

# **Binding Sites, Orders, Site-Specific Affinities, and Composite Allosteric Behavior Revealed by a Single Mass Spectrometry-Based Approach**

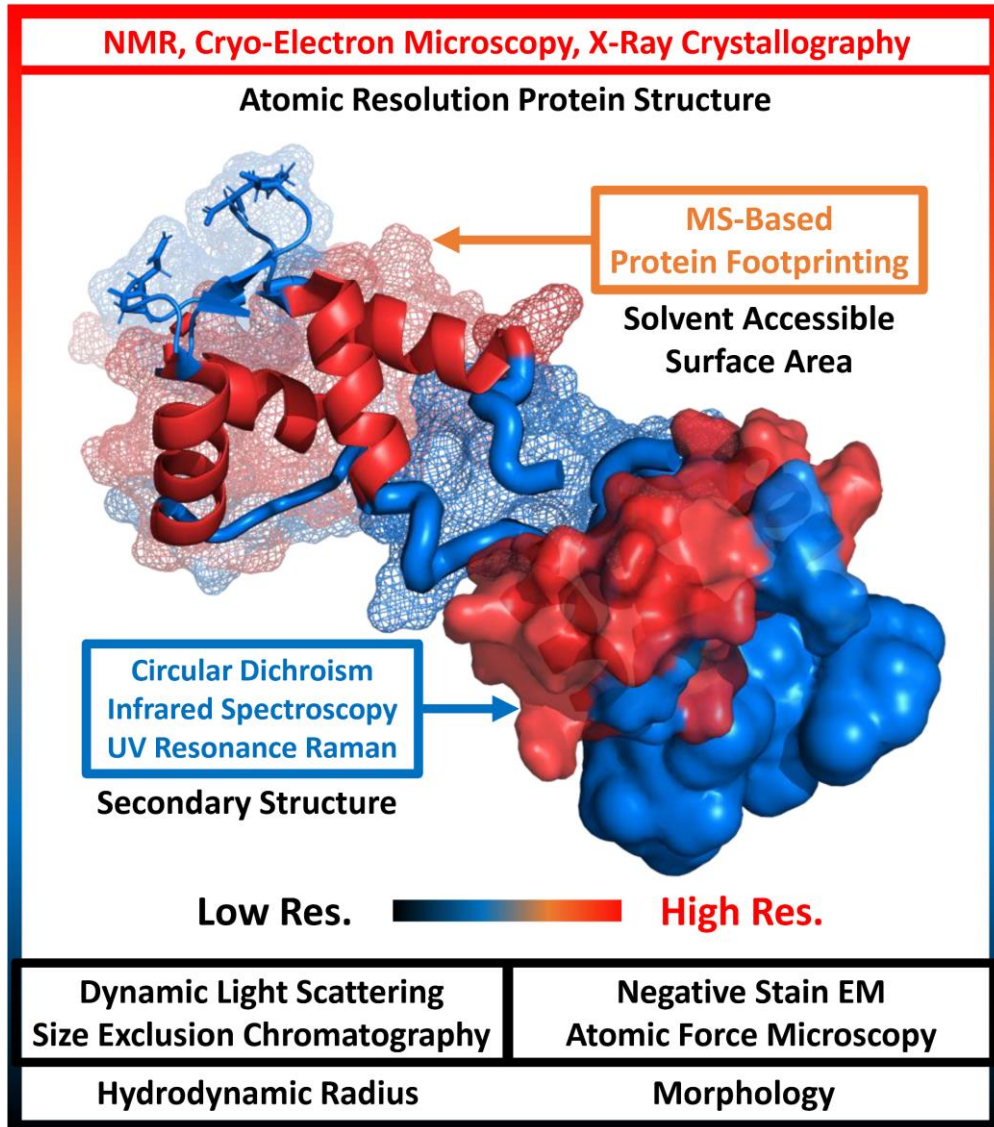
**Roger Liu**

Global Product Development  Bristol Myers Squibb™

04/14/2021



# Mass Spectrometry-based Footprinting in Structural Proteomics

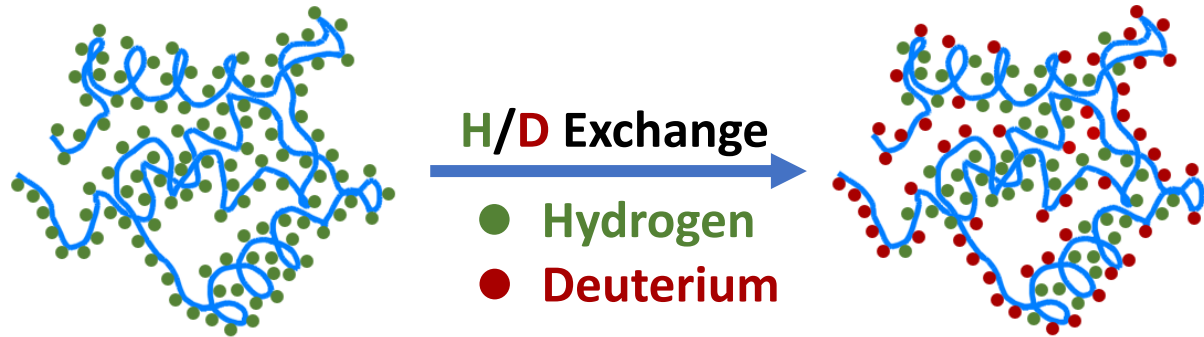


- **Protein Higher Order Structure (HOS)**
  - Secondary to quaternary structures
  - Enables various biological functions
- **Mass Spectrometry in HOS Analysis**
  - High throughput
  - Low sample amount requirement (ng -  $\mu$ g)
  - In-solution characterization
  - High sensitivity
  - Mid to high spatial resolution (peptide and amino acid residue level)



# Reversible and Irreversible Approaches – Solvent Accessible Surface

- Reversible Approaches



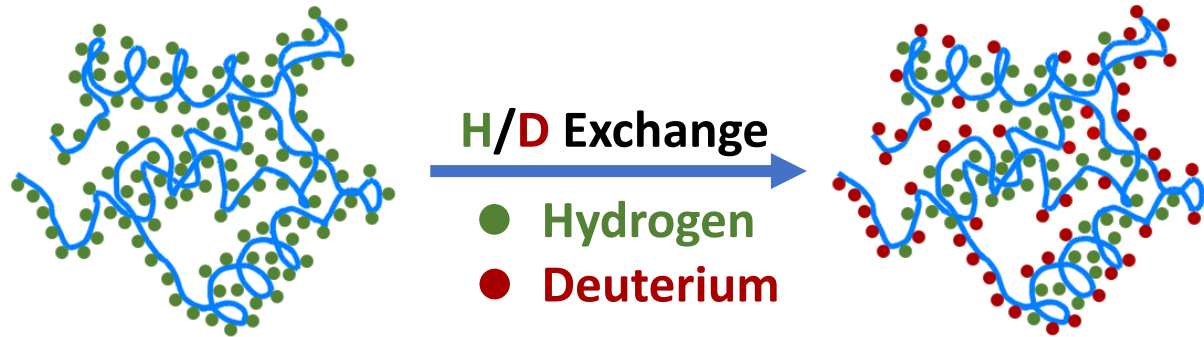
- Hydrogen Deuterium Exchange (HDX)

Solvent accessible backbone amide **hydrogen** reversibly exchange with **deuterium** in solvent



# Reversible and Irreversible Approaches – Solvent Accessible Surface

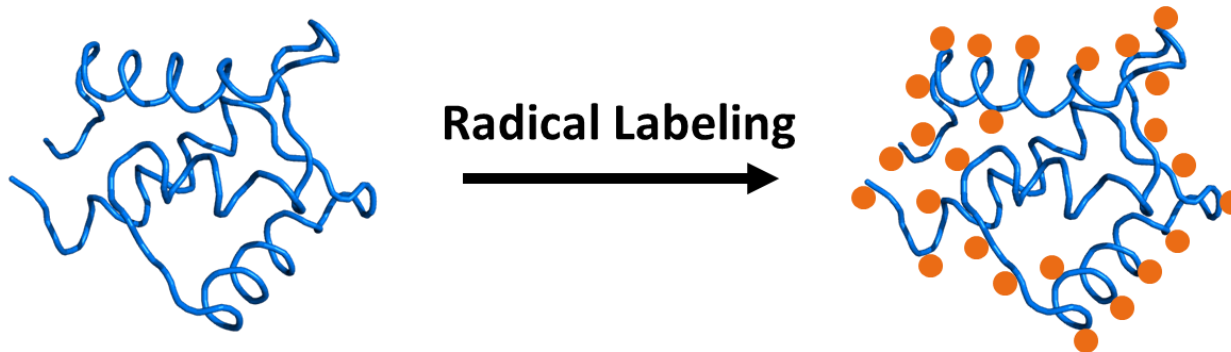
- Reversible Approaches



- Hydrogen Deuterium Exchange (HDX)

Solvent accessible backbone amide **hydrogen** reversibly exchange with **deuterium** in solvent

- Irreversible Approaches



- Radical labeling reagents

e.g.,  $\cdot\text{OH}$ ,  $\text{SO}_4^{\cdot-}$ ,  $\cdot\text{CF}_3$ ,  $\text{CO}_3^{\cdot-}$ ,  $\text{I}^\cdot$

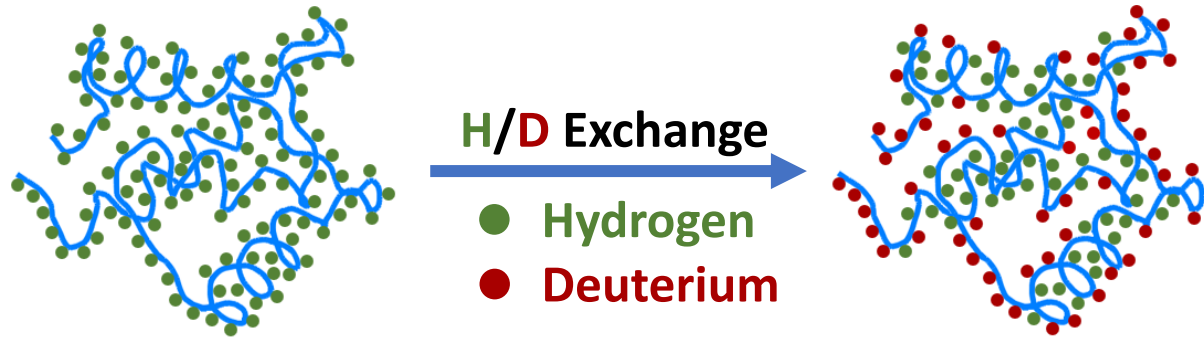
RC:, photo cross-linking

Chemical labels covalently attach onto solvent accessible amino acid side chains



# Reversible and Irreversible Approaches – Solvent Accessible Surface

- Reversible Approaches



- Hydrogen Deuterium Exchange (HDX)

Solvent accessible backbone amide **hydrogen** reversibly exchange with **deuterium** in solvent

- Irreversible Approaches



- Radical labeling reagents  
e.g.,  $\cdot\text{OH}$ ,  $\text{SO}_4^{\cdot-}$ ,  $\cdot\text{CF}_3$ ,  $\text{CO}_3^{\cdot-}$ ,  $\text{I}^\cdot$   
RC:, photo cross-linking
- Targeted labeling reagents (covalent labeling)  
e.g., glycine ethyl ester: Asp & Glu  
Chemical cross-linking

Chemical labels covalently attach onto solvent accessible amino acid side chains



# Measuring P-L Binding Affinity

$$K_a = \frac{k_{on}}{k_{off}} = \frac{[PL]}{[P][L]}$$

$$K_d = \frac{k_{off}}{k_{on}} = \frac{[P][L]}{[PL]}$$

- **Concentration Measurement**

Fluorescence Polarization, Circular Dichroism, NMR, FT-IR

- **Rate Constant Measurement**

Surface Plasmon Resonance

- **Thermodynamic Measurement**

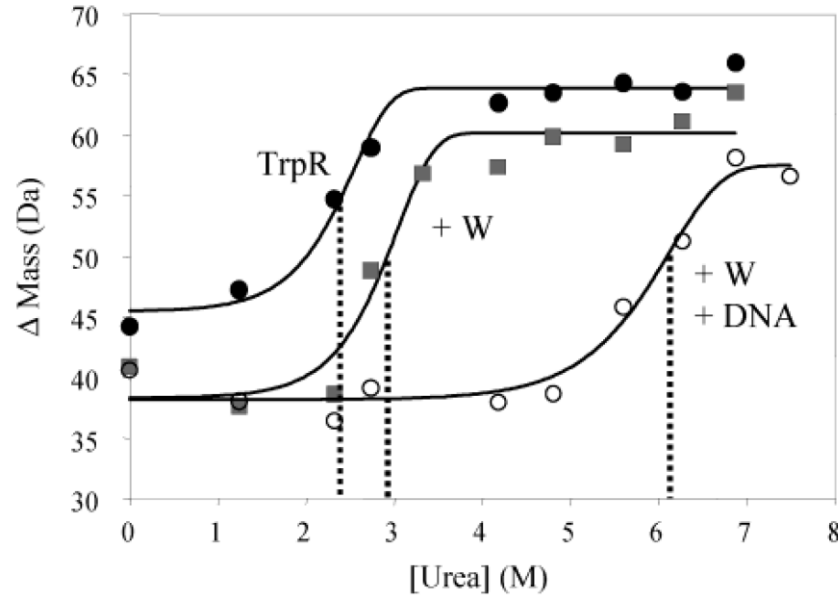
Isothermal Titration Calorimetry (Heat flow during titration)

Specific sample preparation  
Limited spatial resolution

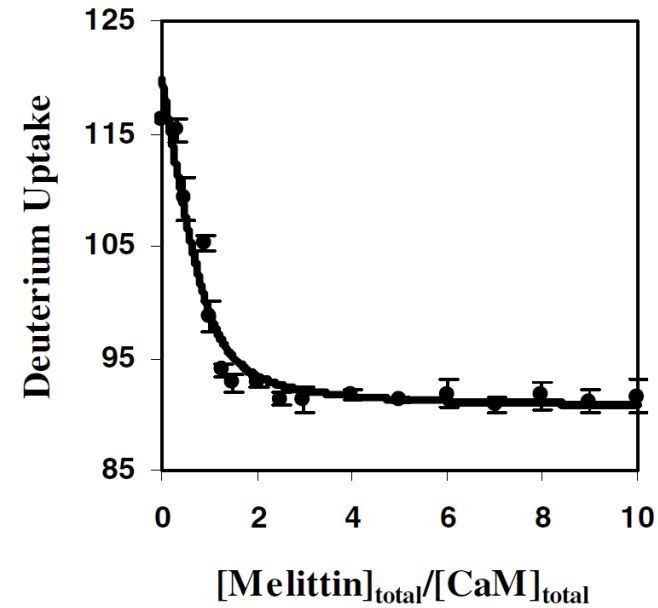
**How could Mass Spectrometry contribute?**



# SUPREX and PLIMSTEX



Stability of Unpurified Proteins from Rates of H/D Exchange

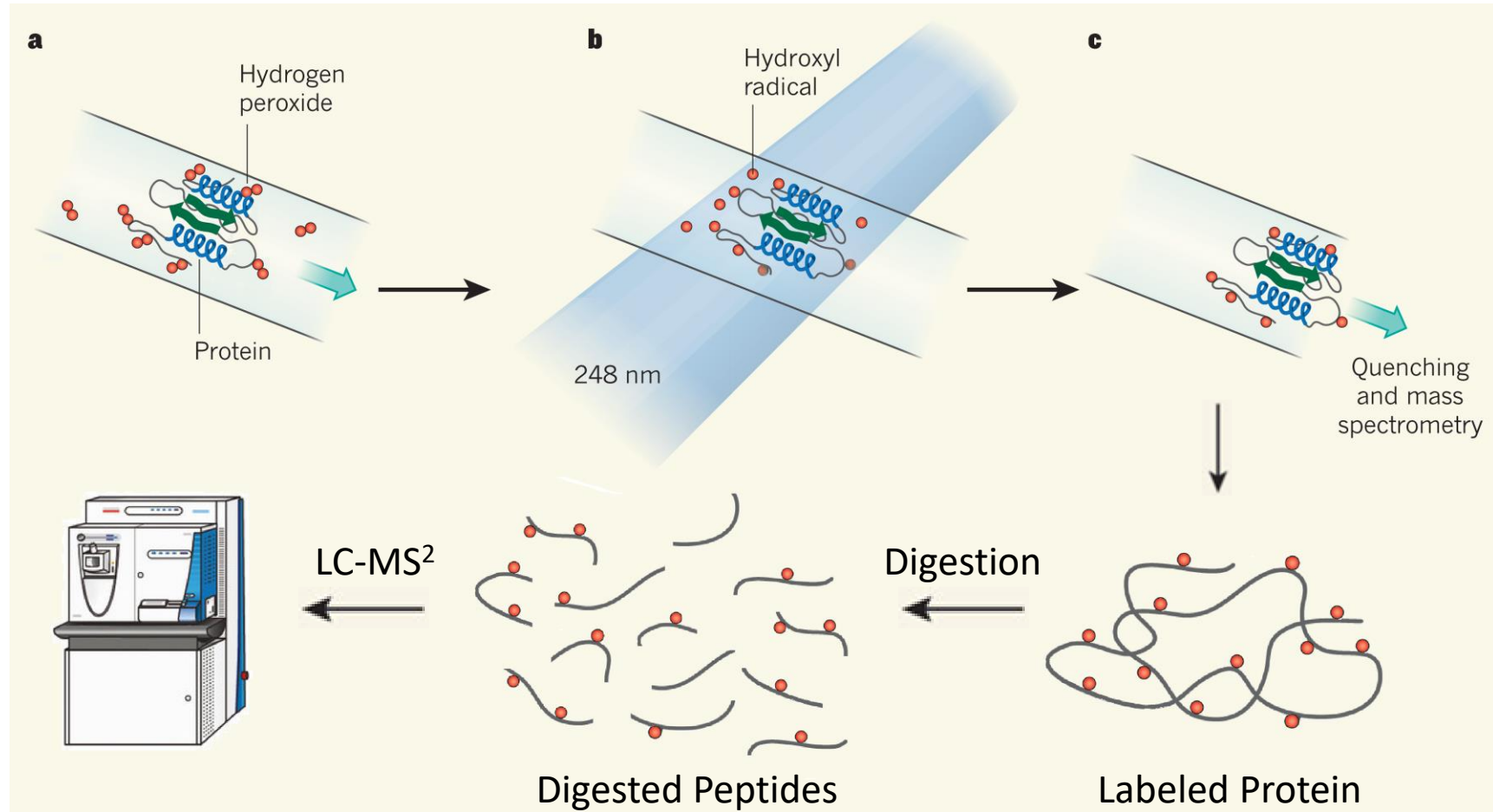


Protein-Ligand Interactions by Mass Spectrometry, Titration, and H/D Exchange

- Universal to various systems, low sample quantities, no special labeling, site-specific
- Backexchange, complicated picture by ligand off-rate and long H/D exchange time



# Fast Photochemical Oxidation of Proteins (FPOP) & Mass Spectrometry



[1] Gruebele, M. *Nature* **2010**, 468, 640-641.

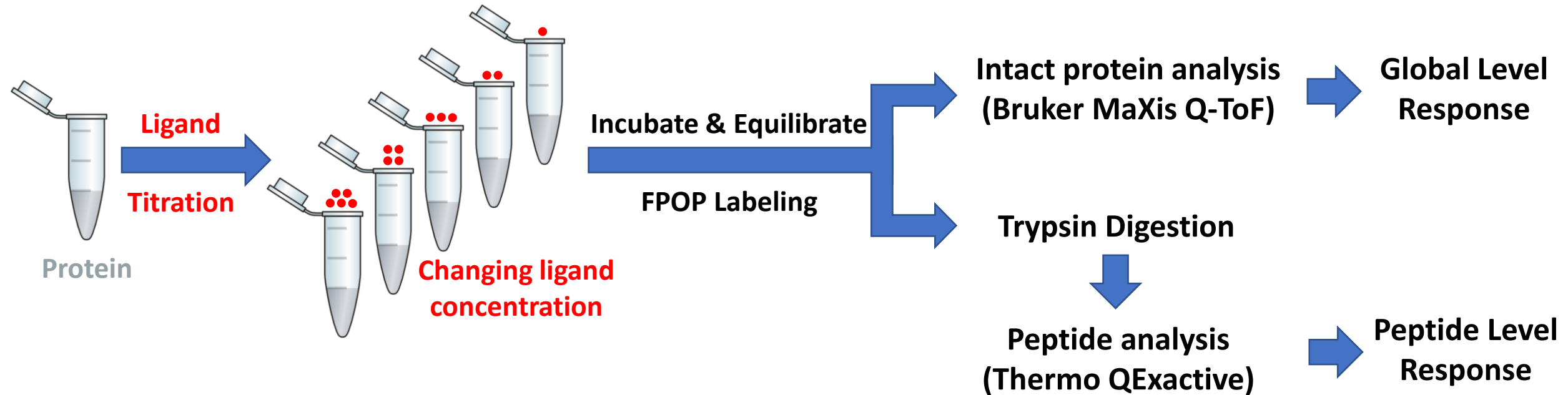
[2] Hambly, D. M.; Gross, M. L. *J. Am. Soc. Mass Spectrom.* **2005**, 16, 2057-2063.

[3] [Liu, X. R.](#)<sup>†</sup>; Zhang, M. M.<sup>†</sup>; Gross, M. L. *Chem. Rev.* **2020**, 120, 4355-4454.

[4] [Liu, X. R.](#); Rempel, D. L.; Gross, M. L. *Nat. Protoc.* **2020**, 15, 3942-3970.



# Protein-Ligand Interaction by Ligand Titration, Fast Photochemical Oxidation of Proteins and Mass Spectrometry: **LITPOMS**

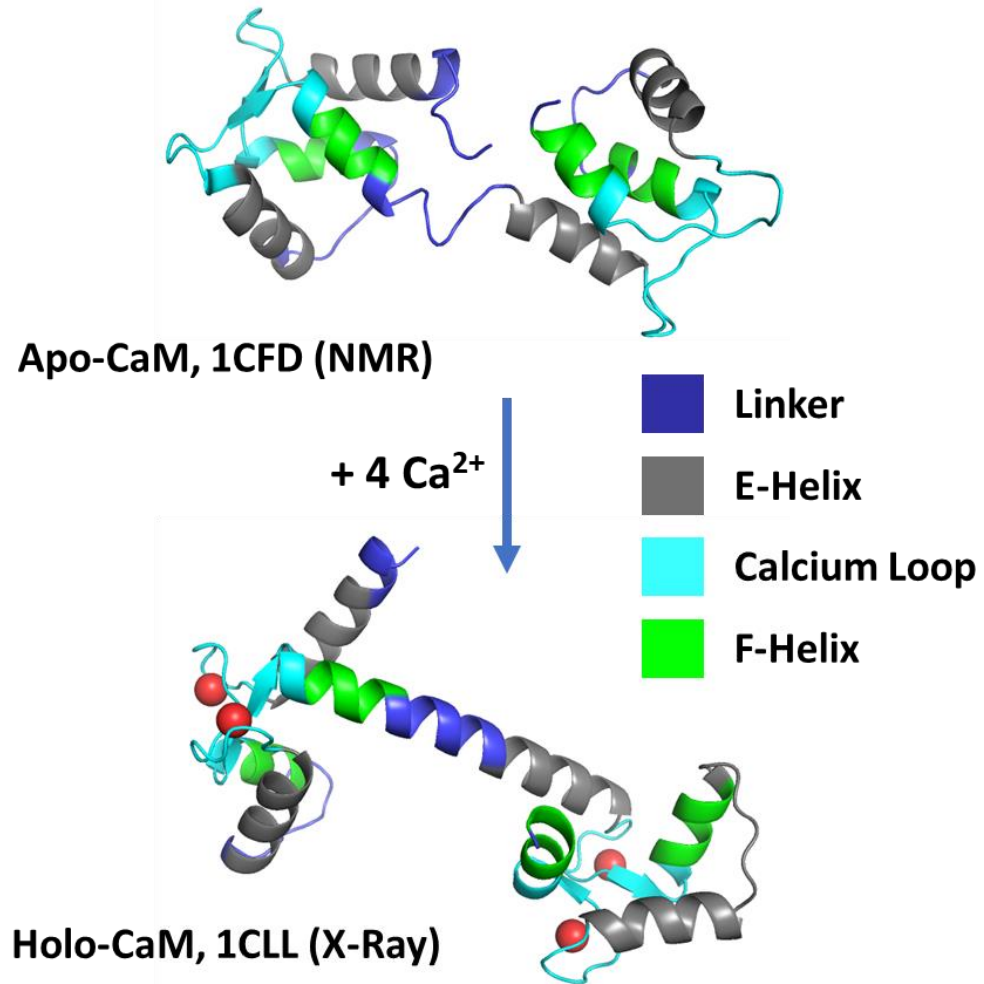


[1] **Liu, X. R.**; Zhang, M. M.; Rempel, D. L.; Gross, M. L. *J. Am. Soc. Mass Spectrom.* **2019**, 30, 213-217.

[2] **Liu, X. R.**; Zhang, M. M.; Rempel, D. L.; Gross, M. L. *Anal. Chem.* **2019**, 91, 5508-5512.



# Calmodulin & Calcium in Tris – 1:4 System



- Found in all eukaryotic cells
- Major Ca<sup>2+</sup> signaling pathway, regulating multiple intracellular process: cell growth, proliferation, apoptosis
- Four EF-hands with “helix-loop-helix” configuration
- Ca<sup>2+</sup> binds to calcium loop through chelating with negatively charged residues
- Binding affinity of  $\mu\text{M}$ , sensitive to stimulations
- Hint of cooperativity / allosteric behavior during binding

[1] Koniwa, H.; Tjandra, N.; Grzesiek, S.; Ren, H.; Klee, C. B.; Bax, A. *Nat. Chem. Biol.* **1995**, 2, 768-776.

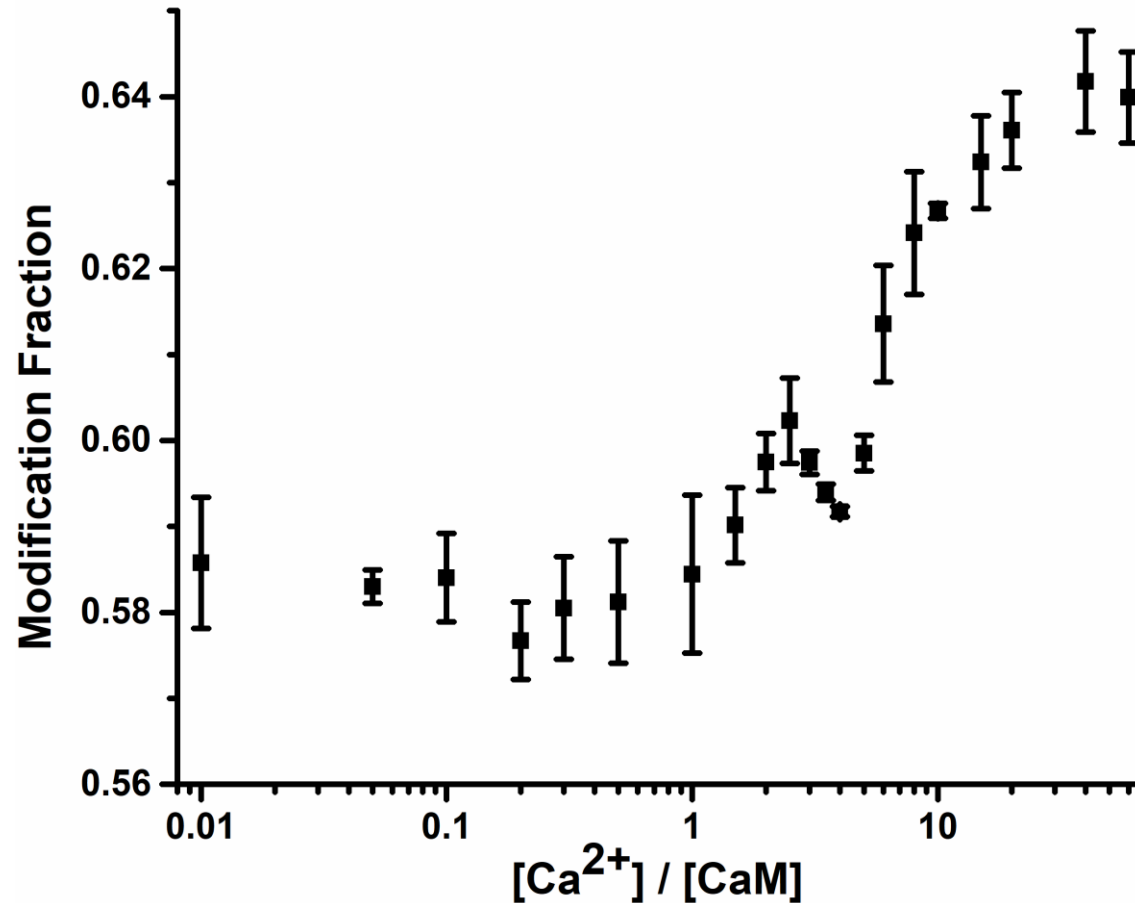
[2] Chattopadhyaya, R.; Meador, W.E.; Means, A.R.; Quiocho, F.A. *J. Mol. Biol.* **1992**, 228, 1177-1192.

[3] Linse, S.; Helmersson, A.; Forsen, S. *J. Biol. Chem.* **1991**, 266, 8050-8054.

[4] Sorensen, R.; Shea, M. A. *Biochemistry* **1998**, 37, 4244-4253.



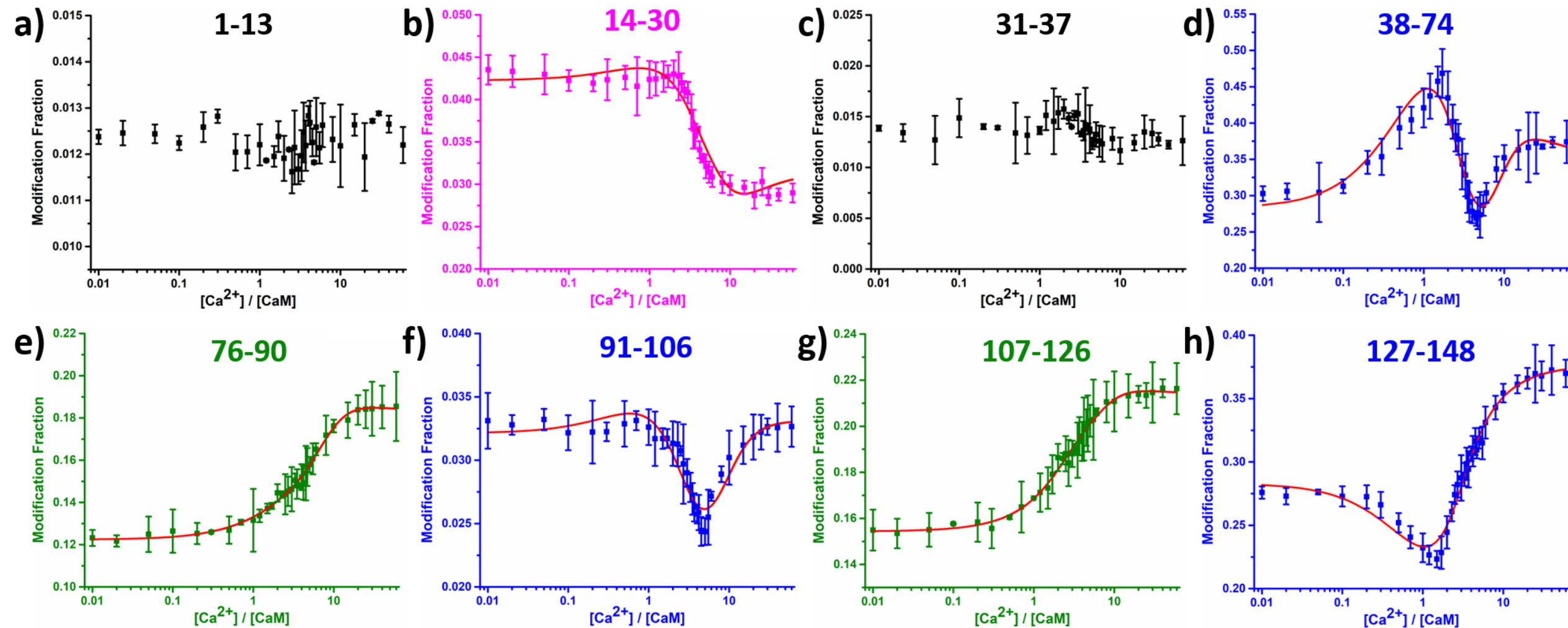
# LITPOMS of $\text{Ca}^{2+}$ & CaM at Global Level



- A more extended confirmation upon binding with calcium
- A decrease in modification fraction indicates a potential binding event
- A promising preliminary check before peptide & residue-level analysis

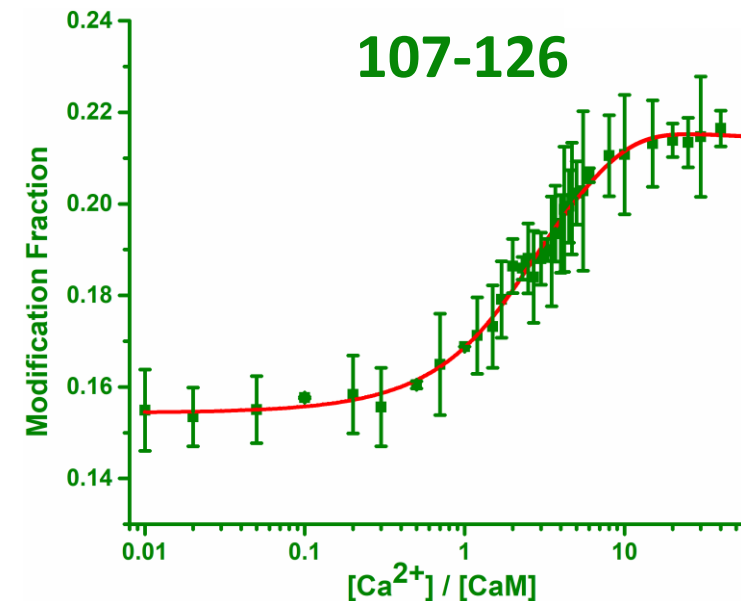
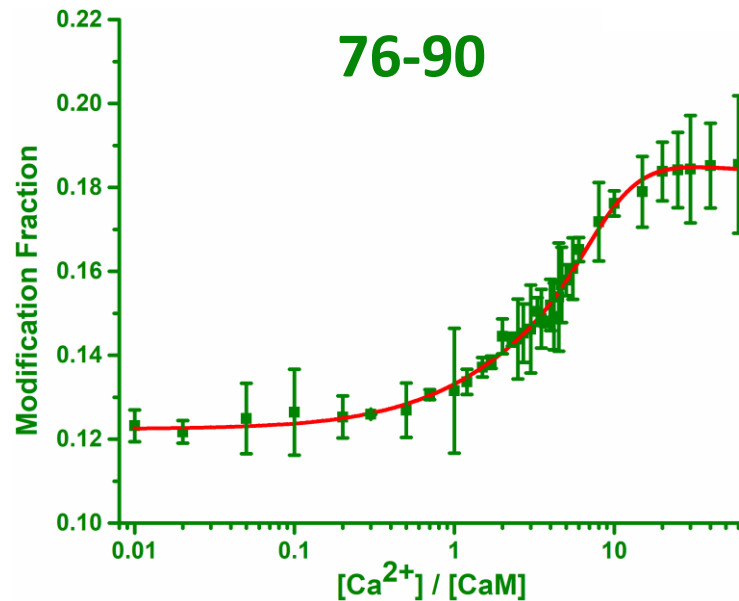


# LITPOMS of $\text{Ca}^{2+}$ & CaM at Peptide Level





# Class I Behavior – Loss of Protection

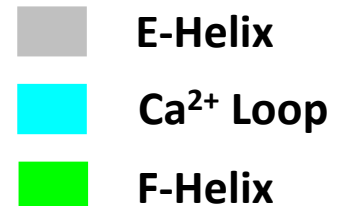
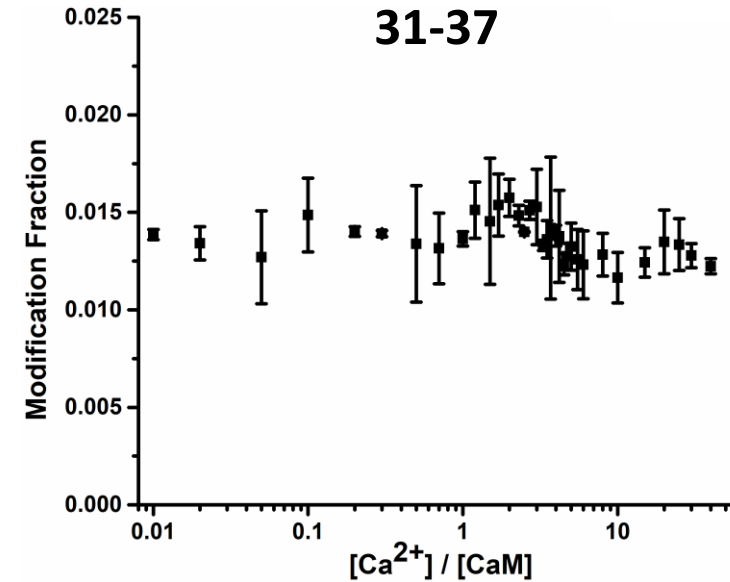
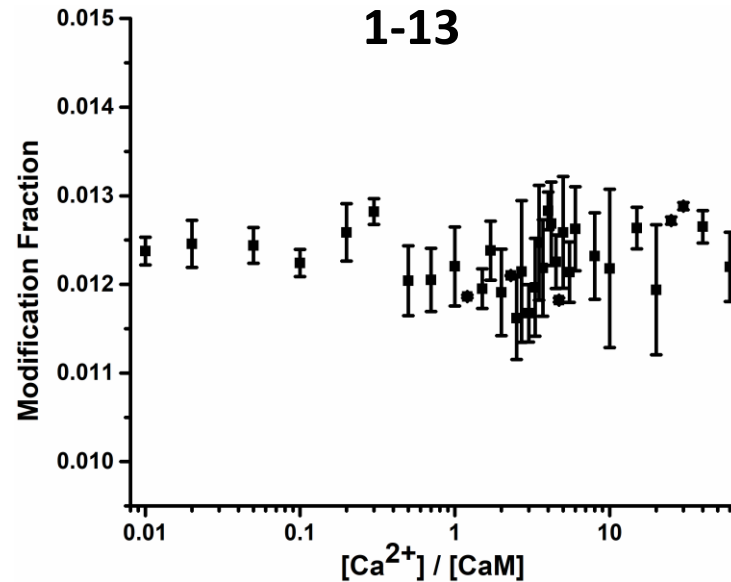


■ E-Helix  
■ F-Helix

- Loss of protection upon binding with calcium, reporting the overall structural transition
- Linker region between EF-hands / more exposed helices



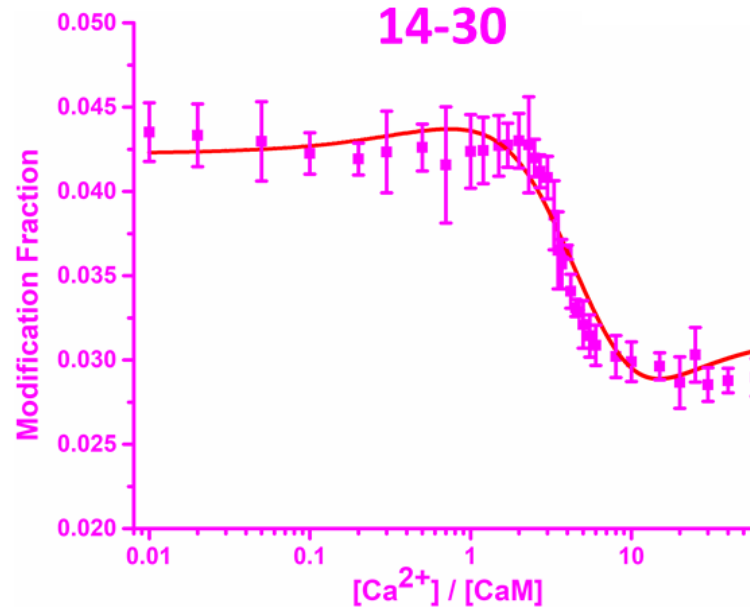
# Class II Behavior – Constant Protection



- Constant protection upon binding with calcium
- Protein N-terminus / F-helix of EF-1



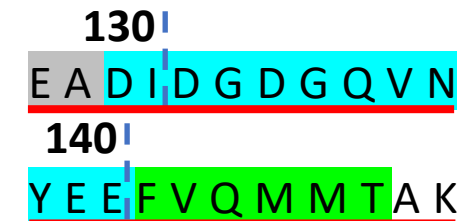
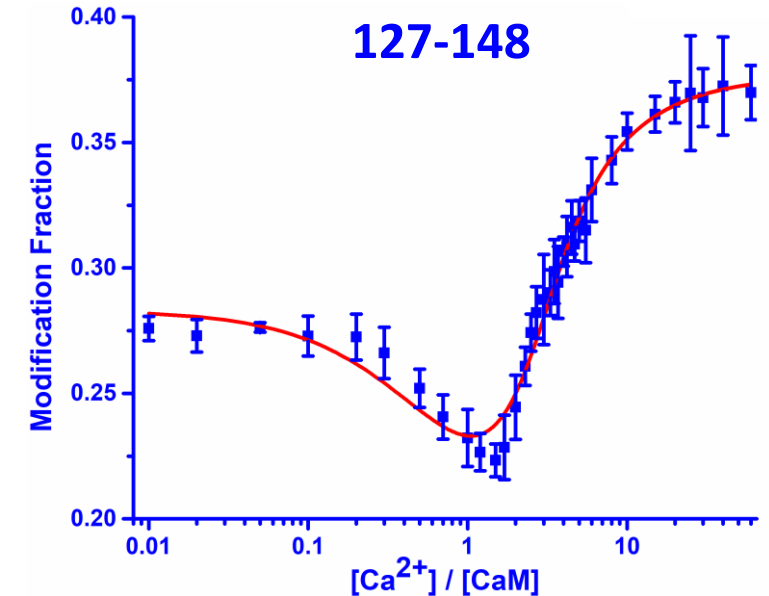
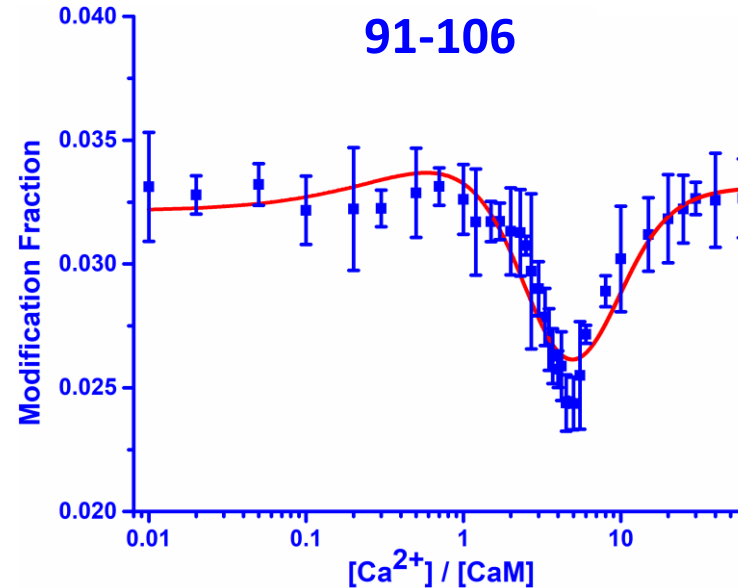
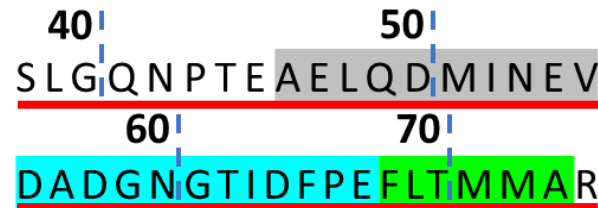
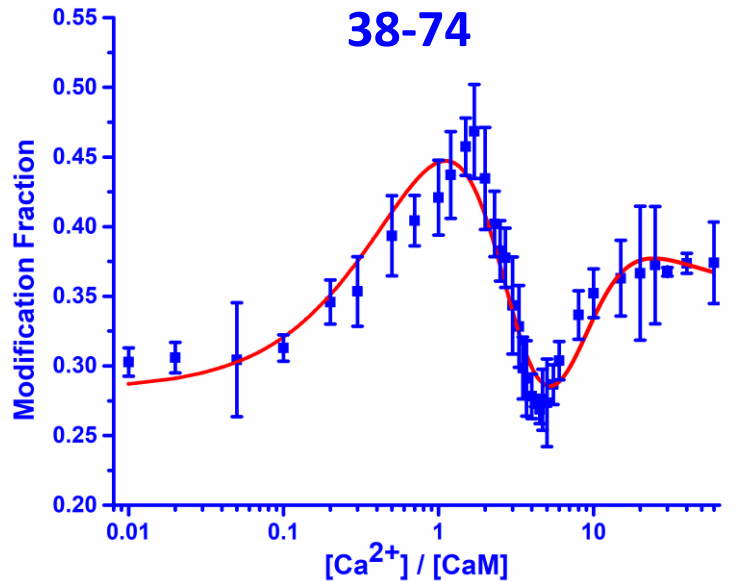
# Class III Behavior – Classical Binding



- E-helix & calcium loop of EF-1
  - More protected upon binding with calcium, a classical binding behavior
- 20 30
- E A F S L F D K D G D G T I T T K
- E-Helix
- Ca<sup>2+</sup> Loop



# Class IV Behavior – Composite Behavior



E-Helix

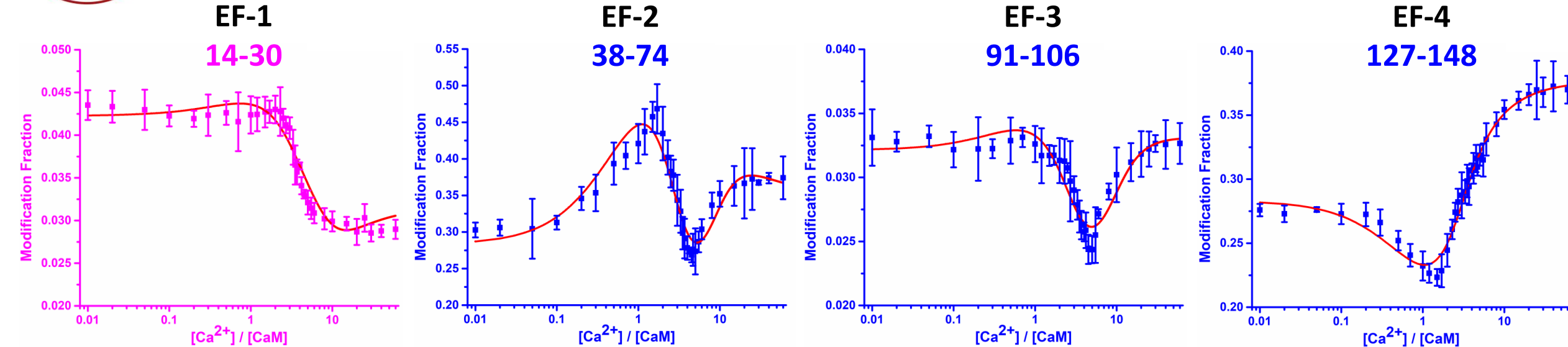
$Ca^{2+}$  Loop

F-Helix

- Covers EF-2 (38-74), EF-3 (91-106) and EF-4 (127-148)
- Binding & Allostery (Conf. change by a remote binding event)



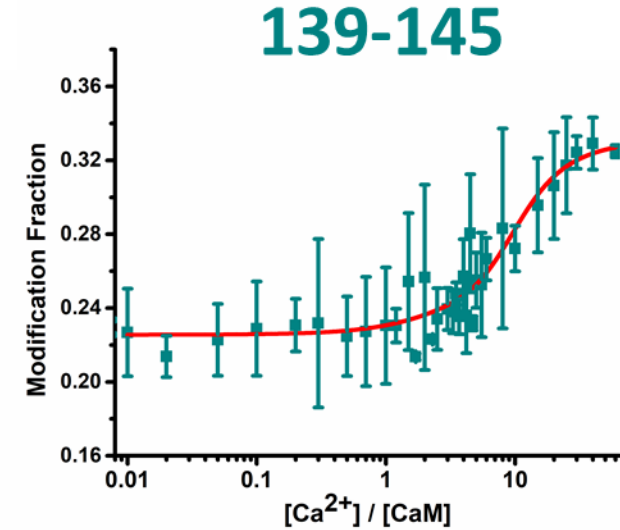
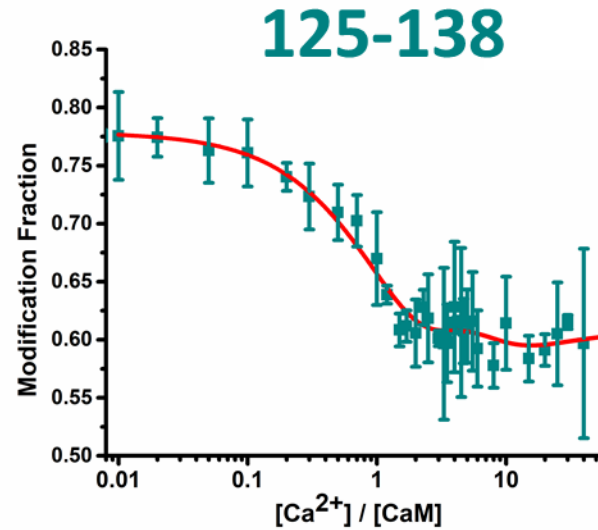
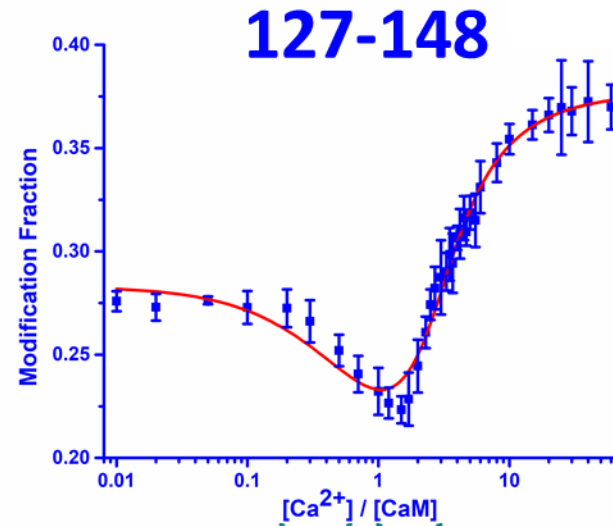
# Allosteric Behavior / Binding Dynamics



- EF-4 (127-148) binds first, followed by a conformational change over helix F (more exposed)
- EF-2 (38-74) becomes more exposed owing to binding at EF-3 (91-106) and EF-4 (127-148), preparing EF-2 for  $Ca^{2+}$  binding
- EF-2, 3 and 4 become more exposed at high  $Ca^{2+}$  owing to binding at EF-1 (14-30)
- Calmodulin is saturated at  $[Ca^{2+}] / [CaM] = 15$



# Dissect Composite LITPOMS by Chymotrypsin



E-Helix

Ca<sup>2+</sup> Loop

F-Helix



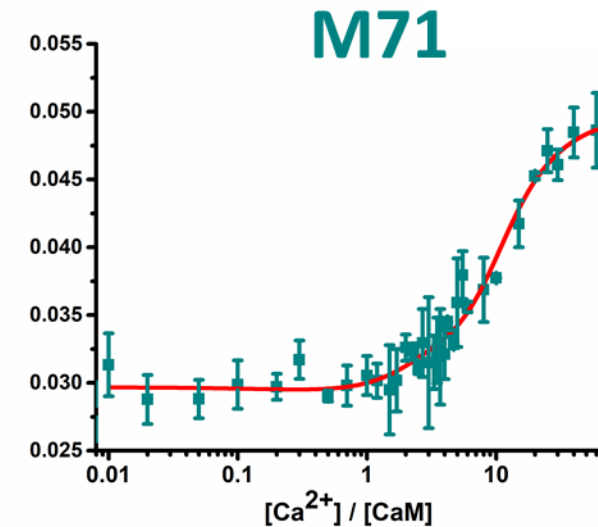
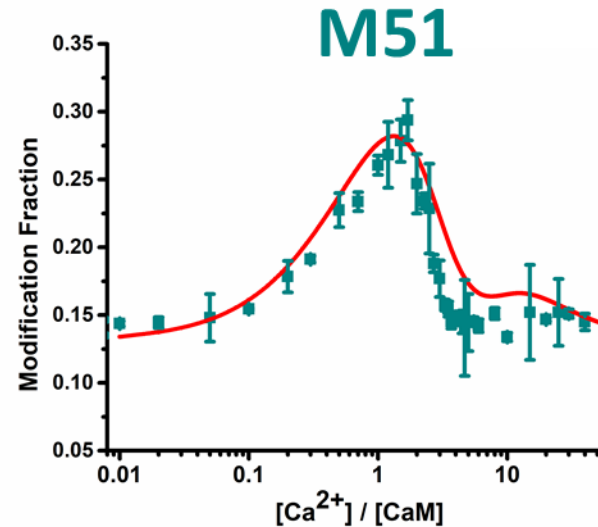
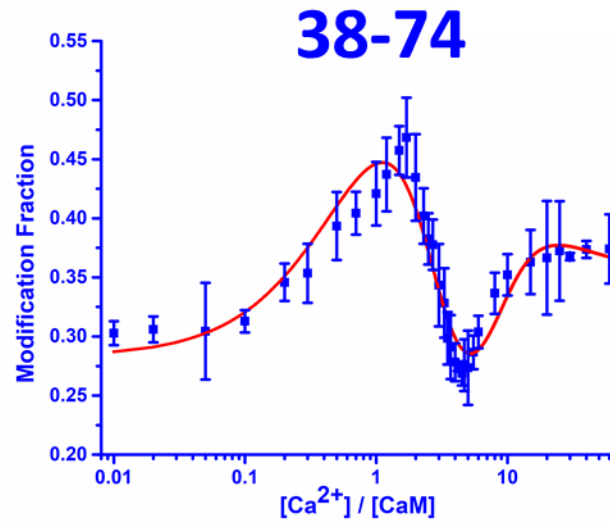
- Tryptic Peptide 127-148 covers EF-4 of calmodulin
- Chymotryptic peptides 125-138 and 139-145 show simple behavior of binding and opening

[1] [Liu, X. R.](#); Rempel, D. L.; Gross, M. L. *Anal. Chem.* **2019**, *91*, 12560-12567.

[2] [Liu, X. R.](#); Zhang, M. M.; Rempel, D. L.; Gross, M. L. *Anal. Chem.* **2019**, *91*, 5508-5512.



# Dissect Composite LITPOMS by Residue-Level Analysis



E-Helix

Ca<sup>2+</sup> Loop

F-Helix



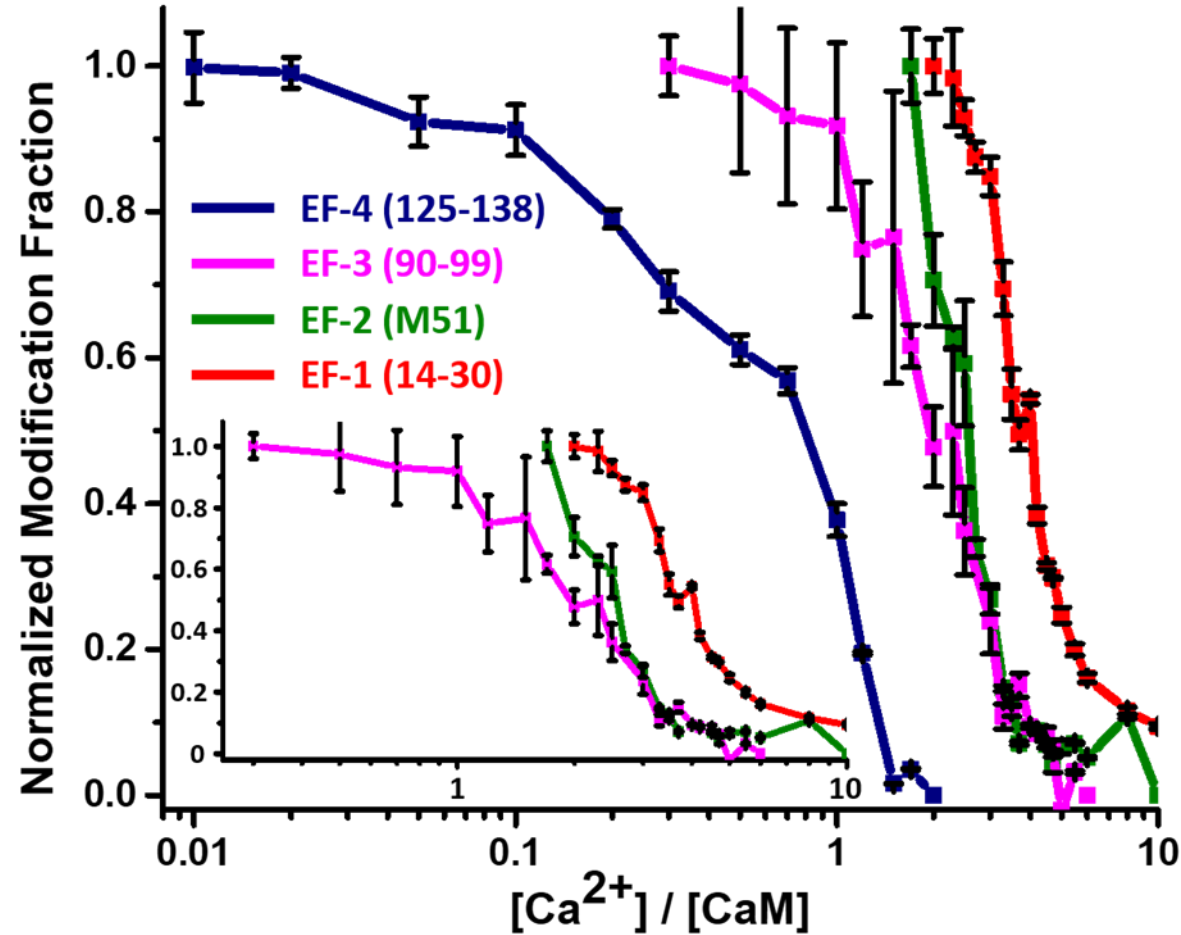
- Tryptic Peptide 38-74 covers EF-2 of calmodulin
- Residue M51 show deprotection initially, followed by binding induced protection and stay protected

[1] **Liu, X. R.**; Rempel, D. L.; Gross, M. L. *Anal. Chem.* **2019**, 91, 12560-12567.

[2] **Liu, X. R.**; Zhang, M. M.; Rempel, D. L.; Gross, M. L. *Anal. Chem.* **2019**, 91, 5508-5512.



# Ca<sup>2+</sup> Binding Order by LITPOMS



**EF-4 > EF-3 > EF-2 > EF-1**



# Site-Specific Binding Affinity

- Four distinct binding affinities through fitting

| Binding Order | EF-hand        | LITPOMS $K_i$ ( $M^{-1}$ ) | Literature $K_i$ ( $M^{-1}$ ) |
|---------------|----------------|----------------------------|-------------------------------|
| 1             | EF-4 (127-148) | $1.4 \times 10^6$          | $8.0 \times 10^4$             |
| 2             | EF-3 (91-106)  | $6.2 \times 10^6$          | $4.0 \times 10^6$             |
|               | C-Term Lobe    | $8.6 \times 10^{12}$       | $3.2 \times 10^{11}$          |
| 3             | EF-2 (38-74)   | $4.1 \times 10^4$          | $2.5 \times 10^4$             |
| 4             | EF-1 (14-30)   | $2.9 \times 10^6$          | $4.0 \times 10^5$             |
|               | N-Term Lobe    | $1.2 \times 10^{11}$       | $1.0 \times 10^{10}$          |

- Agree with literature values within 20-fold

[1] **Liu, X. R.**; Zhang, M. M.; Rempel, D. L.; Gross, M. L. *Anal. Chem.* **2019**, 91, 5508-5512.

[2] Weis, D. D. *Hydrogen Exchange Mass Spectrometry of Proteins, Fundamentals, Methods, and Applications*; Wiley: Chichester, 2016.

[3] Linse, S.; Helmersson, A.; Forsen, S. *J. Biol. Chem.* **1991**, 266, 8050-8054.



# Conclusion

- LITPOMS for the first time reveals the calcium binding order and allostery of calmodulin upon binding with calcium
- March through the protein region-by-region, characterize binding at peptide & residue level
- Composite LITPOMS behavior can be dissected by a different protease or by analyzing residue-level LITPOMS curves
- Successfully demonstrate the capability of dealing with
  - Systems with various binding stoichiometries
  - Systems with binding affinities of  $\mu\text{M}$  to nM (both loose and tight binders)
  - Systems that bind with small peptides
  - Systems that bind with metal ions
- Readily applicable to other systems with complex binding schemes, e.g., signaling proteins

[1] **Liu, X. R.**; Zhang, M. M.; Rempel, D. L.; Gross, M. L. *J. Am. Soc. Mass Spectrom.* **2019**, *30*, 213-217.

[2] **Liu, X. R.**; Zhang, M. M.; Rempel, D. L.; Gross, M. L. *Anal. Chem.* **2019**, *91*, 5508-5512.

[3] **Liu, X. R.**; Rempel, D. L.; Gross, M. L. *Anal. Chem.* **2019**, *91*, 12560-12567.



# Acknowledgement



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**R24GM136766**

**R01GM131008**



**PROTEIN METRICS**



**Stay Safe!**

