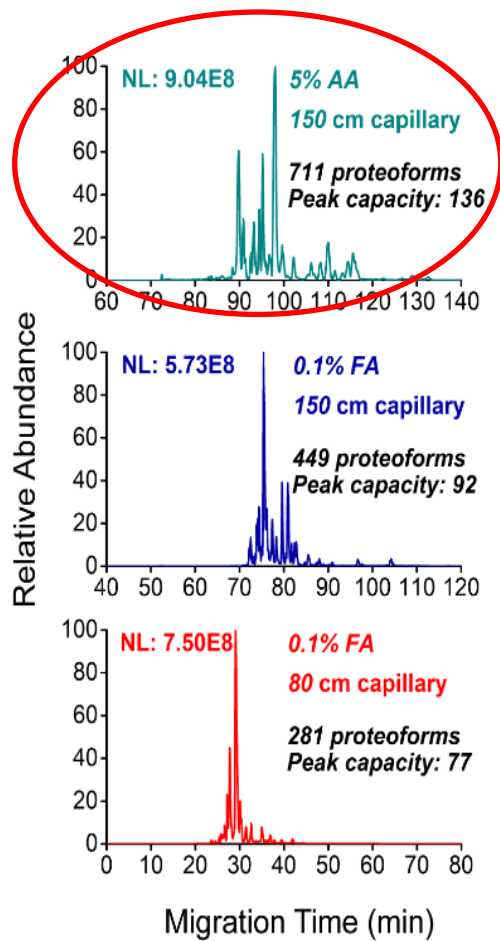


Automated cIEF-MS/MS for TDP of *E. coli*

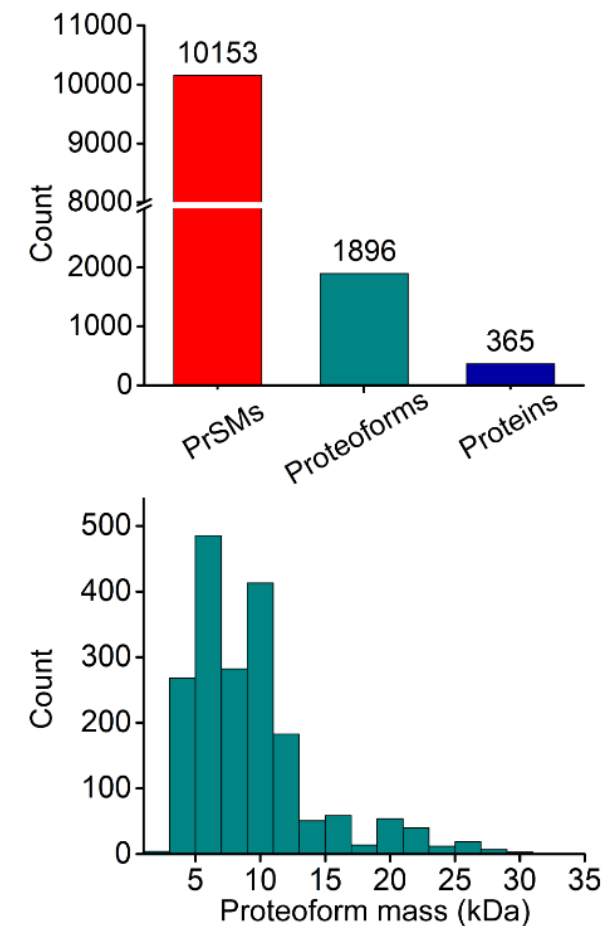
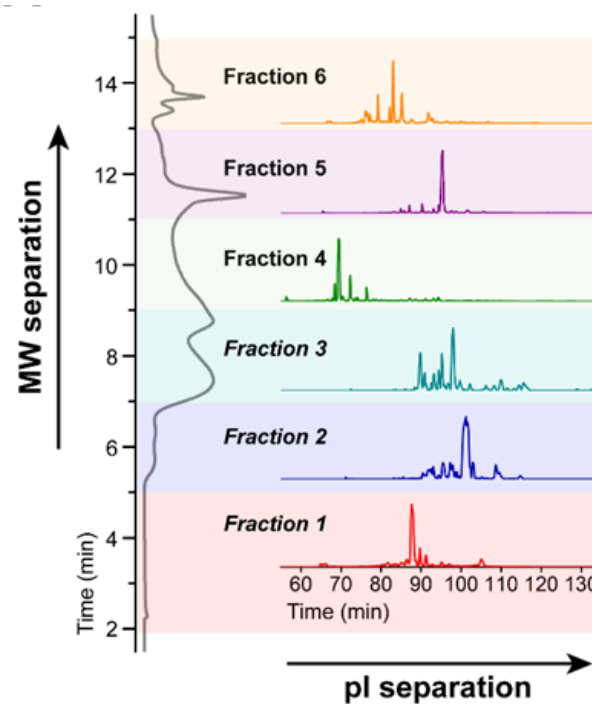
High throughput



High capacity



SEC-cIEF-MS/MS analysis of *E. coli*

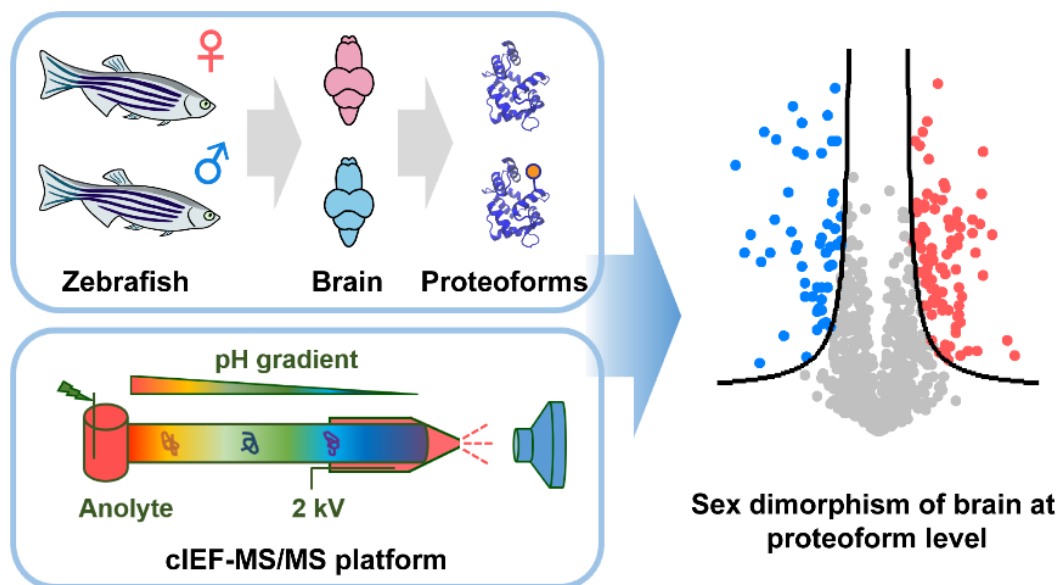


The data represents the first TDP dataset using cIEF-MS/MS



Quantitative TDP of zebrafish male and female brains

- Sexual dimorphism of brains: *the expression of sex chromosome genes and effects of hormones secreted from gonads.*
- **No TDP studies have been done to study the sex dimorphism.**
- Zebrafish is an important model organism in developmental biology for both embryogenesis studies and drug development.



Label-free quantification (LFQ)

TopPIC software from
Dr. Xiaowen Liu's group
(Tulane University)



LFQ of zebrafish brain proteoforms

A

Calmodulin; mass: 16779.83 Da;
63 fragment ions; E-value: 5.07e-019

Acetyl

M]A D Q L T E E Q I A E F K E A F S L F
D K D G D G T I T T K E L G T V M R S
L G Q N P T E A E L Q D M I N E V D A
D I N G T I D F P E F L T M M A R K M
K D T D S E E E I R E A F R V F D K D
G N G Y I S A A E L R H V M T N L G E
K L T D E E V D E M I R E A D I D G D
G Q V N Y E E F V Q M M T A K

Mass shift:
42.0 Da

B

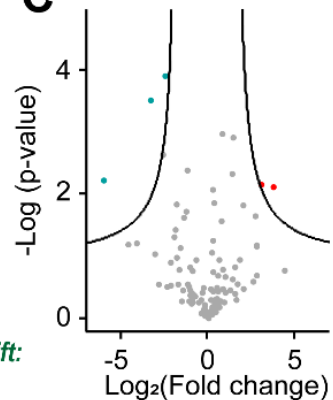
Caveolae-associated protein 4a;
mass: 6254.05 Da; 22 fragment ions;
E-value: 3.10e-007

N-terminal truncated

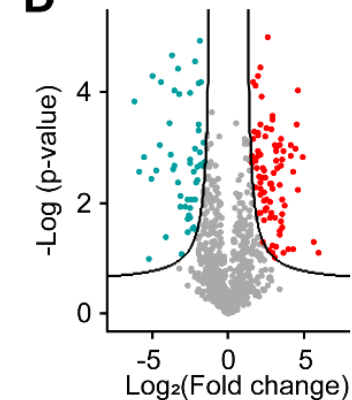
--- E S F K I K L K K E R T V A E G Q E
G A E A E P A V T P P K G R K S S P D
V T Y T E V V T E N K R E G P V S E E
G A T R I H E D

Mass shift:
79.0 Da

C

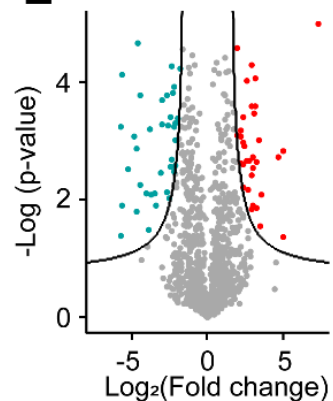


D

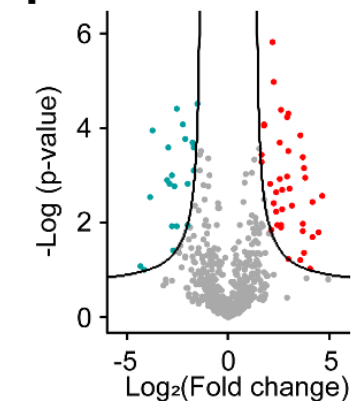


female/male

E



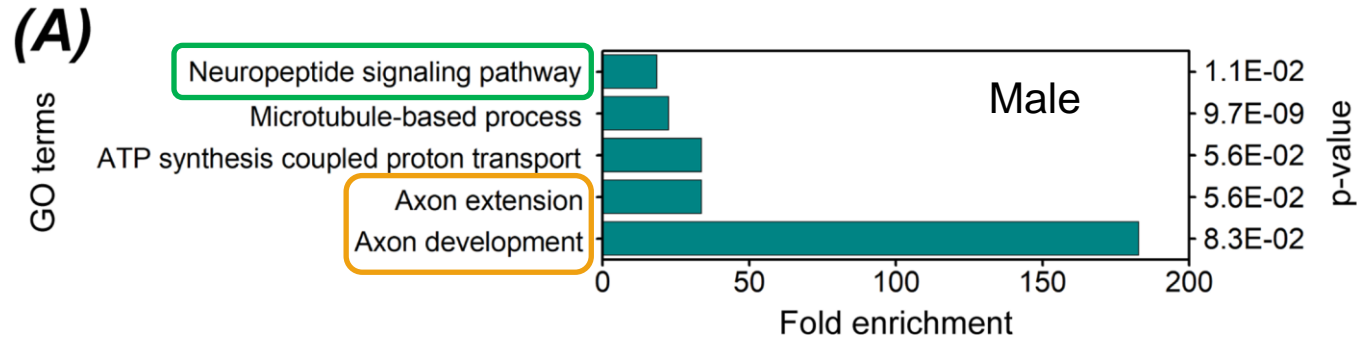
F



263 proteoforms showed statistically significant difference in abundance.

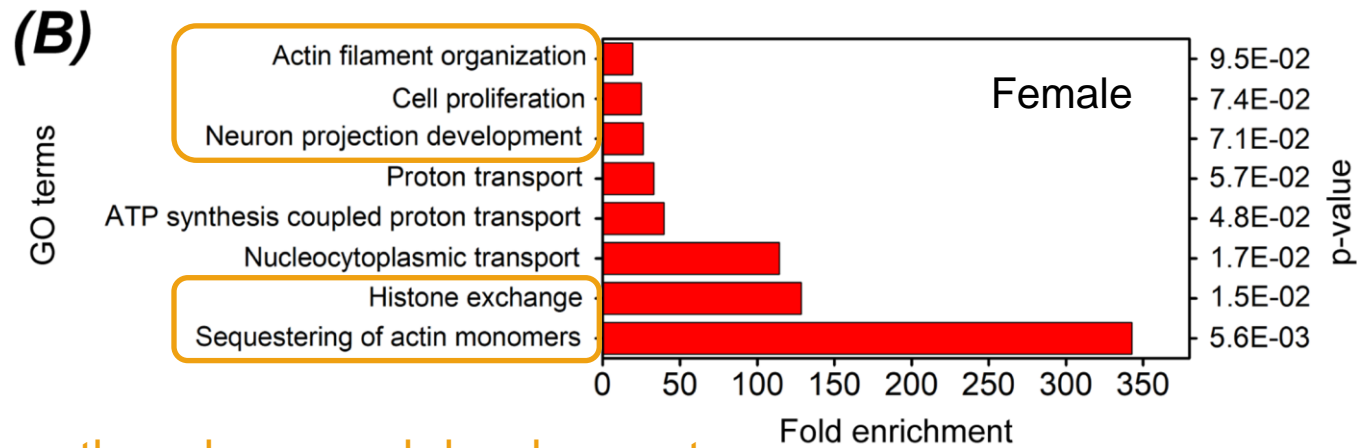


GO enrichment analysis of differentially expressed proteoforms



Peptide hormones

POMC and *PDYN*
(*pro-opiomelanocortin*
and *prodynorphin*)



Neurite outgrowth and neuronal development



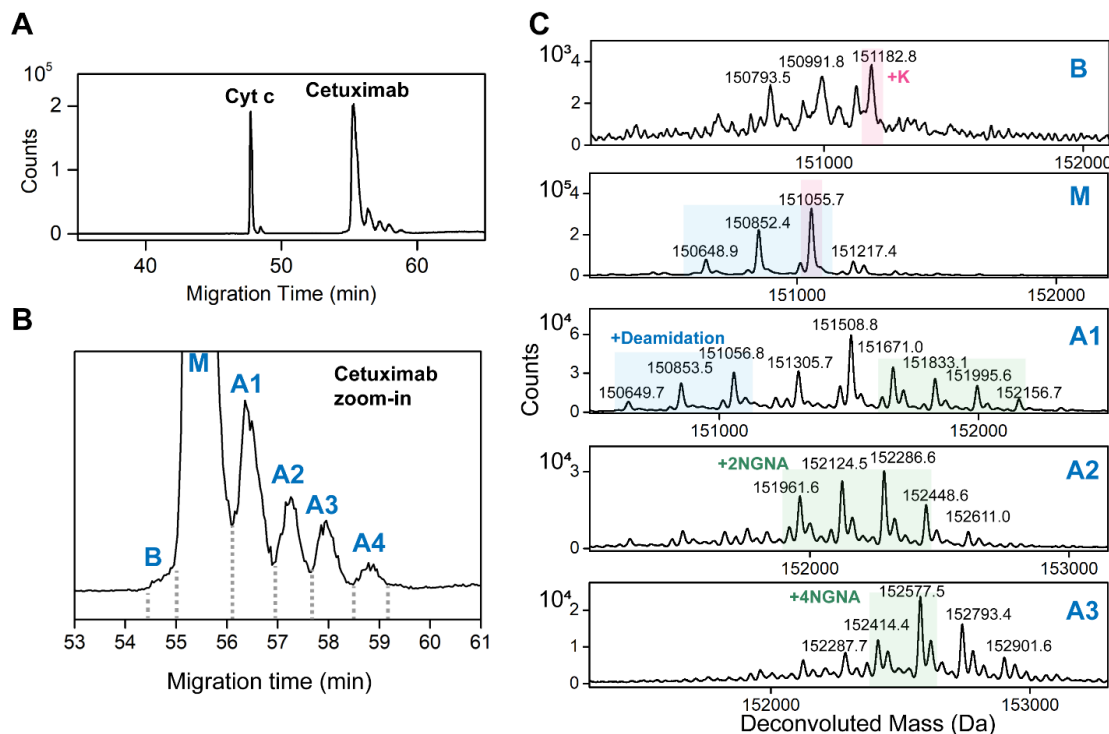
Automated cIEF-MS for monoclonal antibody (mAb) charge variants

- cIEF-MS is a powerful tool for characterizing mAb charge variants.
 - High-resolution separation.
 - automated operation.
 - much better identification of charge variants than the traditionally used cIEF-UV.
- Further improvement in stability and separation resolution is needed.
- Our work:
 - Bettered the quality of linear polyacrylamide (LPA) capillary coating.
 - Reduced catholyte pH to 10.
 - Systematically optimized the cIEF-MS separation conditions for characterizing of mAbs.



Charge variants of cetuximab

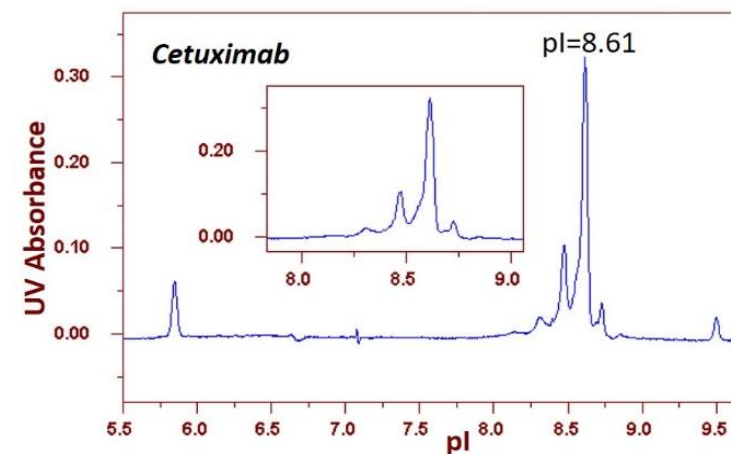
cIEF-MS



N-glycolyl neuraminic acid (NGNA)

Parameters for cIEF: 75 cm LPA-coated capillary, 30 cm catholyte plug, 45 cm sample plug, 0.8 mg/mL cetuximab, 0.05 mg/mL cytochrome c, 2% three-ampholyte mixture (pH range of 3-10, 5-8 and 8-10.5, ratio 1:2:4), 20 kV separation voltage, 10 mbar pressure applied at 20 min.

cIEF-UV

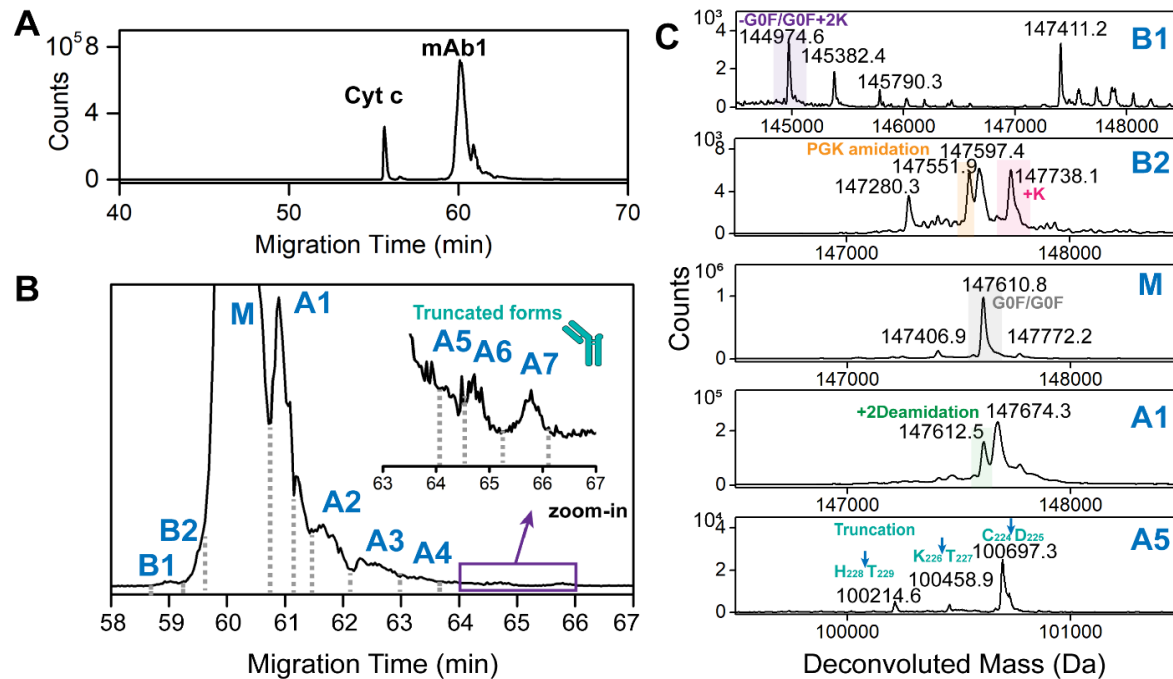


Cartridge: FC-Coated cIEF Capillary Cartridge, ProteinSimple Catalog No. 101701. Pharmalyte: Pharmalyte 3-10. Final prepared sample: 0.8 M Urea, 0.28% methyl cellulose, 4% carrier ampholytes, 0.2% pI 5.85 marker, 0.2% pI 9.50 marker, ~1.0 mg/mL protein. Focusing period 1: 1500V for 1 min; Focusing period 2: 3000V for 8 min. UV detection wavelength: 280 nm. System: iCE3 (ProteinSimple).

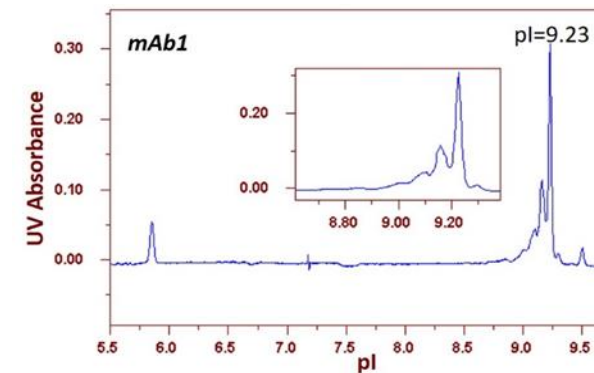


Charge variants of mAb1

cIEF-MS



cIEF-UV



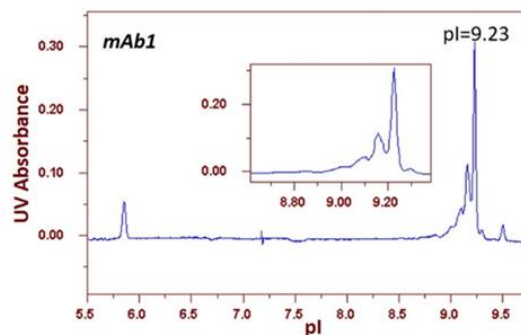
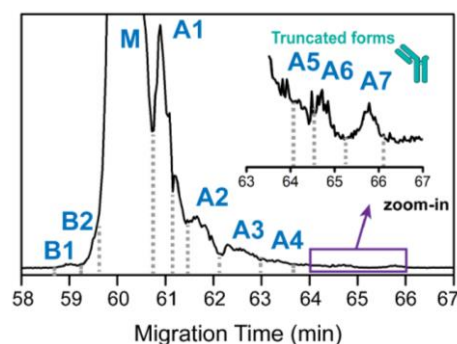
- Discovered ten charge variants with PTMs: PGK amidation, incomplete C-terminal lysine clipping, glycosylation, and deamidation.
- Achieved much more complete analysis than the cIEF-UV.



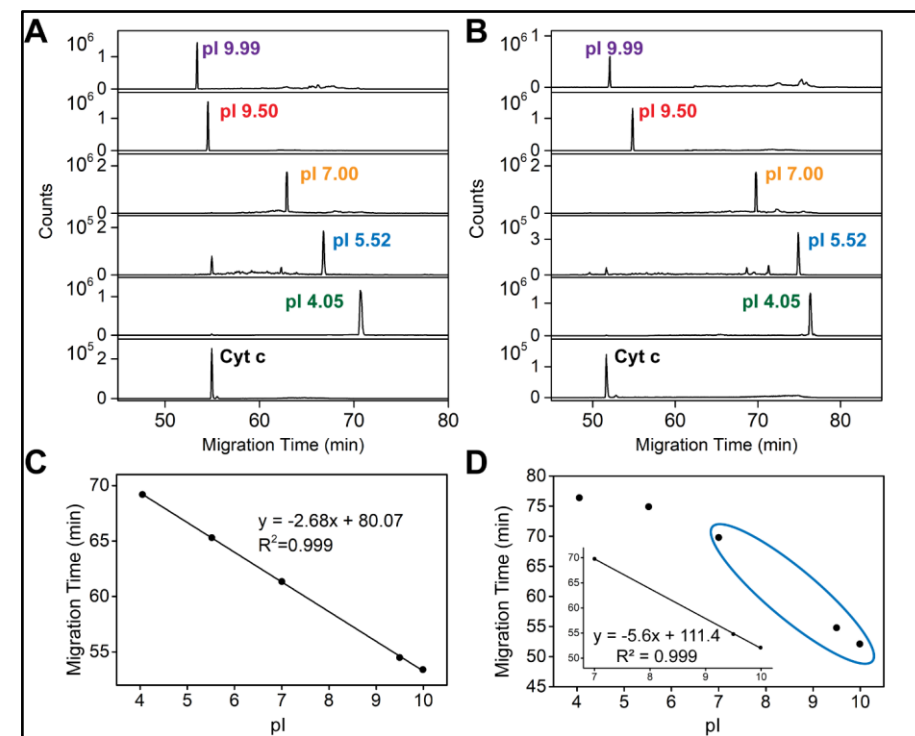
pI determination of mAb1 charge variants

	Data 1			Data 2			Data 3			Mean	SD
	t1	t2	pI	t1	t2	pI	t1	t2	pI		
M	59.6	57	9.15	53.3	56.9	9.17	56.3	56.9	9.17	9.16	0.01
A1	60.5	57.9	9	54.2	57.8	9.01	57.2	57.8	9.01	9.01	0.01
A2	61.4	58.8	8.85	55.2	58.8	8.85	58.2	58.8	8.85	8.85	0.00
A3	62.1	59.5	8.73	56	59.6	8.71	59	59.6	8.71	8.72	0.01

Main peak pI:
9.23 (UV) vs. 9.16 (MS)

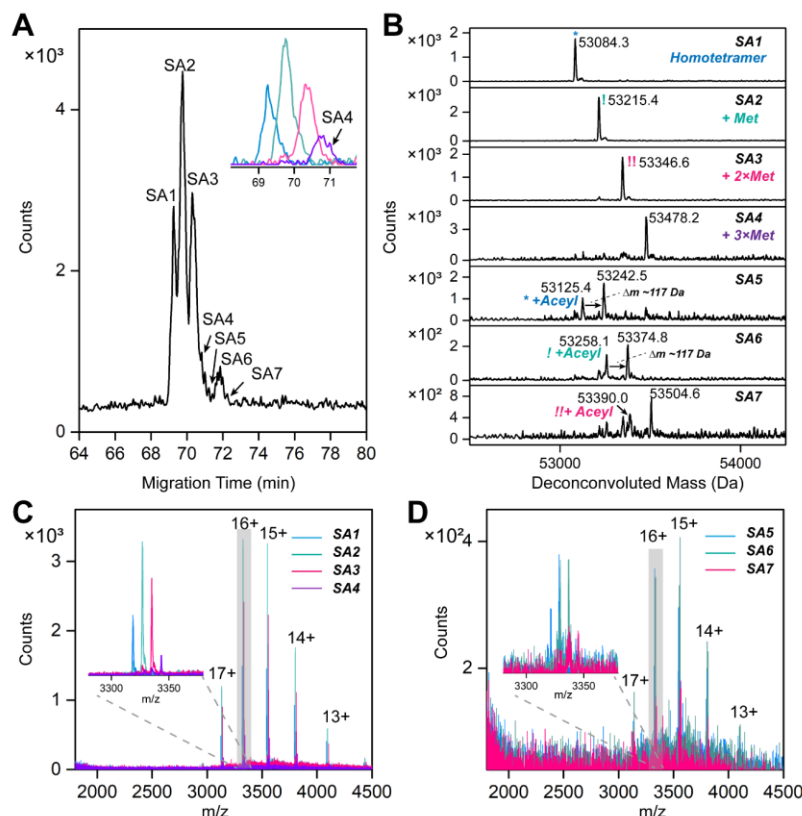


- Achieved reproducible measurements of pIs of charge variants (cyto.c for normalization)
- UV and MS data agreed well.
- Provide a useful approach for studying how PTMs influence pIs of proteoforms in TDP.





Automated and native cIEF-MS for protein complexes



- Streptavidin (SA): Homo-tetramer
- High-resolution separations of seven proteoform complexes of SA.
- Determined the pls of SA homo-tetramers and studied how sequence variations and acetylation influence the pl of SA homo-tetramer.
- Provide a useful tool for native top-down MS delineation of protein complexes.

Collaboration with Abbvie (Dr. Linjie Han)

SA variants	Normalized migration time (min)			pI			pI (Mean±STD)
	Run#1	Run#2	Run#3	Run#1	Run#2	Run#3	
SA1	62.82	62.60	62.70	7.36	7.36	7.40	7.37±0.02
SA2	63.09	62.84	62.90	7.26	7.29	7.33	7.29±0.03
SA3	63.38	63.26	63.22	7.16	7.16	7.22	7.18±0.03
SA4	63.67	63.43	63.50	7.07	7.06	7.12	7.08±0.04
SA+acetylation	64.14	63.73	63.90	6.90	6.94	6.99	6.94±0.04



Acknowledgments

- **Advisor:**
Prof. Liangliang Sun
- **Collaborator:**
Dr. Linjie Han (Abbvie)
- **Group members:**
 - Qianjie Wang
 - Qianyi Wang
 - Jorge Colonrosado
 - Fei Fang

Support:



abbvie