

ENHANCING THE SPEED OF VACCINE DEVELOPMENT BY UTILIZING FULLY AUTOMATED HIGH THROUGHPUT CELL POTENCY ASSAYS



MERCK

INVENTING FOR LIFE

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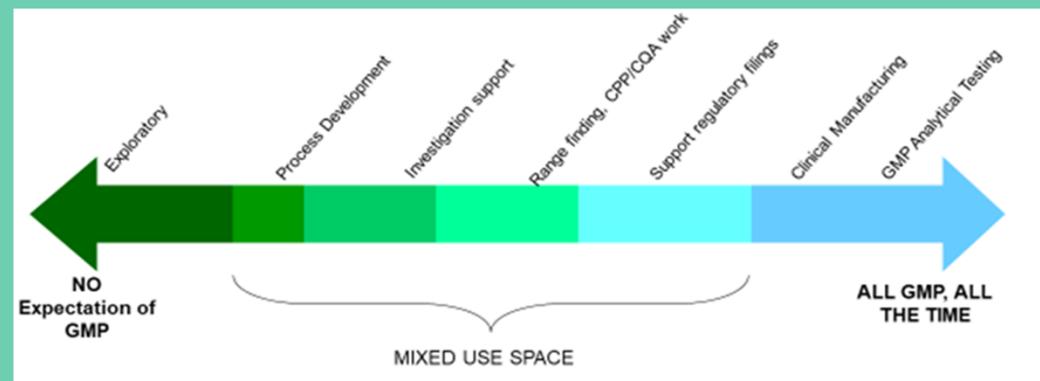
High-throughput Automation

High-throughput Automation is a popular solution to accelerating process development by increasing sample numbers and data quality while decreasing data turnaround time, and assay variability. A wide range of analytical techniques and specialized equipment can be used for high throughput automation.

Who are our partners:

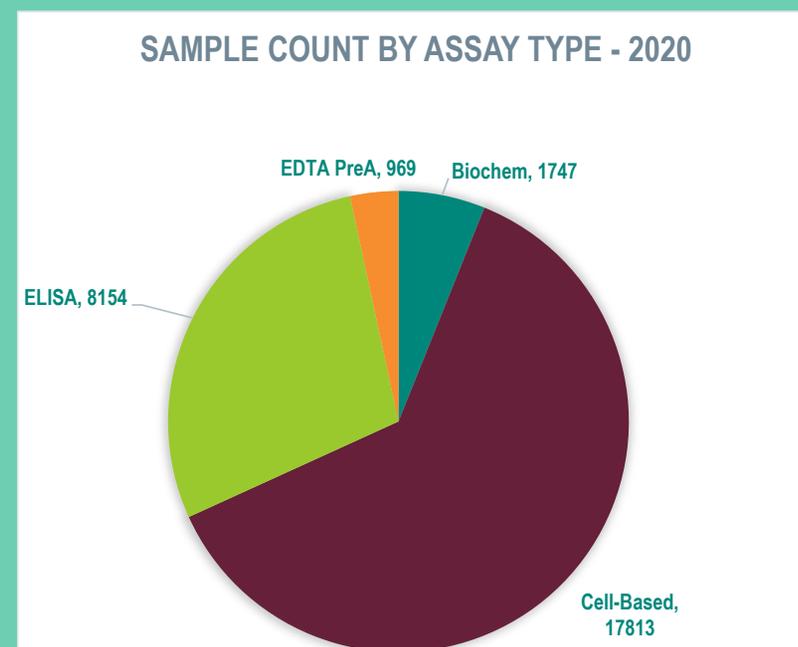
- Process development teams
- Formulation
- Continually evaluating partnerships

Phases we support:



Automation Strategy

- Focus
 - Labor intensive assays (ELISAs, Cell Based Assays...)
 - High sample volume assays
 - In use for long periods of time
- “Walk-away” automation
 - Maximize resource liberation
 - 1 day ELISA assays
- Automation of data reduction and reporting
- 10 assays
- 8 vaccine candidates

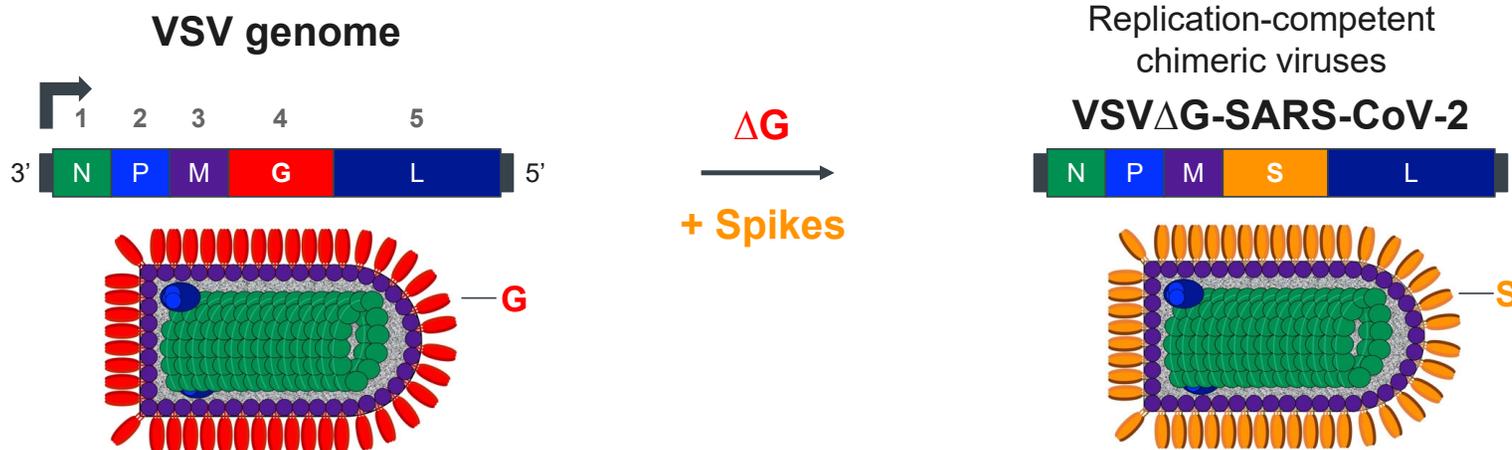


CASE STUDY

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Background on Covid Vaccine candidates



Utilize a recombinant Vesicular Stomatitis Virus (VSV) platform by replacing the VSV-G protein with the SARS-CoV-2 S Protein

The CoV Spike gene is large - VSV genome size is increased by ~ 3kb

The VSV and CoV replication cycles differ

- Replication occurs in different cellular compartments
- CoV Spikes concentrate in different regions of the cell compared to VSV G
- CoV Spikes tend to be fusogenic



Assay Concept / Comparison

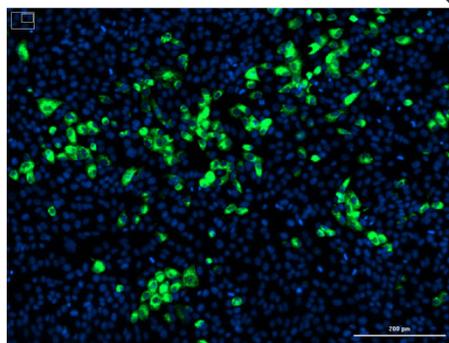
- Plaque Assays – to quantify infectious viral units that are capable of infection and cell-to-cell transmission (i.e. viral replication)
 - Goal is to allow multiple infection rounds per infectious virion to form plaque colonies
 - Generally direct quantitation assay (pfu/mL read out)
 - Typically used as a dose assay for viral vaccines

- Infectivity Assays – to quantify viral units capable of infecting a cell and expressing viral protein(s) on the surface of a cell
 - Goal is to measure only primary infection events per viral unit
 - Generally a relative potency read-out (%RP); can also be direct quantitation (FFU/mL or IU/mL)
 - Typically used as a measure of potency for process and formulation development; can also be used as dose assay

HTA Cell Based Assays Platforms



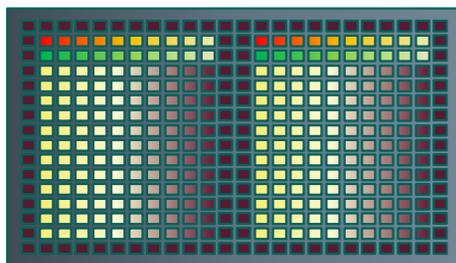
HighRes Biosolutions System "Selene"



V590 Infectivity using BioTek Cytation5

HT Infectivity Assay Platform

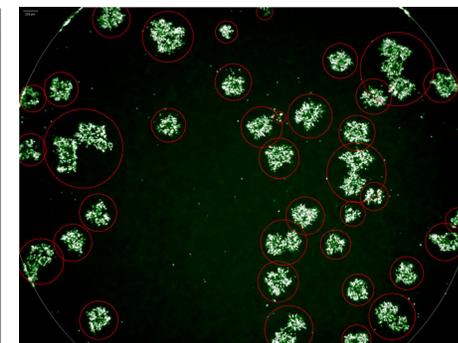
- Fully automated (48-60) 384 well plate run capacity – 12 samples/plate (in duplicate)
- 576 - 720 samples per run
- 2-3 runs/week
- 96-capacity built-in
- BioTek Cytation5



Our first HighRes Biosolutions System "Phoenix" →
Set up for both Assay Platforms in lower capacity and throughput



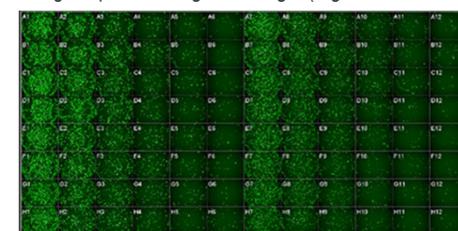
PAA System "Helios"



V590 Immuno-plaque
Image captured using PE EnSight (Algorithm Counted: 44)

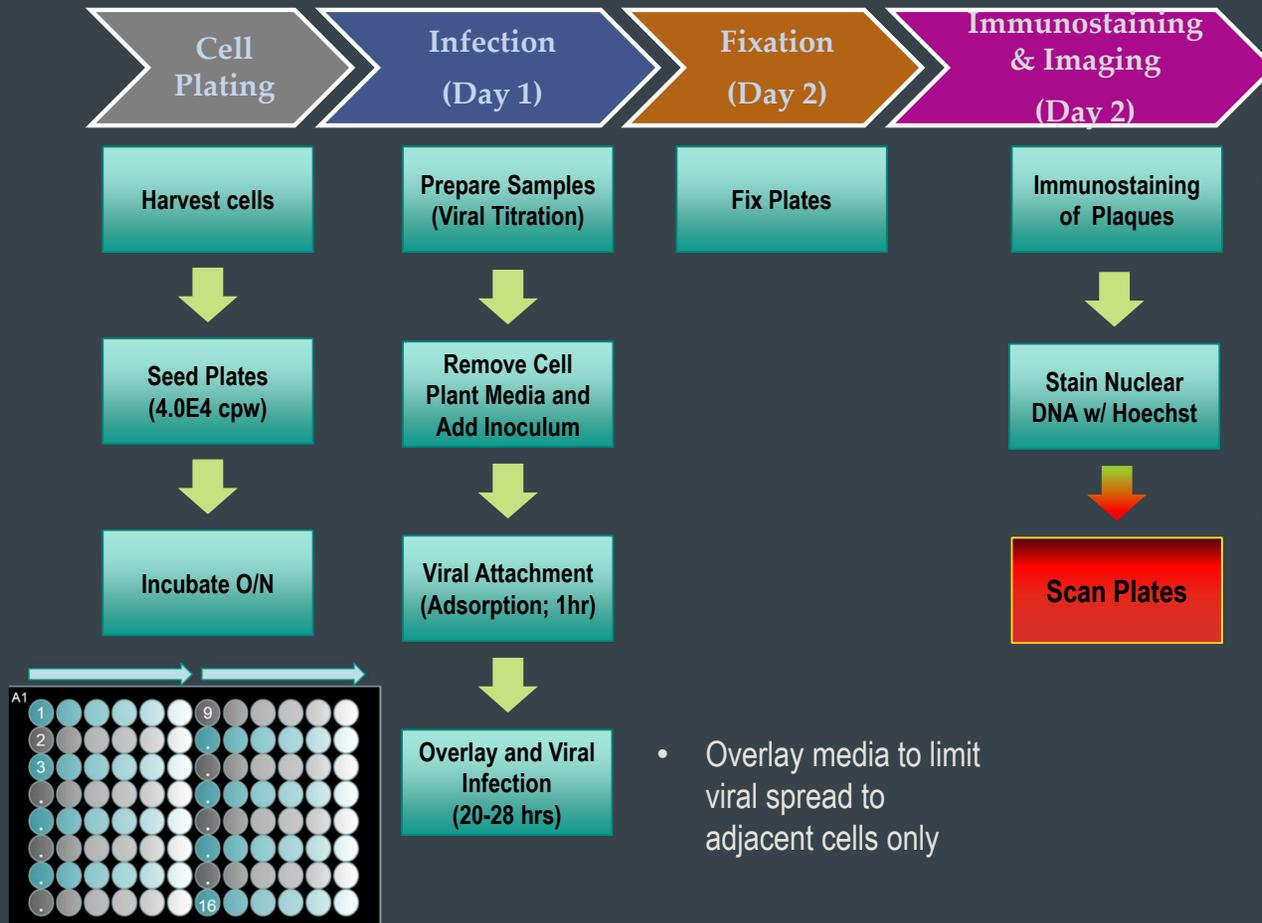
HT μ Plaque Assay Platform

- Fully automated (44-48) 96 well plate run capacity – 14 samples/plate (in singleton)
- ~600-700 samples per run
- 1-2 runs/week
- 384 well capability built-in
- PE EnSight & BioTek Cytation3





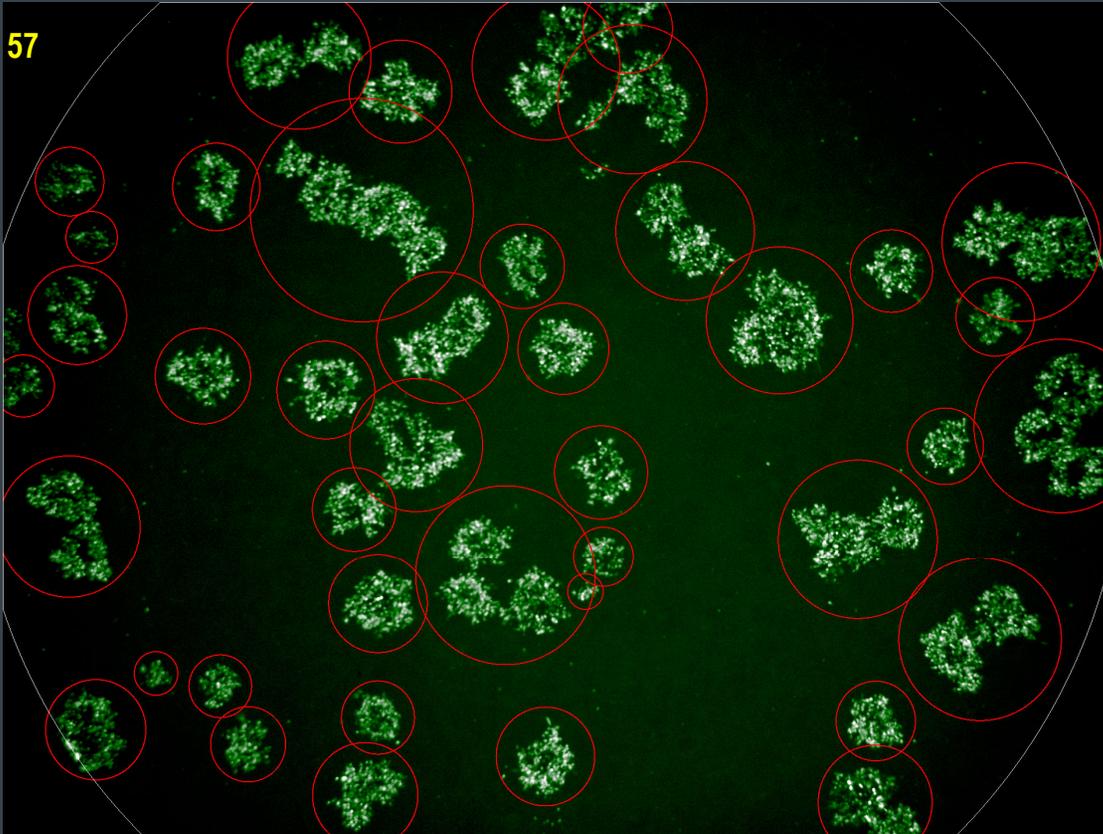
96-well Plaque Assay Workflow



- Immunostaining of plaques with viral specific *mAb*
 - AlexaFluor secondary
- Stain nuclei with Hoechst
- Read plates on PE Enight
 - Brightfield (% Confl.)
 - GFP (Plaques)
 - Nuclear Count



Plaque Counting Algorithm on PE EnSight / Kaleido Software



1. Count single, continuous fluorescent objects
2. Depending on sizing parameters, partition objects, by area alone, into estimated plaques
 - Doesn't leverage gaussian or watershed algorithms; these are computationally intensive

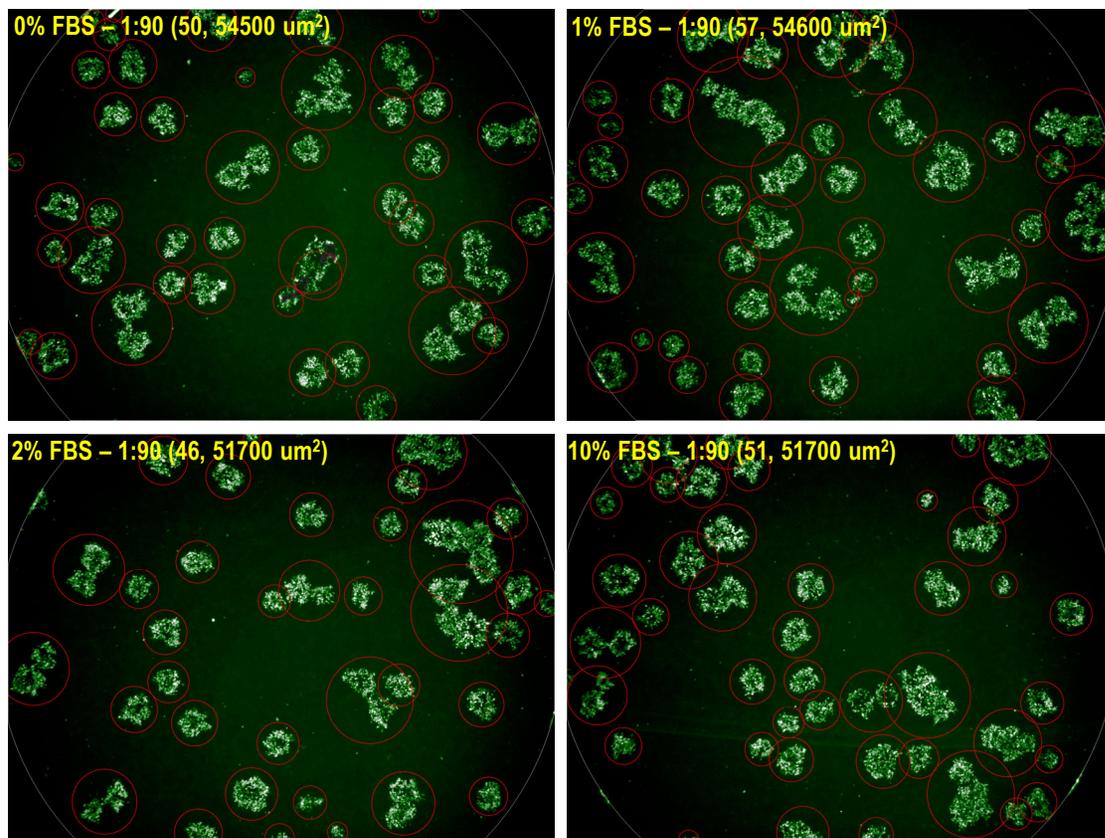
Experience with this for about 4-5 years → Algorithm has evolved over time and is well defined

Multi-Factored DOE for μ Plaque

Assay Variables Adjustments

1. Cell substrate, plate seeding density, and passaging concentrations
2. FBS concentration in assay media and overlay media
3. Trypsin concentration in assay media and overlay media; Impact of Trypsin on potency
4. Pen/Strep in assay media and overlay media
5. Primary and secondary antibody candidates for immuno-staining
6. Basal media used for cell growth, planting, and assay dilutions
7. Viral adsorption and infection kinetics
8. Sample stability through freeze thaw cycles, time on deck (diluted) and time at stock conc.
9. Sample dilutions for various stages of process and formulation development
10. Imaging parameters including plaque size, brightness, and overall plaque quality

Serum Content – Viral Attachment (10% FBS in Overlay)



	PFU/mL	%CV
0% FBS during Infection	2.01E+05	6%
1% FBS during Infection	2.83E+05	12%
2% FBS during Infection	2.46E+05	10%
10% FBS during Infection	1.61E+05	24%

Constant Conditions:

Final Bulk Drug Substance

10% FBS in Overlay Media

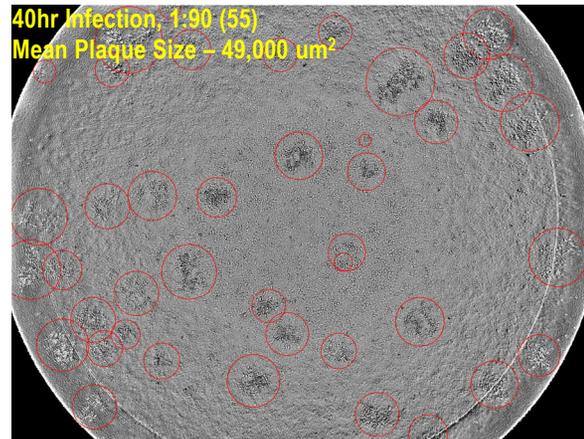
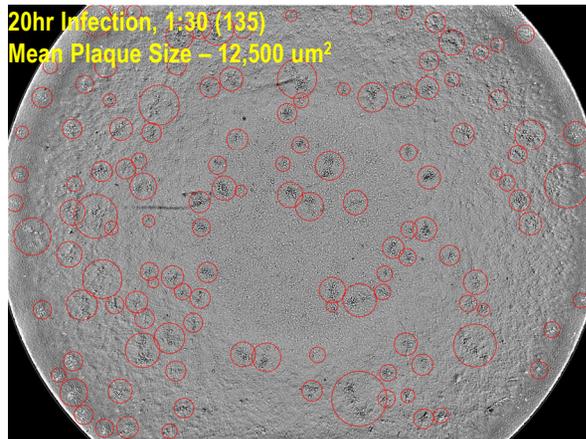
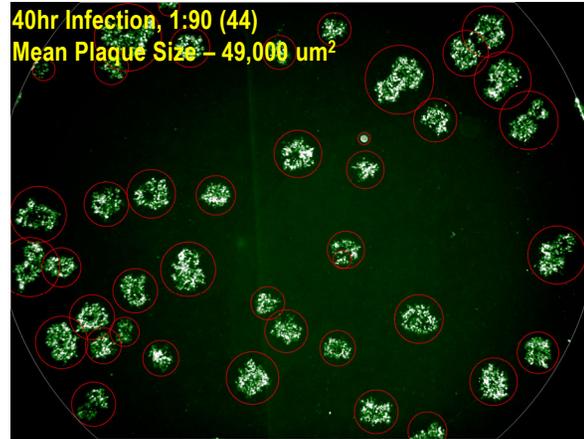
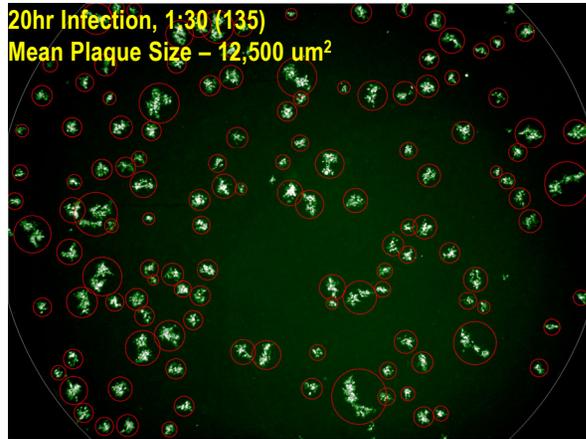
Conclusions:

Serum is critical for cell health but can be inhibitory on viral uptake

Not a significant difference in titers between serum content;
decrease 10% FBS

- Cell health improves with increasing FBS conc.
- 1% and 2% FBS optimal for sensitivity and cell health

20 hour vs 40 hour infection



Sample Used:

Final Bulk Drug Substance

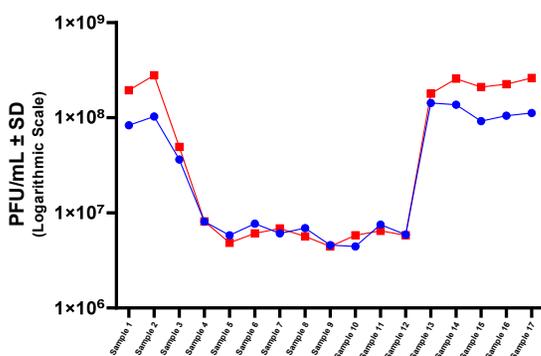
Conclusions:

20 hour incubation time has smaller but quantifiable plaques

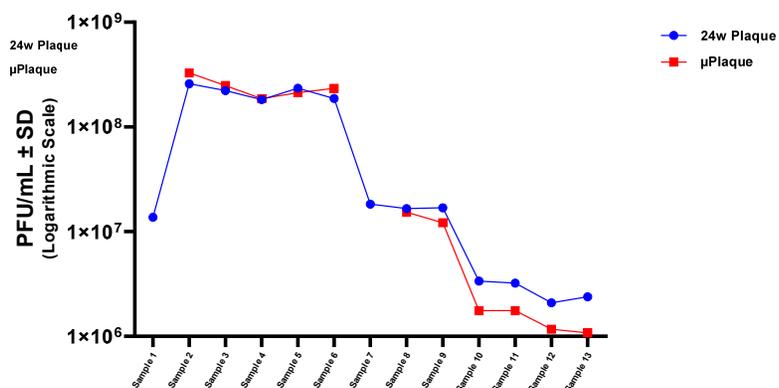
- Allows earlier detection
- Wider quantifiable range per well (i.e. more plaques per surface area)

Process Development Downstream Samples Correlation

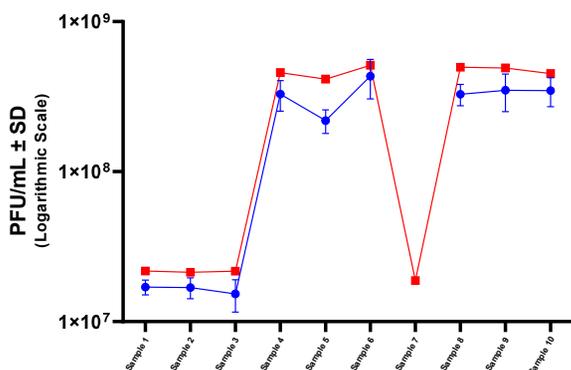
24w Plaque vs. 96w μ Plaque
Downstream Samples



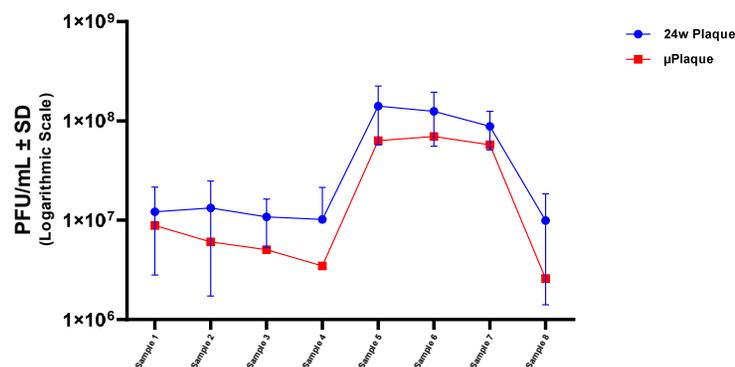
24w Plaque vs. 96w μ Plaque
Downstream Samples



24w Plaque vs. 96w μ Plaque
Downstream Samples



24w Plaque vs. 96w μ Plaque
Downstream Samples



Notes

Unit differences between assays

- 24w \rightarrow Geomean titers with %RSD variance
- 96w \rightarrow Mean titers with %CV variance

Correlation studies not controlled for cell passage, same vials, etc.

Summary

Good trending between the assays pointing towards a strong correlation

Better alignment in peak response observed between the assays

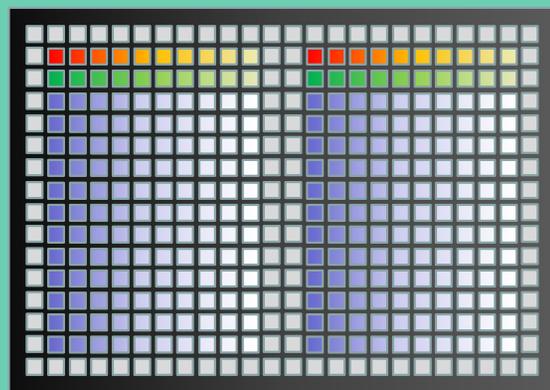
- Variance in trends likely due to cell passage \rightarrow known issue with Vero cells and has been observed with other projects as well

Infectivity Assay Steps



Serial Dilution
of Virus

Incubate at
37°C, 5%CO₂ for
8-20 Hours



- 14 rows of standard / samples *in duplicate*
 - High Throughput: 12 samples / plate
- Outer edges not infected – Assay Blanks

Permeabilize

Block &
Hoechst Stain

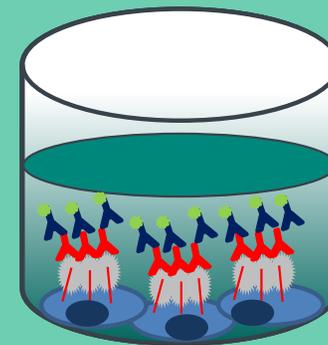
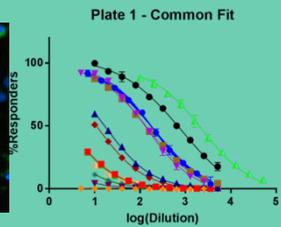
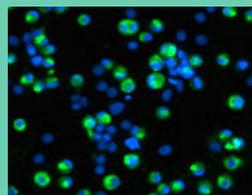
1° Ab

2° Ab
(AlexaFluor)

Add PBS and Transfer to
reader

Cytation 3

Relative
Potency



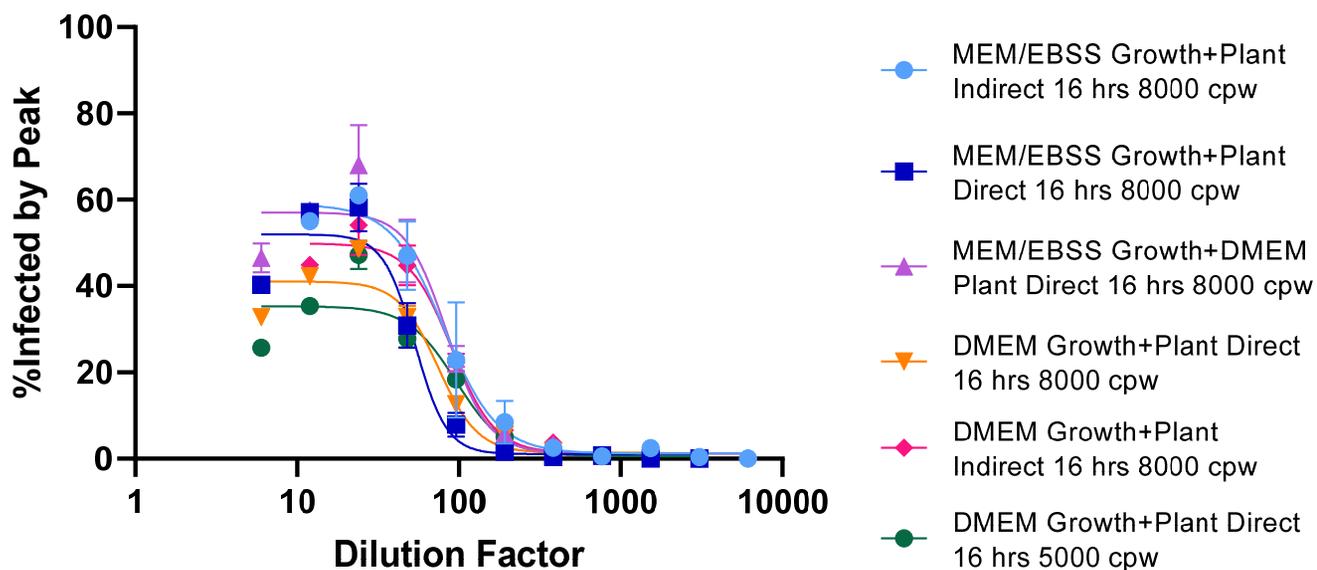
Multi-Factored DOE for Infectivity

Assay Variables Adjustments

1. Cell substrate, plate seeding density, and passaging concentrations
2. Basal media used for cell growth, planting, and assay dilutions
3. Direct vs. Indirect infection kinetics and incubation periods
4. Assay media FBS source
5. Primary and secondary antibody candidates for immuno-staining
6. Sample stability through freeze thaw cycles, time on deck (diluted) and time at stock conc.
7. Sample dilutions for various stages of process and formulation development
8. Plate washer height, cycle count, and aspirate/dispense rate
9. Mitigation of “hook effect” through cell seeding density, media refeed/refresh, and addition of ApoE and rTrypsin
10. Concordance with μ Plaque data

Infectivity Data – Hook Effect

0.20 U/mL TAME rTrypsin - Comparisons



Sample Used:
Final Bulk Drug Substance

- Conditions Varied**
- cell seeding density,
 - Media
 - media refeed/refresh,

Conclusions:
Unfortunately, Hook Effect still present

Conclusions

- The 96-well μ Plaque assay was successfully developed, optimized, and deployed for COVID-19 program support within 6 weeks
- Since then, it has provided key data for more than 7300 samples at an average turn around time of 8 days and is being run, at minimum, once a week.
- While the Infectivity assay did not successfully complete development, due to hook effects we gained additional understanding on platform support.

THANK YOU!

Acknowledgements

HTA Group

- John Loughney
- Amy Bowman
- Pete DePhillips
- Kristine Kearns
- Matt Troutman
- Carl Hofmann
- Purvit Patel
- James Devlin
- Josh Petersheim
- Nisarg Patel

Formulation

- Erica Stable
- Lynne Isopi
- Jenny Kriss
- Marie Glebocki

APL

- Chris Tubbs
- Kelsey Hines

Cell Based Science

- Melissa Whiteman
- Ashley Gruber

QUESTIONS?

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