

Biologics Process Development Analytics

The Integration of Mass Spectrometry to the Process Development of Fusion Protein Therapeutics

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2021 CASSS MASS SPEC

Outline

- **Introduction**
- **Case Studies**
 - Fab Site-Specific Glycosylation Monitoring
 - LMW Analysis for Fc-fusion Protein
 - Complex Charge Profile of Fc-fusion Protein
- **Summary**
- **Acknowledgement**

Bioprocess Development Analytical Support



Analytical Support

- High-throughput Platform Assays
- Sample Automation
- PAT for Real time Analytics
- Mass Spectrometry for In-Process Support

Fc-fusion Protein Therapeutics

- Fc-fusion protein therapeutics are one of the most successful classes of IgG-based products;

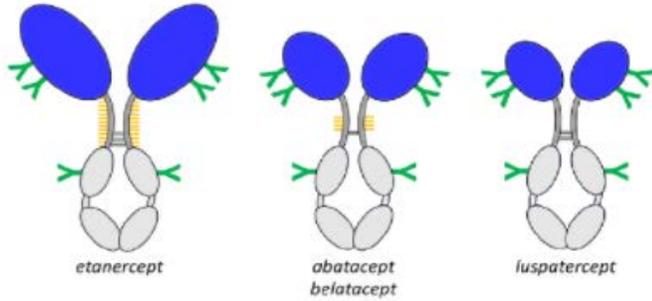
Product	Sales in 2019 (billion, USD)
Eylea	7.5
Enbrel	7.2
Trulicity	4.3
Orencia	3.2
Elocta	1.2

- Fc-fusion protein combine the pharmacological properties of biological ligands with the additional stability and inherent properties of IgG Fc domain;
- Fc-fusion protein can significantly improve the clinical potential of active protein drugs such as extend the plasma half life as well as engage immune-mediated effector functions;
- To date, ~**37** therapeutic fusion proteins are in clinical development and **13** products have been approved by the FDA, CFDA, and EMA;
 - Enbrel (TNFR-Fc fusion, FDA approval in 1998) and Orencia (CTLA4-Fc fusion, FDA approval in 2005) for treatment of rheumatoid arthritis;

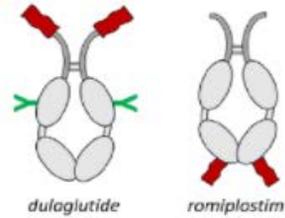
Duivelshof BL, et al. J Sep Sci. 2021. Jan; 44(1):35-62.

Major Groups of Fc-fusion Proteins

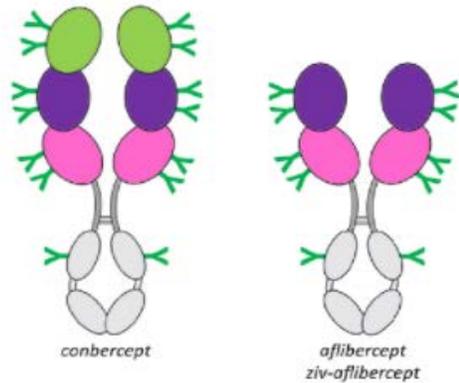
A) ECD-Fc



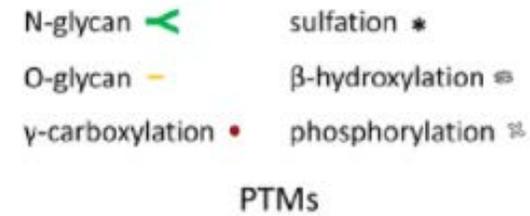
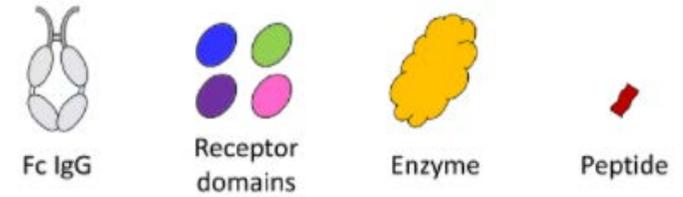
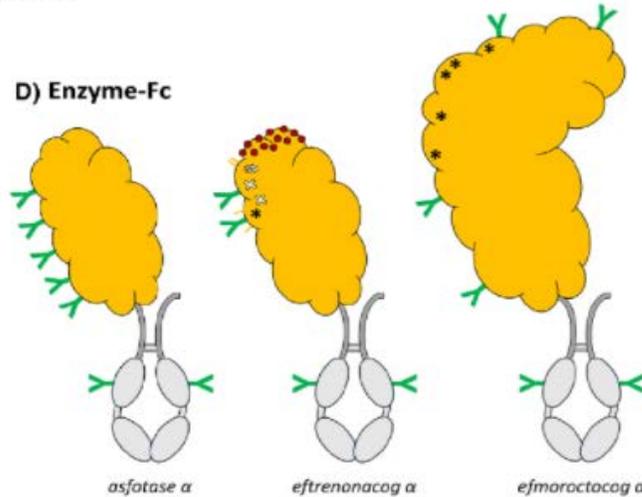
B) Peptide-Fc



C) Cytokines traps



D) Enzyme-Fc



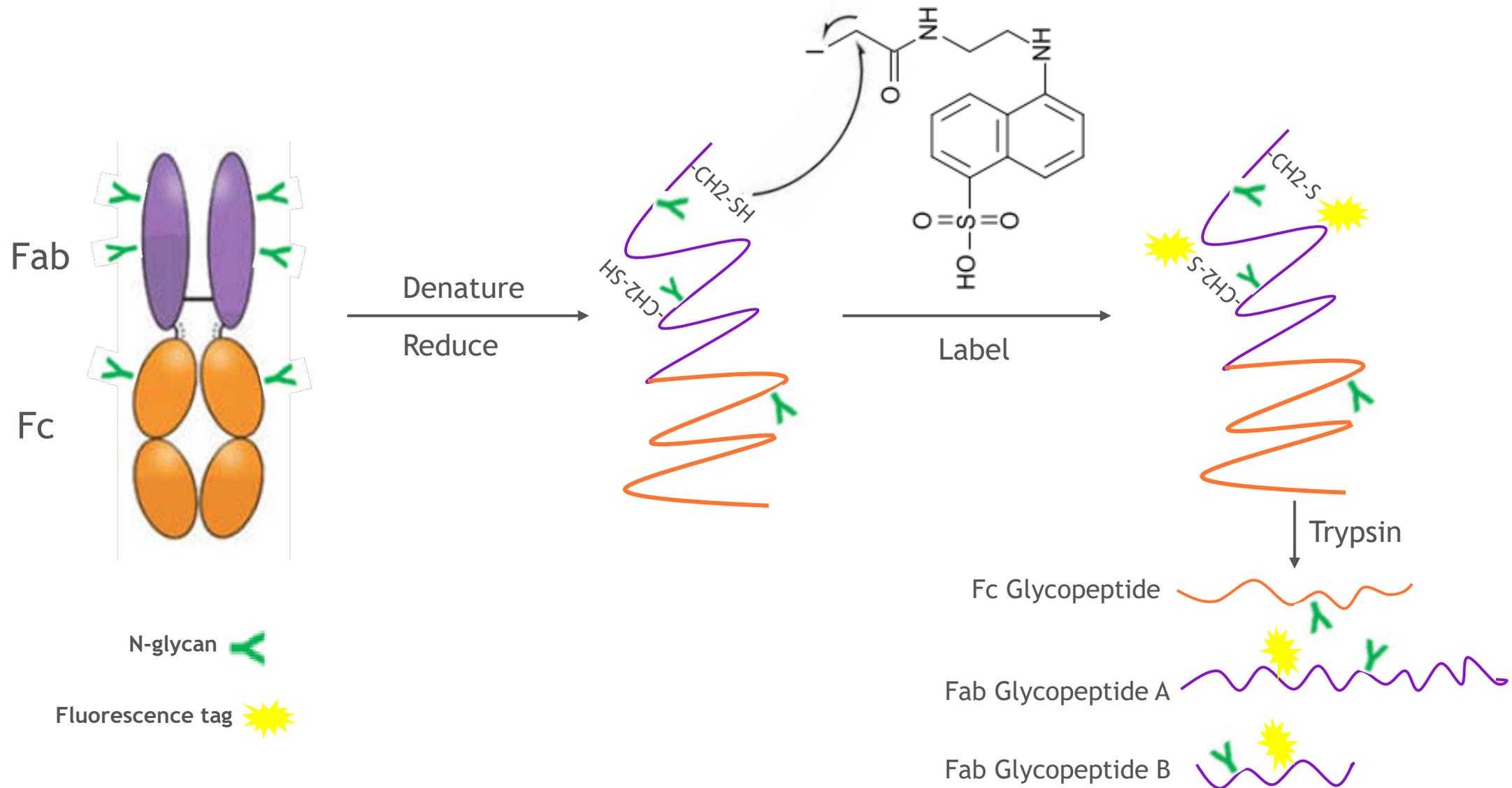
Therapeutic Fc-fusion proteins: Current analytical strategies
Duivelshof BL, et al. J Sep Sci. 2021. Jan; 44(1):35-62.

- Due to highly **heterogenous** structure (the presence of sialic acid, complex glycan structure, etc.), the analysis of Fc-fusion proteins is more challenging and complex than monoclonal antibodies;
- **Product-specific** methods over conventional generic or platform methods are often desirable to support process development;

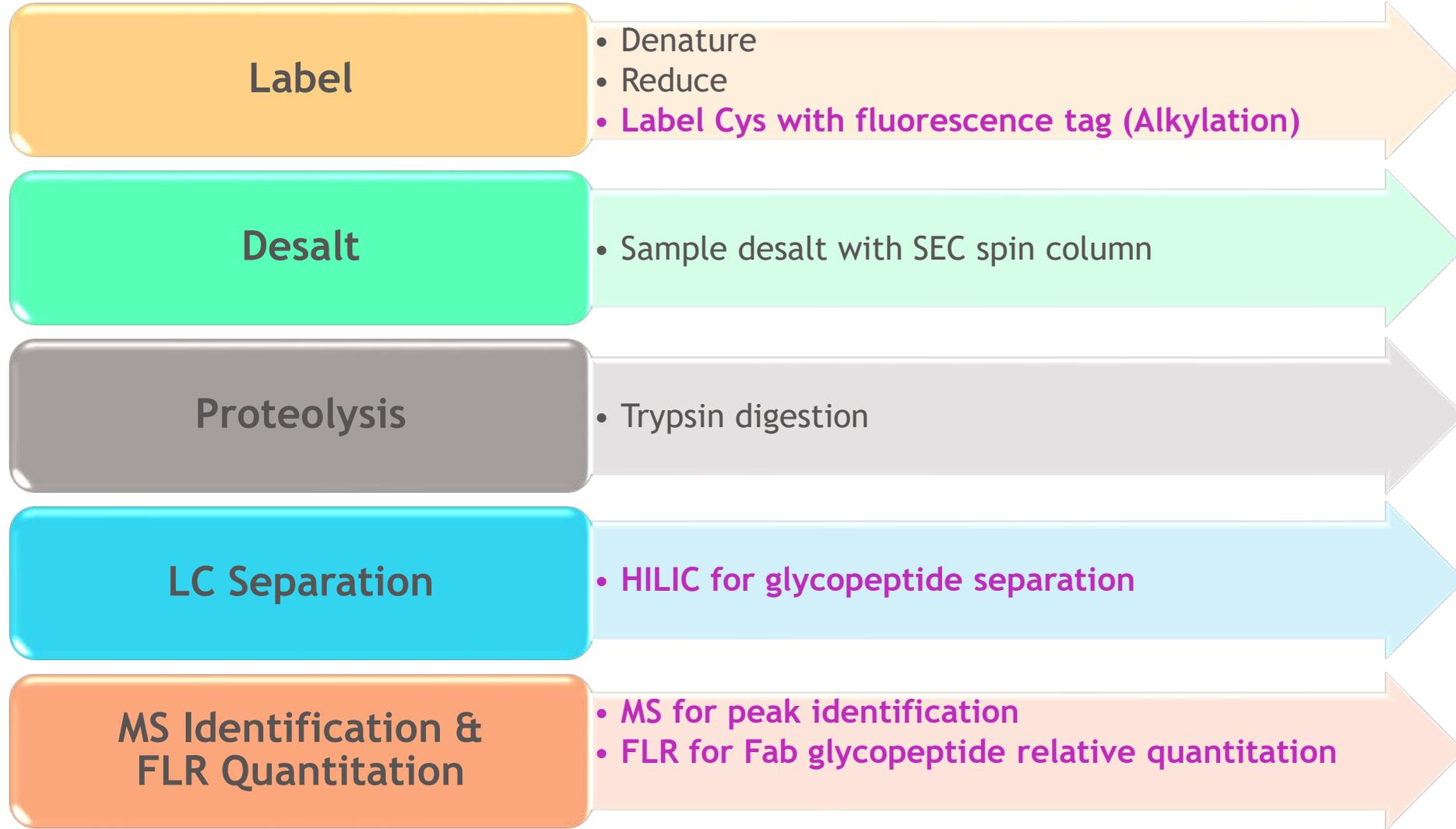
Case Study I: Fab Site-Specific Glycosylation Monitoring

- A Fc-fusion protein with **four** *N*-glycosylation sites in Fab region and **two** *N*-glycosylation sites in the Fc region;
- A LC method was developed to monitor Fab site-specific glycoforms (G0F, G1F, G2F, G2FS1, and G2FS2) as the understanding about site-specific glycosylation as pCQA is continuously evolving during product development lifecycle;
- Mass spectrometry is coupled with LC for the peak identification of the complex chromatogram and the optimization of LC method parameters;
 - Mobile phase screening
 - Fluorescence tag screening
 - LC gradient optimization

Glycopeptide Chemical Labeling

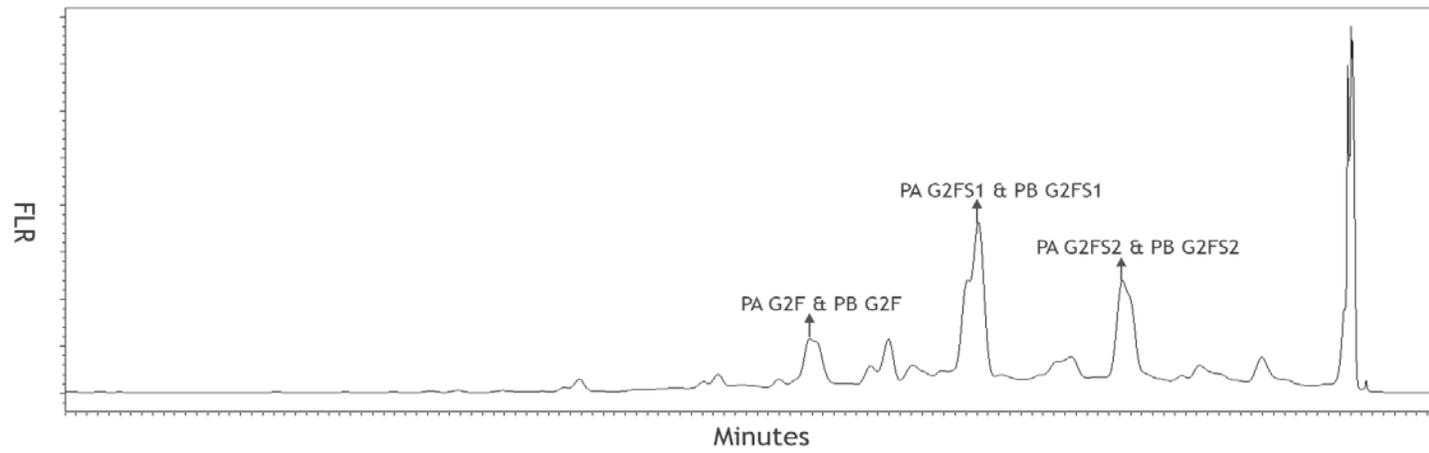


Workflow Overview



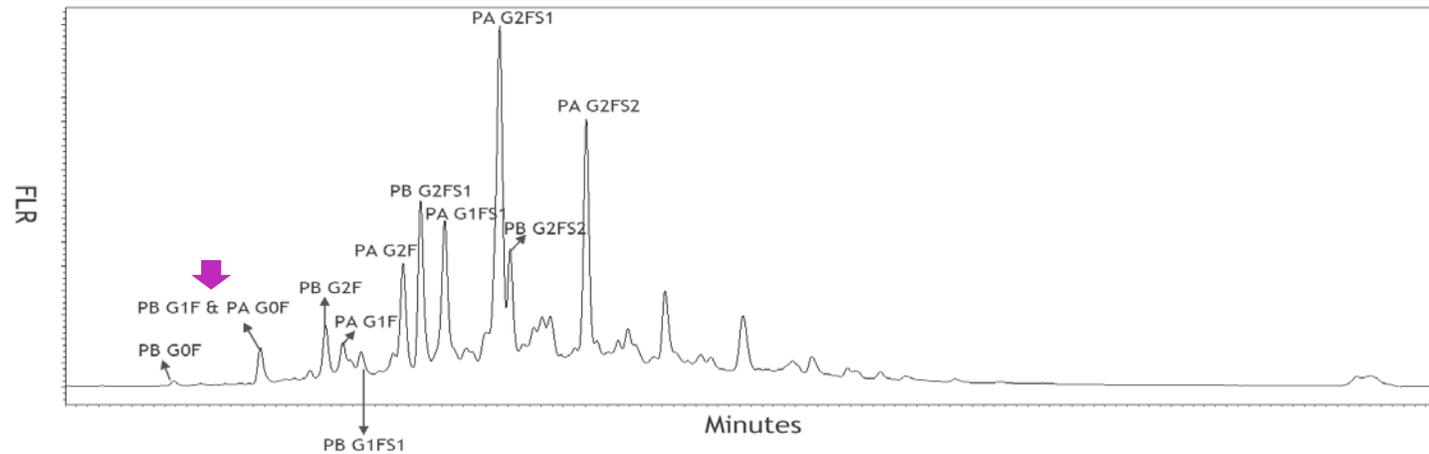
Ammonium Acetate vs Ammonium Formate as Mobile Phase

IASD, Ammonium Acetate



☐ Glycopeptide A and B coelution

IASD, Ammonium Formate

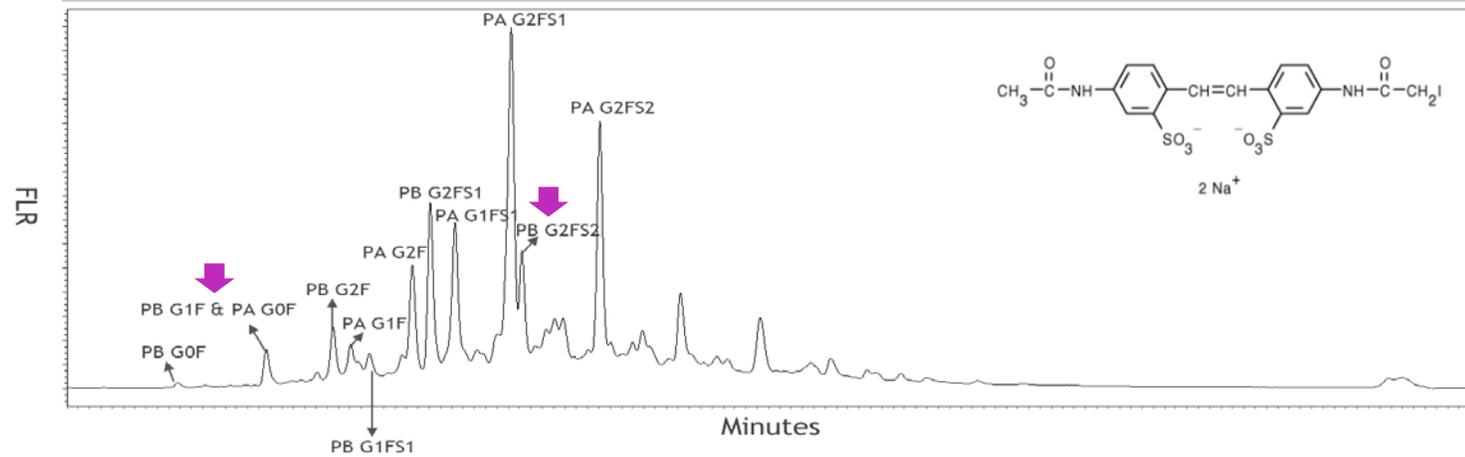


☐ Only Glycopeptide A + G0F and Glycopeptide B + G1F coelutes

- Significant improvement in separation resolution with ammonium formate for IASD FLR tag;

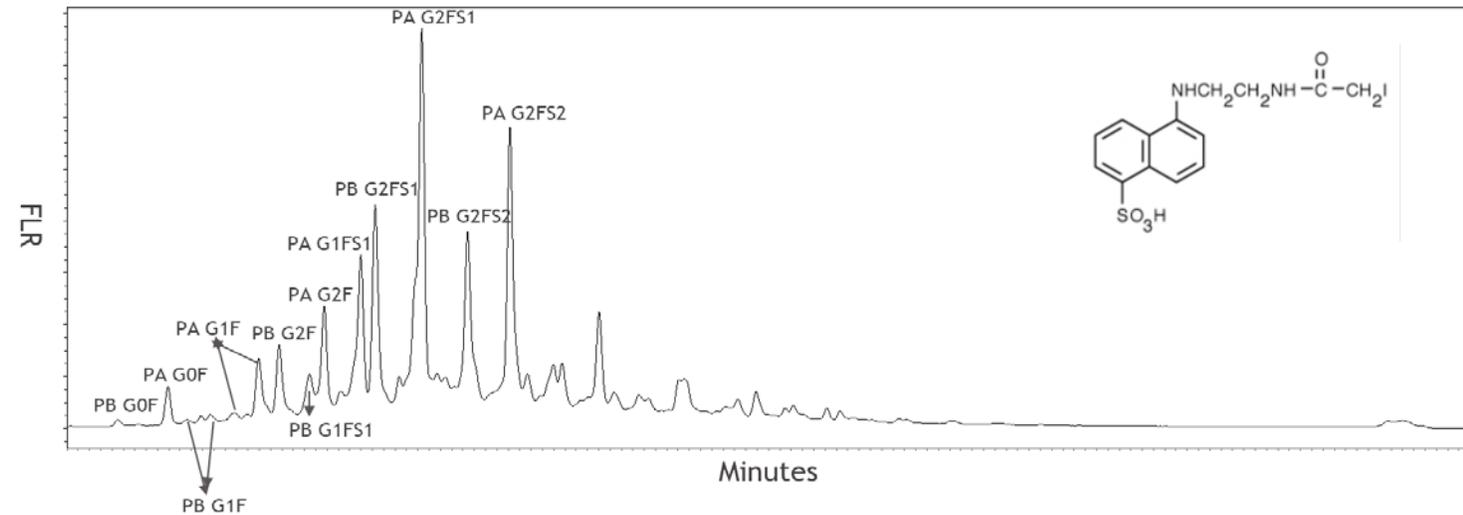
IASD vs IAEDANS as Fluorescence Tag

IASD, Ammonium Formate



- ❑ Only Glycopeptide A + G0F and Glycopeptide B + G1F coelutes

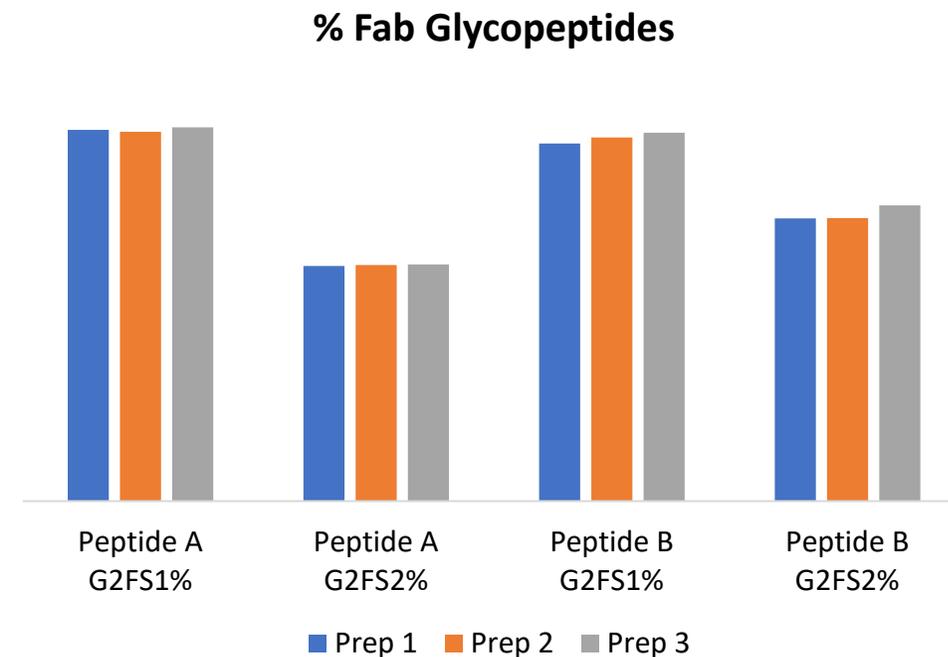
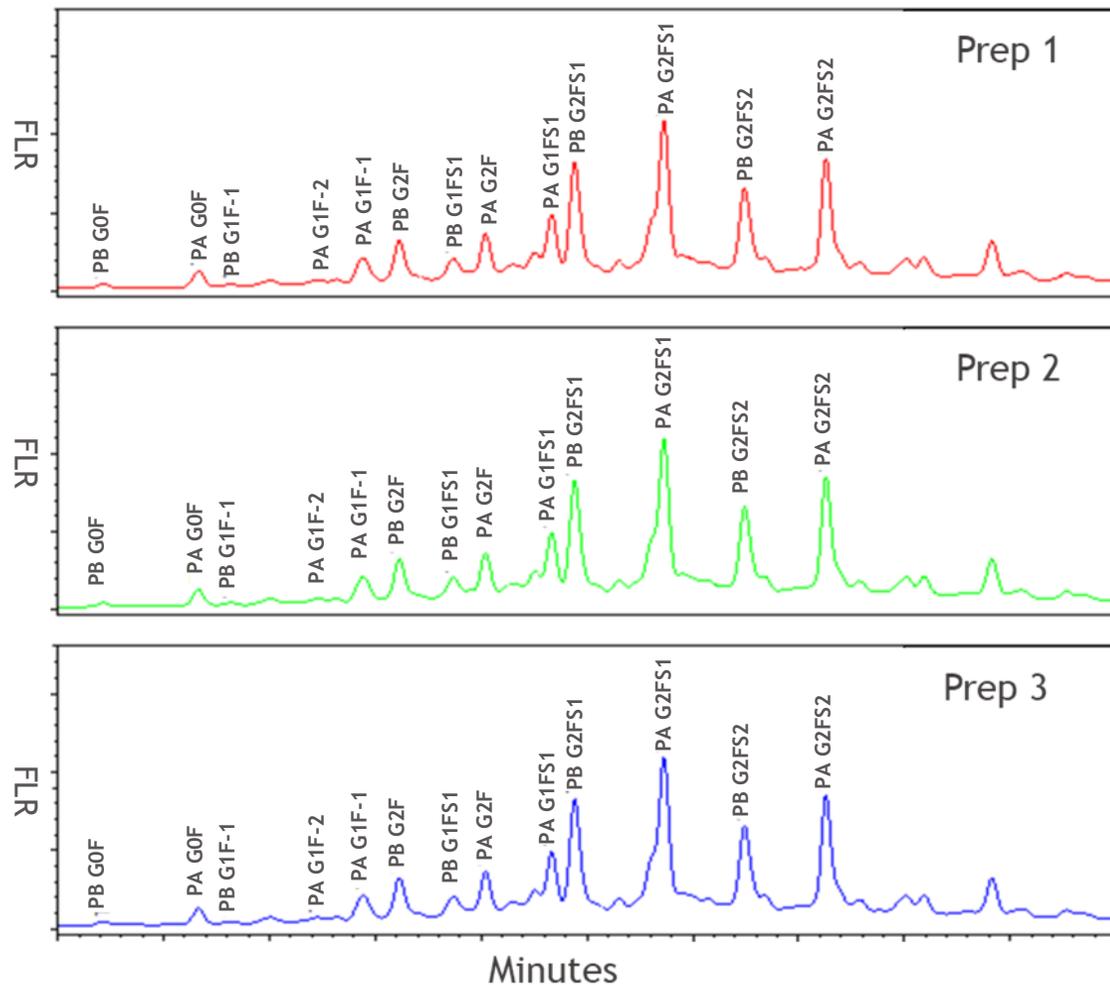
IAEDANS, Ammonium Formate



- ❑ Glycopeptide A + G0F and Glycopeptide B + G1F were separated
- ❑ Better resolution between Glycopeptide A+ G2FS1 and Glycopeptide B+ G2FS2

- Different glycoforms of Fab glycopeptide A and B are fully resolved with IAEDANS FLR tag;

Quantitation of Fab Site-Specific Glycopeptides through Fluorescence Peak Intensity



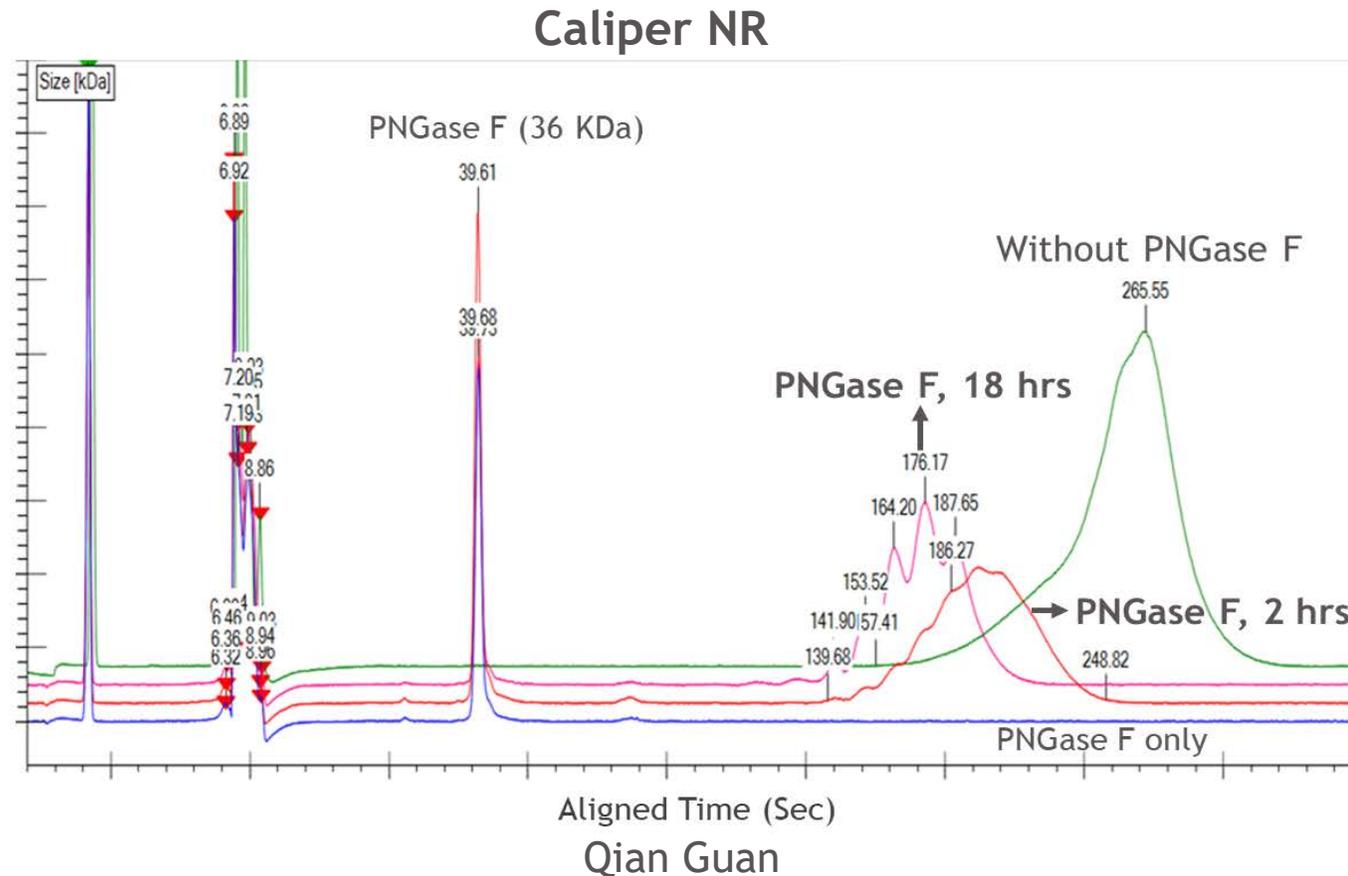
- Reproducible quantitation for glycopeptides;

Case Study I Conclusions

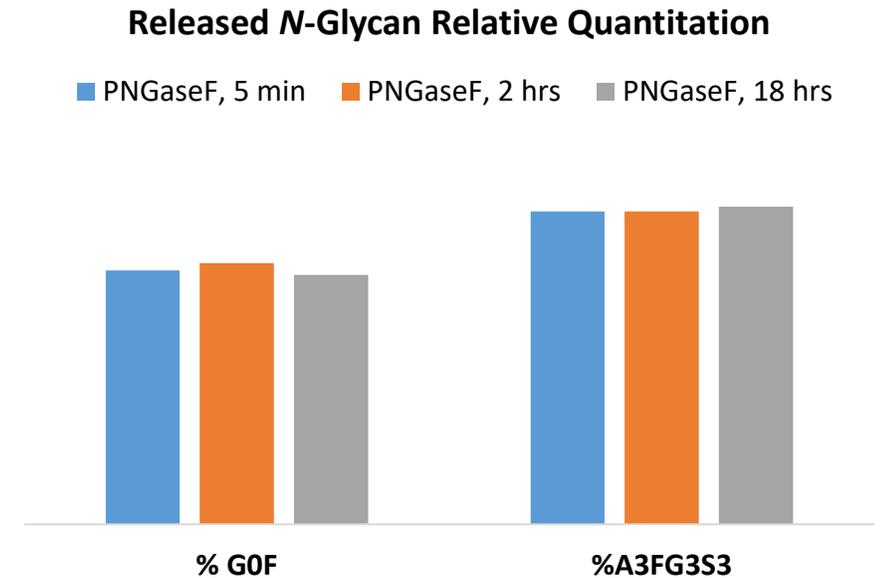
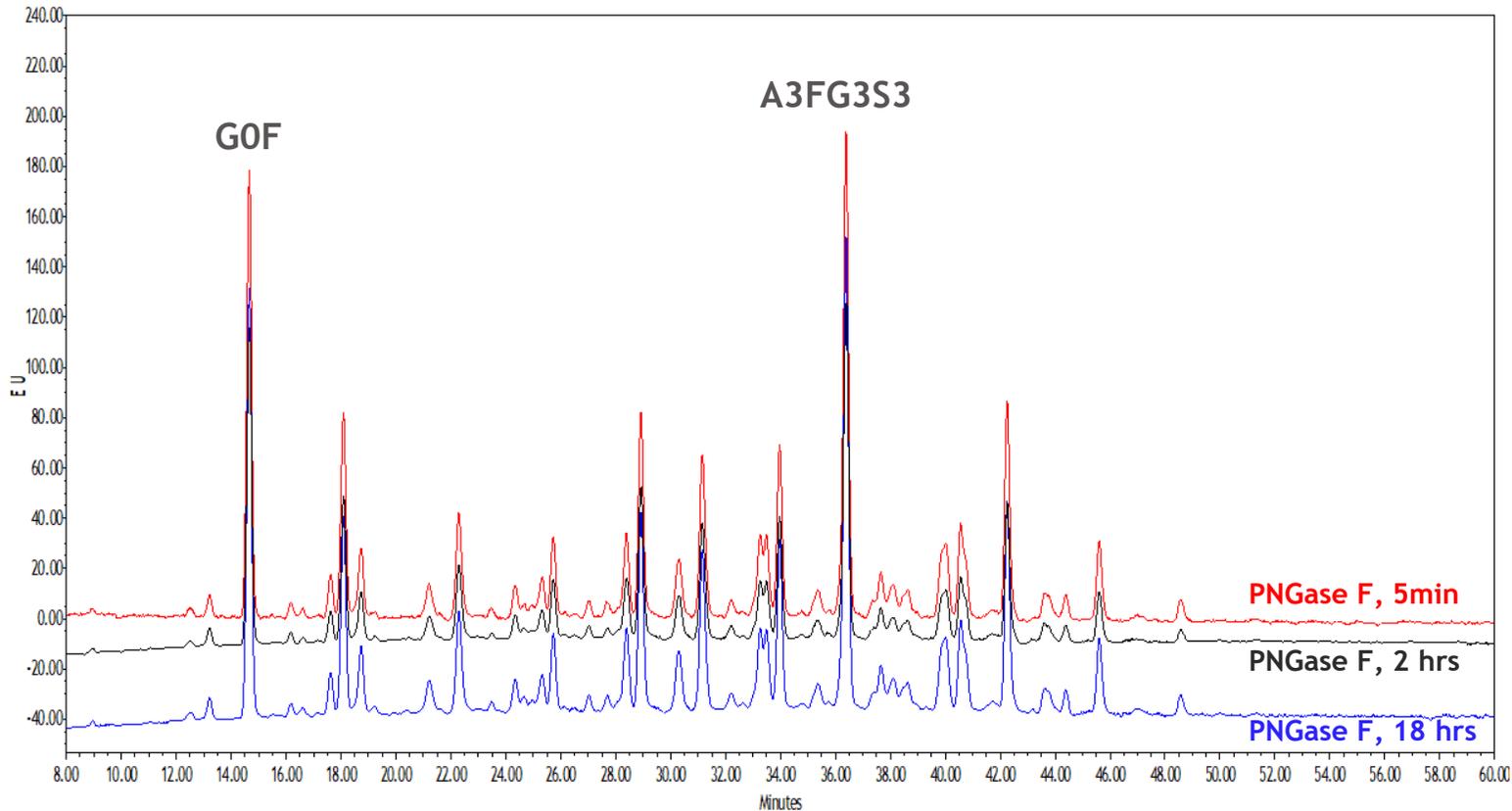
- A HILIC LC-FLR-MS method was established to separate glycopeptides and quantify Fab site-specific glycosylation for a Fc-fusion protein;
- Mobile phase and fluorescent tags played significant role in glycopeptide separation with HILIC column;

De-glycosylation by PNGase F with Native Conditions

- Peaks started to split but still broad, after 18-hour PNGase F digestion under native conditions (37° C);
 - Is the digestion complete?



Consistent *N*-glycan Profile with PNGase F Digestion under Native Conditions

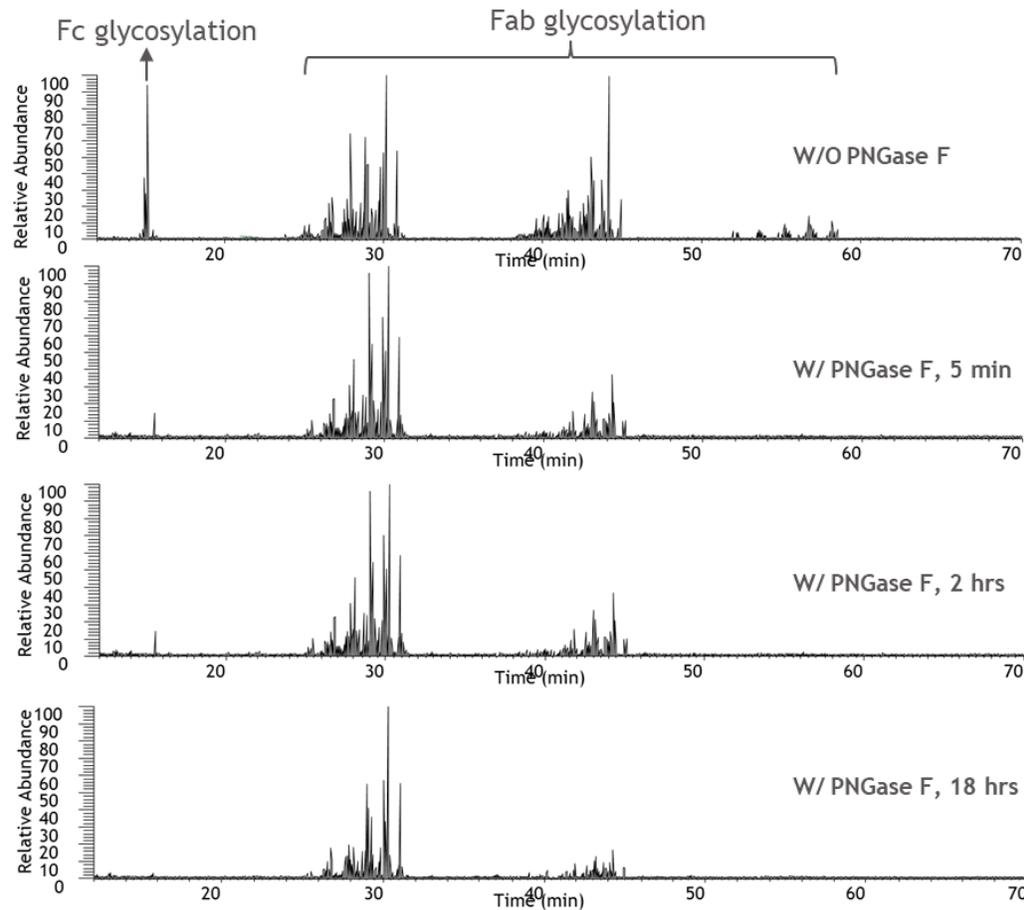


Xia Xu

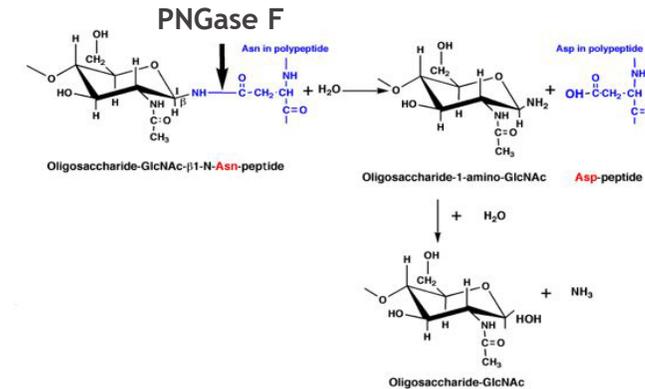
- Whether *N*-glycans from all the glycosylation sites were completely removed remains inclusive;

Glycopeptide Site-Occupancy Analysis by LC-MS

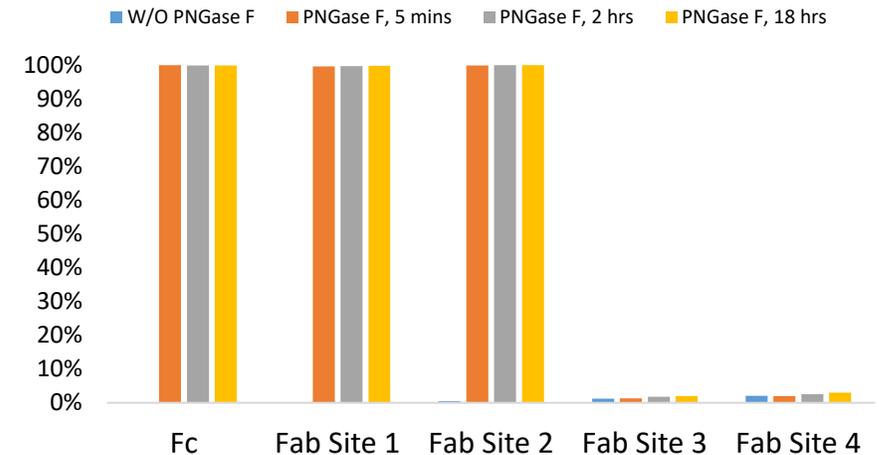
EIC of $m/z=204.0867$
Signature ions of glycopeptide fragmentation



- PNGase F cleaves between Asn and the innermost GlcNAc, transforming Asn to Asp;



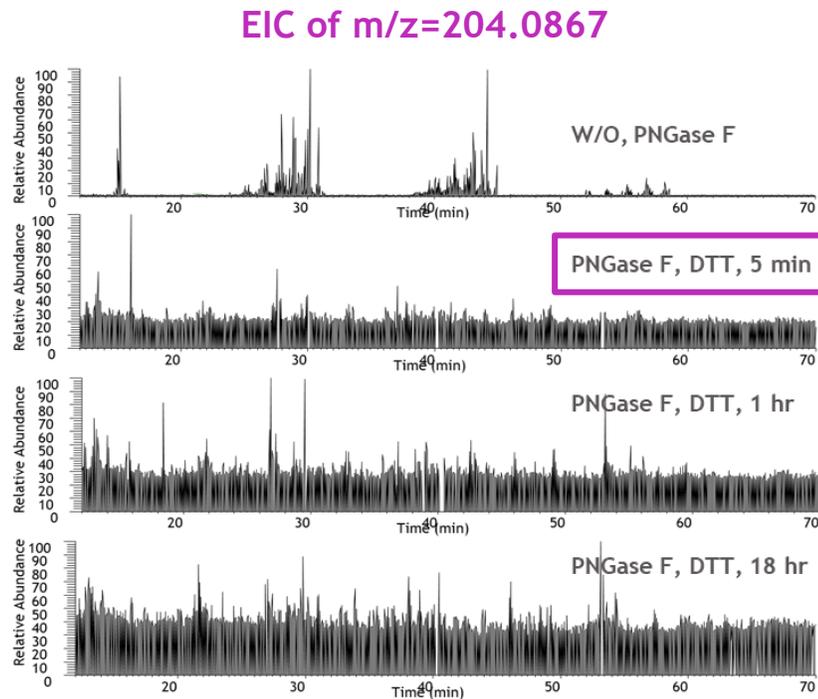
% Deamidation of Glycopeptides after PNGase F



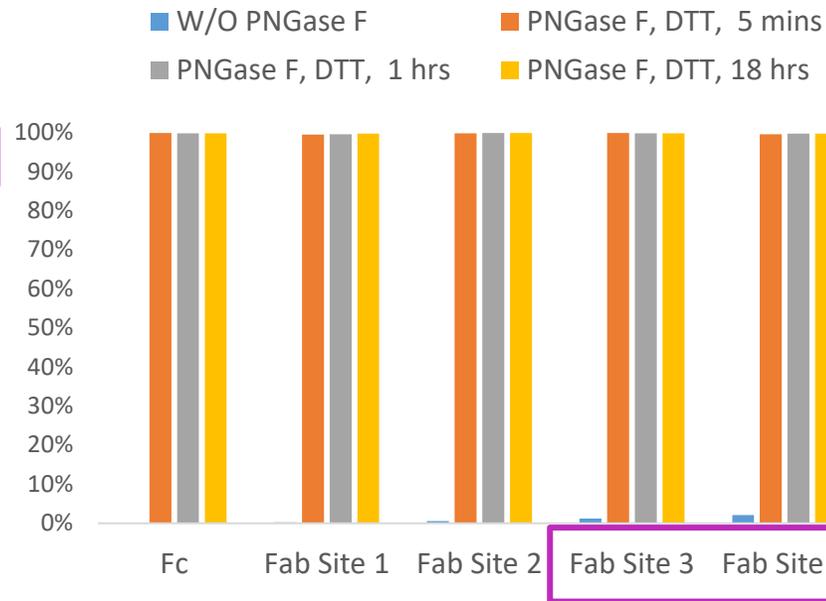
- Fab site-dependent preference by PNGase F under native conditions;

PNGase F Digestion under Reducing Conditions Completely Removes *N*-glycans

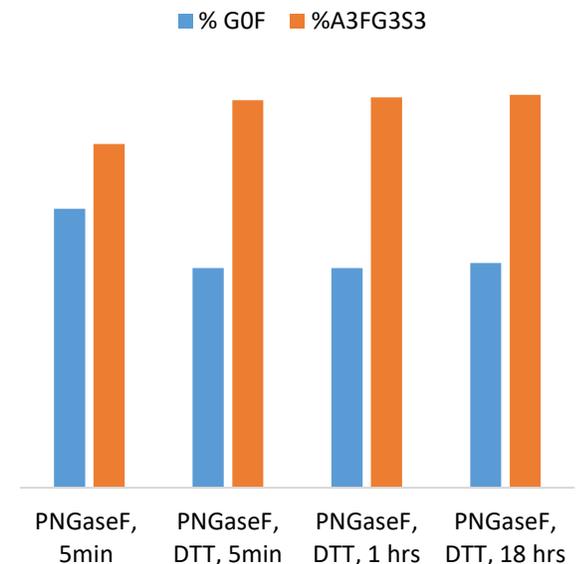
- Fab glycosylation site 3 and site 4 are surrounded by **disulfide bonds**, which could potentially block the enzyme access to the glycosylation sites;
- Therefore, PNGase F digestion under reducing conditions was evaluated;



% Deamidation of Glycopeptides after PNGase F



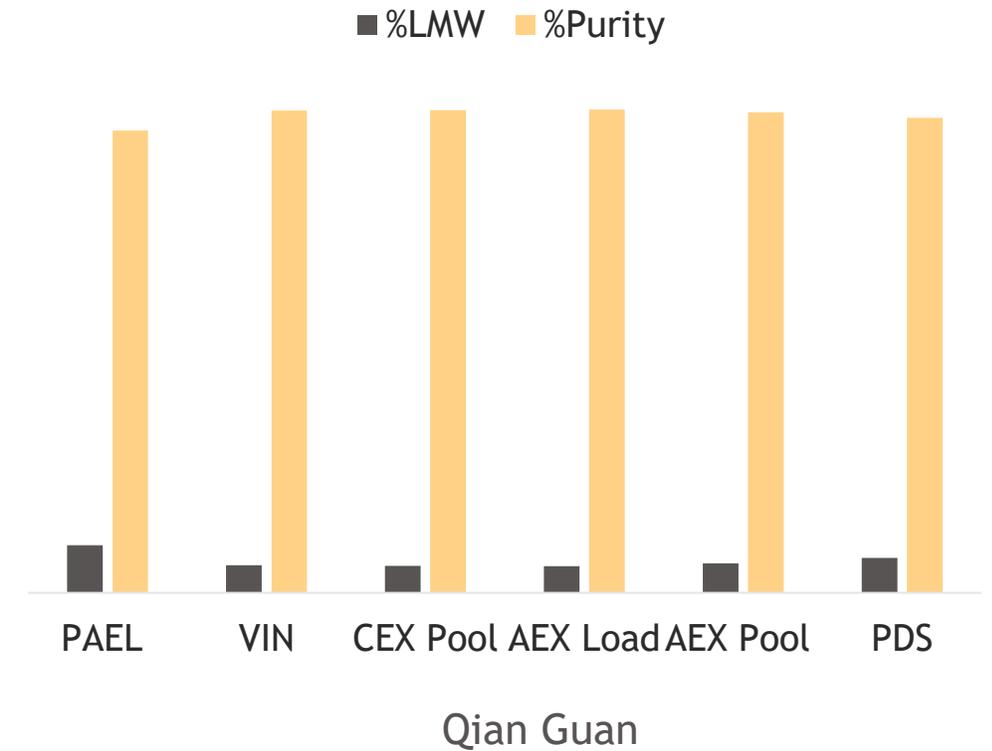
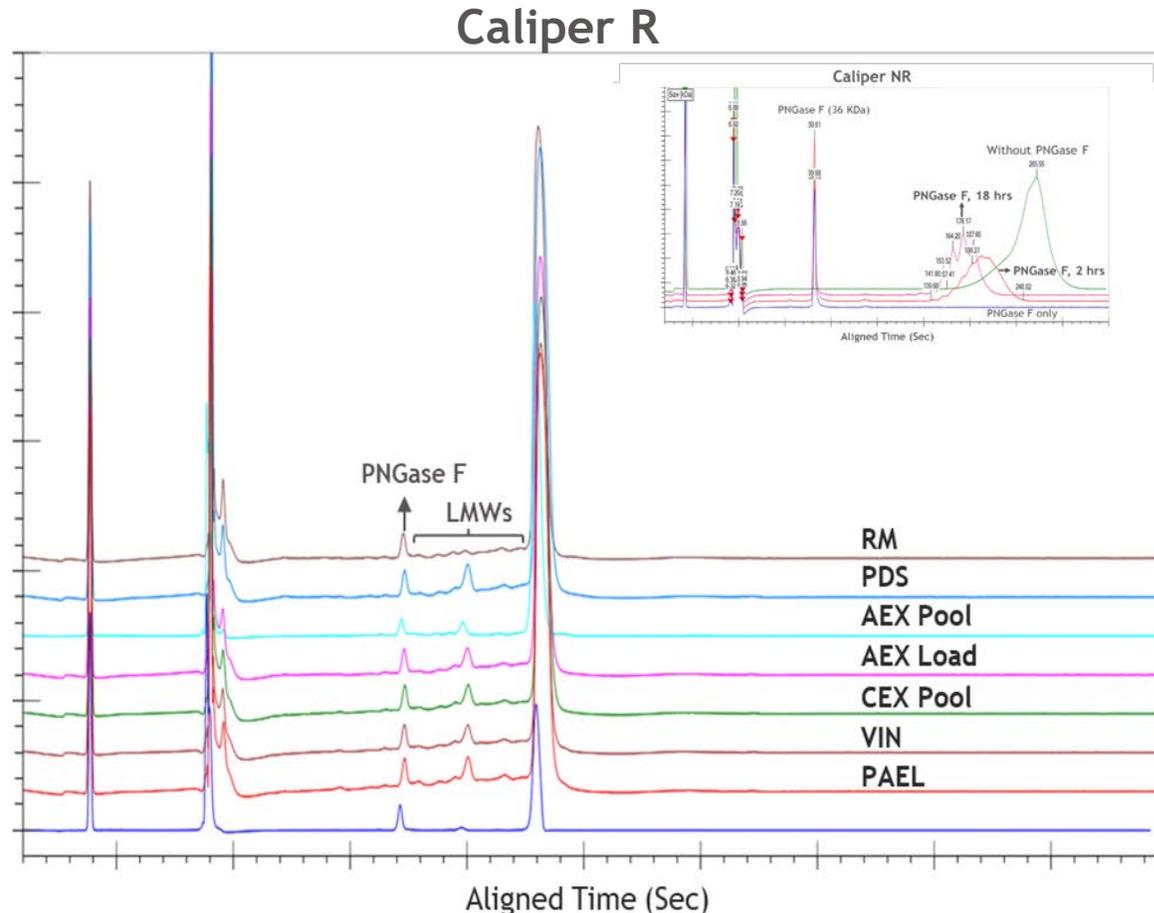
Released *N*-glycan Analysis



- **PNGase F digestion for 5min with DTT is sufficient to remove *N*-glycans from all the modification sites;**

Optimized Caliper (Reduced) Method for In-Process Sample Support

- PNGase F digestion protocol under reducing condition was applied to Caliper sample preparation;
- VIN step removed LMWs; As a result, % purity increased;

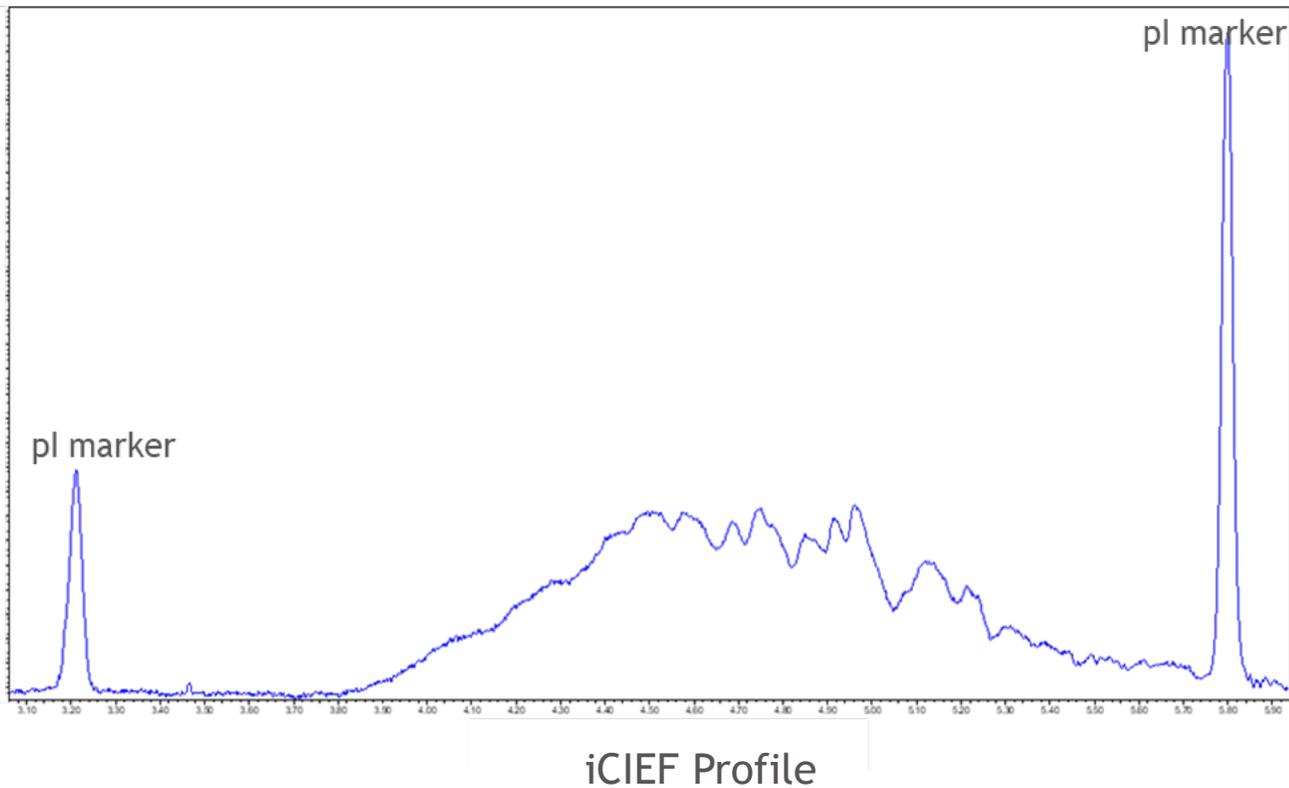


Case Study II Conclusions

- LMWs were not resolved from intact protein with Caliper for a Fc-fusion protein when *N*-glycans are present;
- PNGase F digestion under reducing condition was implemented during sample preparation to enable the purity analysis with Caliper for in-process samples;
- Site-specific glycopeptide analysis with LC-MS is a useful tool for establishing molecule specific method to analyze the sample purity;

Case Study III: Complex Charge Profile of Fc-fusion Protein

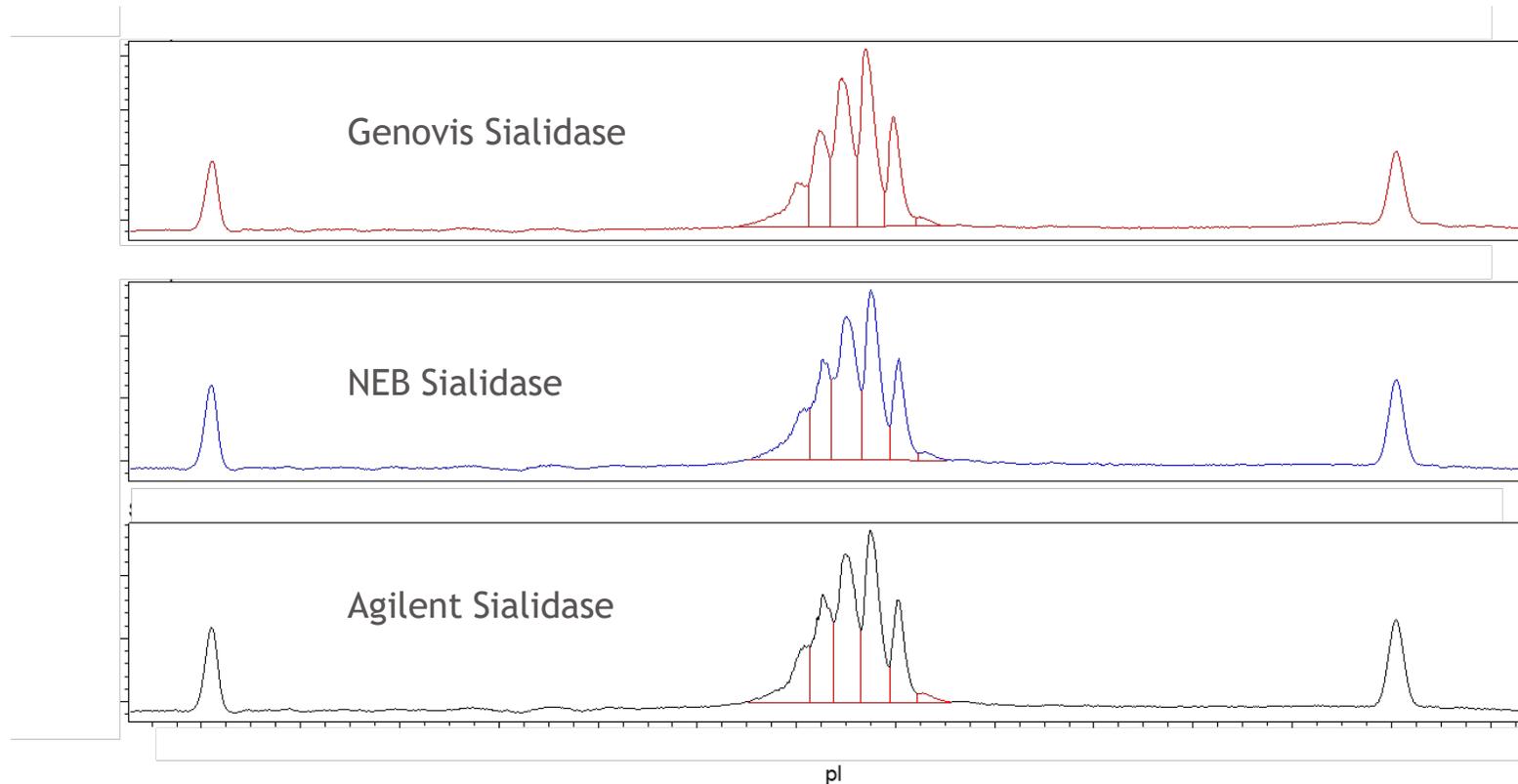
- A Fc-fusion protein with >10 *N*-glycosylation sites;
- Complex charge profile due to sialic acid modification;



- ❑ Difficult for peak integration;
- ❑ Challenging for results reporting;
- ❑ Limited information for process;

Simplified Charge Profile by Sialidase Treatment

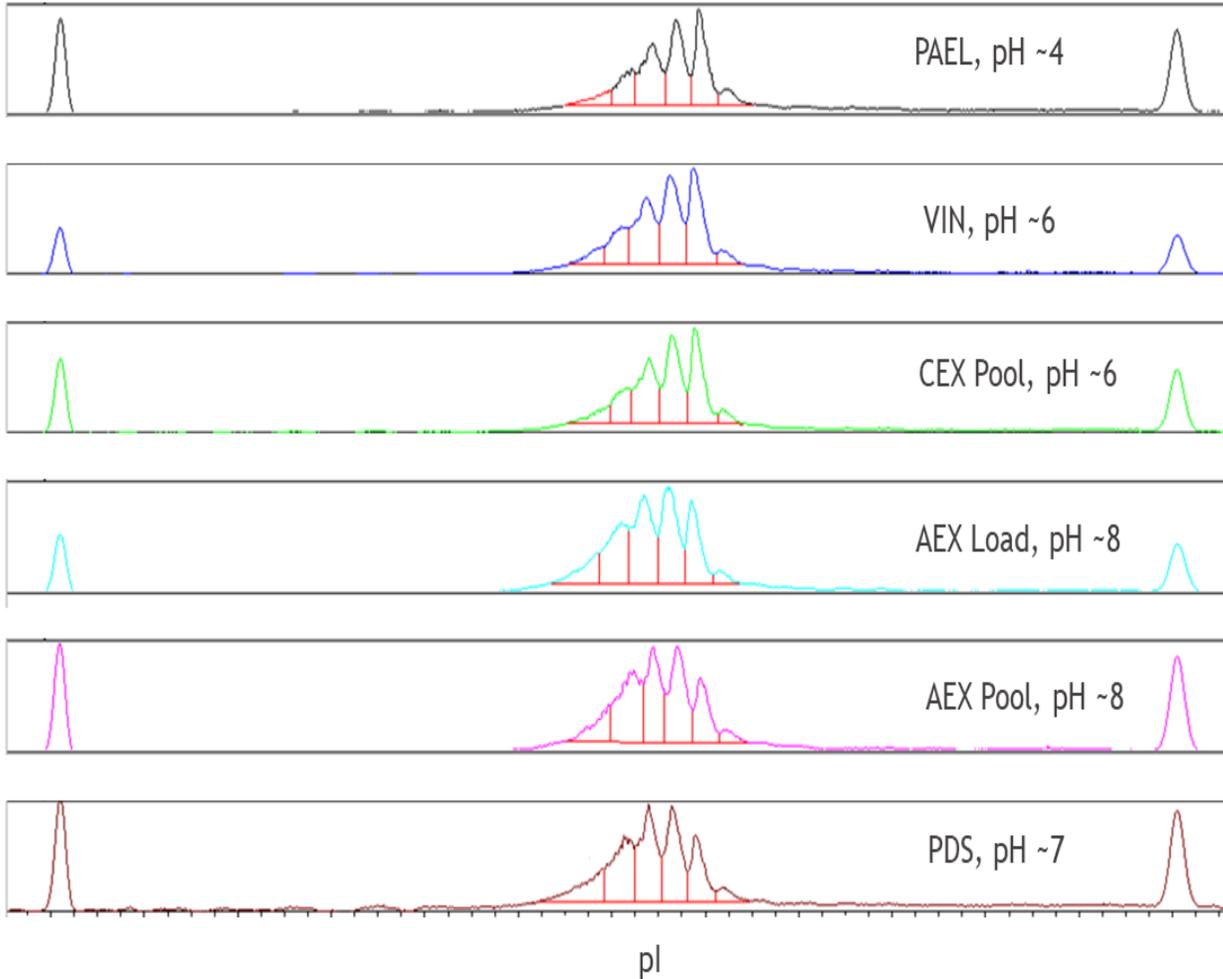
- In-process sample testing strategy
 - Reduce the charge heterogeneity by sialidase treatment;
 - Desialyated iCIEF is for monitoring other attributes that could impact the charge profile;
 - Sialic acid can be monitored by SA quantitation method, *N*-glycan method, and/or glycopeptides method;



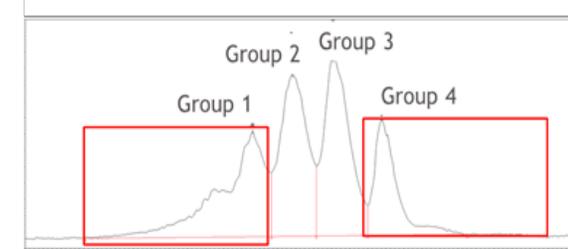
- ❑ 30 mins Sialidase digestion;
- ❑ Comparable results for three vendors' sialidase;

Helen Zhao, Li Zhang, Joann Max

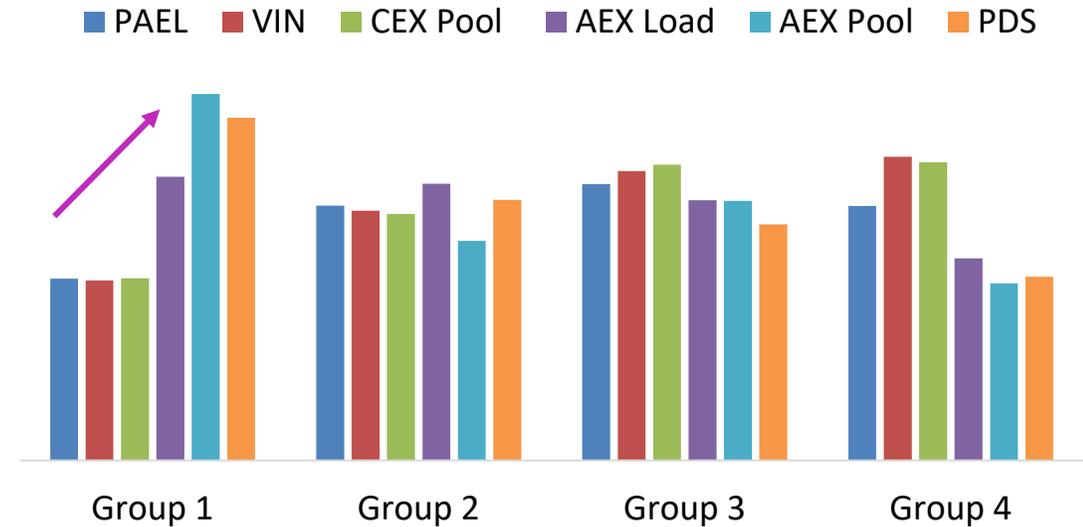
iCIEF Profile for Downstream Steps



Li Zhang, Helen Zhao



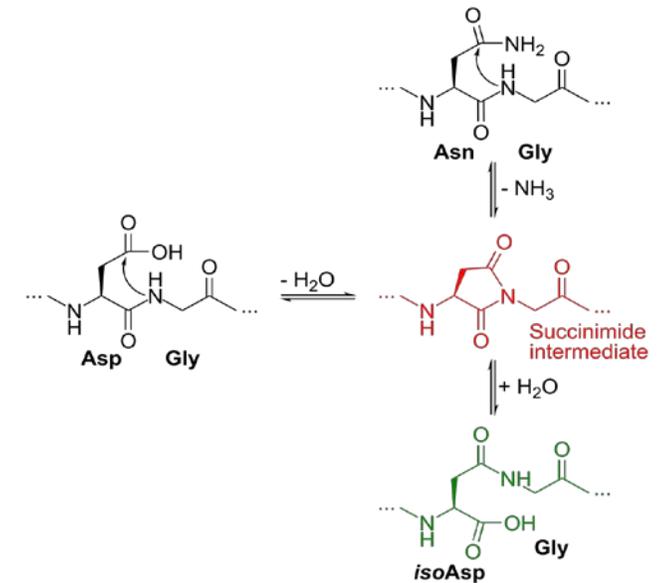
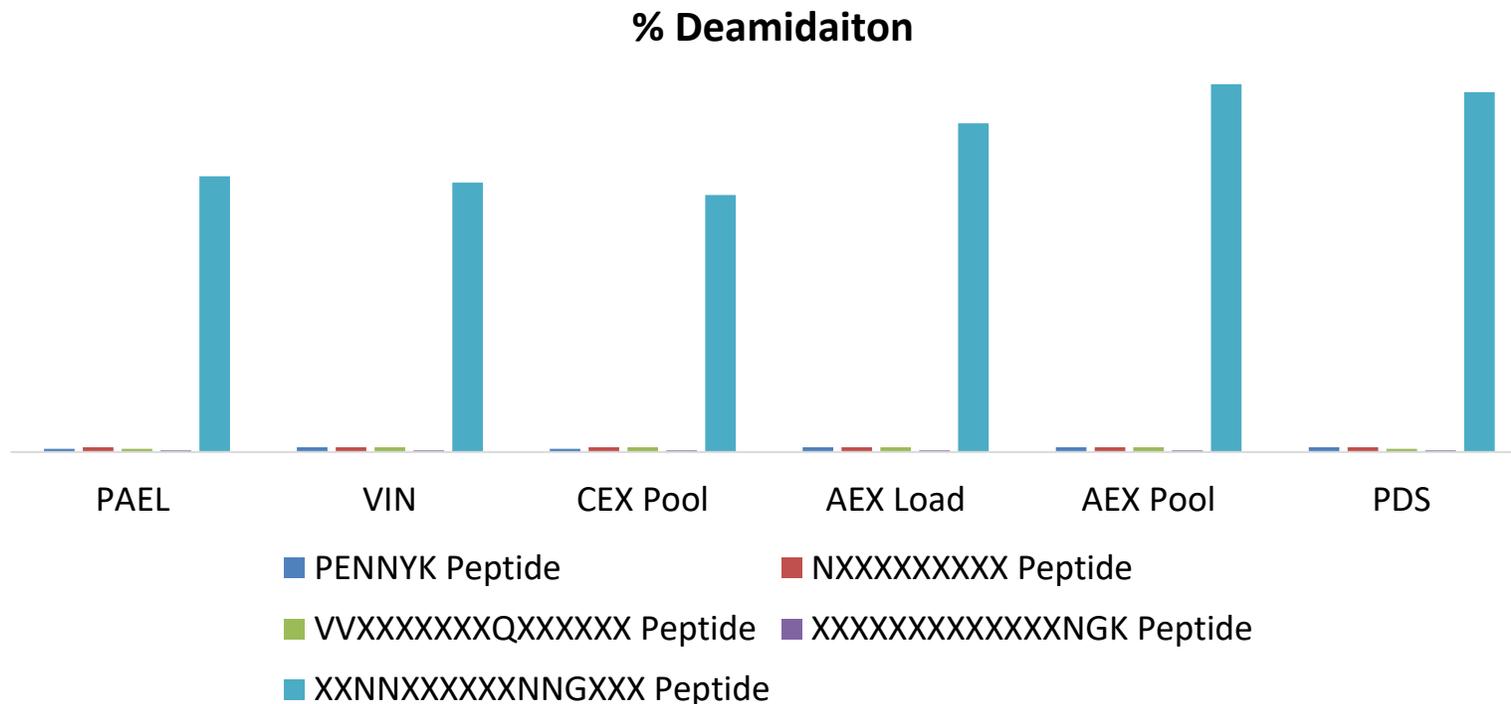
iCIEF Grouped Peak Intensity (%)



- % Group 1 increases starting from AEX load

Deamidation by LC-MS Peptide Mapping

- CEX pool and AEX load sample pH is different: pH ~6 vs pH ~8;
- Deamidation is checked by peptide mapping because it's known as acidic variant and can be triggered under basic pH;
- XXNNXXXXXXXXNNGXXX peptide deamidation increase during downstream process;



<https://en.wikipedia.org/wiki/Deamidation>

Case Study III Conclusions

- Complex iCIEF profile due to terminus sialic acid;
- Simplified iCIEF profile after sialidase treatment is useful to monitor other attributes (e.g. deamidation) that could impact the charge profile;
- Peptide mapping suggested the deamidation of peptide XXNNXXXXXXXXNNGXXX could potentially contribute to the increase in iCIEF acidic region during downstream steps;

Summary

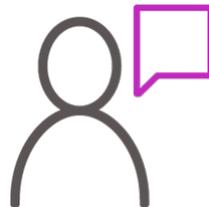
- The analysis of Fc-fusion proteins is challenging;
- Molecule specific methods are often required to support the process development of complex protein therapeutics;
- Mass spectrometry is a critical analytical technique to assist the development of non-platform methods for process development support;

Acknowledgement

Partnership among Process Development Analytics, Upstream, and Downstream

- Qian Guan
- Xia Xu
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- Kyle McHugh
- Lye Lin Lock
- Hangtian Song
- Diane Worrell
- Keith Coleman
- Li Tao
- Yunping Huang
- Junyu Ma
- Mengxiao Lu

Thank You for Your Attention!





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